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(54) Title: CHIMERIC PROTEINS COMPRISING BORRELIA POLYPEPTIDES: USES THEREFOR

Novel chimeric nucleic acids, encoding chimeric Borrelia proteins consisting of at least two antigenic polypeptides from corresponding and/or non-corresponding proteins from the same and/or different species of Borrelia, are disclosed. Chimeric proteins encoded by the nucleic acid sequences are also disclosed. The chimeric proteins are useful as vaccine immunogens against Lyme borreliosis, as well as for immunodiagnostic reagents.

(57) Abstract

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CHIMERIC PROTEINS COMPRISING BORRELIA POLYPEPTIDES: USES THEREFOR

Background of the Invention

Lyme borreliosis is the most common tick-borne infectious disease in North America, Europe, and northern Asia. The causative bacterial agent of this disease, Borrelia burgdorferi, was first isolated and cultivated in 1982 (Burgdorferi, W.A. et al., Science 216: 1317-1319 (1982); Steere, A.R. et al., N. Engl. J.

- Med. 308: 733-740 (1983)). With that discovery, a wide array of clinical syndromes, described in both the European and American literature since the early 20th century, could be attributed to infection by B. burgdorferi (Afzelius, A., Acta Derm. Venereol. 2: 120-
- 15 125 (1921); Bannwarth, A., <u>Arch. Psychiatr.</u>

 <u>Nervenkrankh. 117</u>: 161-185 (1944); Garin, C. and A.

 Bujadouz, <u>J. Med. Lyon 71</u>: 765-767 (1922); Herxheimer,

 K. and K. Hartmann, <u>Arch. Dermatol. Syphilol. 61</u>: 57-76,

 255-300 (1902)).
- The immune response to B. burgdorferi is characterized by an early, prominent, and persistent humoral response to the end of lagellar protein, p41 (fla), and to a protein constituent of the protoplasmic cylinder, p93 (Szczepanski, A., and J.L. Benach,
- Microbiol. Rev. 55:21 (1991)). The p41 flagellin antigen is an immunodominant protein; however, it shares significant homology with flagellins of other microorganisms and therefore is highly cross reactive. The p93 antigen is the largest immunodominant antigen of
- 30 B. burgdorferi. Both the p41 and p93 proteins are physically cryptic antigens, sheathed from the immune system by an outer membrane whose major protein constituents are the outer surface proteins A and B

(OspA and OspB). OspA is a basic lipoprotein of approximately 31 kd, which is encoded on a large linear plasmid along with OspB, a basic lipoprotein of approximately 34 kd (Szczepanski, A., and J.L. Benach, 5 Microbiol. Rev. 55:21 (1991)). Analysis of isolates of B. burgdorferi obtained from North America and Europe has demonstrated that OspA has antigenic variability, and that several distinct groups can be serologically and genotypically defined (Wilske, B., et al., World J. 10 Microbiol. 7: 130 (1991)). Other Borrelia proteins demonstrate similar antigenic variability. Surprisingly, the immune response to these outer surface proteins tends to occur late in the disease, if at all (Craft, J. E. et al., <u>J. Clin Invest. 78</u>: 934-939 (1986); Dattwyler, R.J. and B.J. Luft, Rheum. Clin. 15 North Am. 15: 727-734 (1989)). Furthermore, patients acutely and chronically infected with B. burgdorferi respond variably to the different antigens, including OspA, OspB, OspC, OspD, p39, p41 and p93.

Vaccines against Lyme borreliosis have been attempted. Mice immunized with a recombinant form of OspA are protected from challenge with the same strain of B. burgdorferi from which the protein was obtained (Fikrig, E., et al., Science 250: 553-556 (1990)).

Furthermore, passively transferred anti-OspA monoclonal antibodies (Mabs) have been shown to be protective in mice, and vaccination with a recombinant protein induced protective immunity against subsequent infection with the homologous strain of B.burgdorferi (Simon, M.M., et al., J. Infect. Dis. 164: 123 (1991)). Unfortunately, immunization with a protein from one strain does not necessarily confer resistance to a heterologous strain (Fikrig, E. et al., J. Immunol. 7: 2256-1160 (1992)), but rather, is limited to the homologous 'species' from

35 which the protein was prepared. Furthermore,

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immunization with a single protein from a particular strain of Borrelia will not confer resistance to that strain in all individuals. There is considerable variation displayed in OspA and OspB, as well as p93, including the regions conferring antigenicity. Therefore, the degree and frequency of protection from vaccination with a protein from a single strain depend upon the response of the immune system to the particular variation, as well as the frequency of genetic variation in B. burgdorferi. Currently, a need exists for a vaccine which provides immunogenicity across species and to more epitopes within a species, as well as immunogenicity against more than one protein.

Summary of the Invention

The current invention pertains to chimeric Borrelia 15 proteins which include two or more antigenic Borrelia polypeptides which do not occur naturally (in nature) in the same protein in Borrelia, as well as the nucleic acids encoding such chimeric proteins. The antigenic polypeptides incorporated in the chimeric proteins are 20 derived from any Borrelia protein from any strain of Borrelia, and include outer surface protein (Osp) A, OspB, OspC, OspD, p12, p39, p41, p66, and p93. proteins from which the antigenic polypeptides are derived can be from the same strain of Borrelia, from 25 different strains, or from combinations of proteins from the same and from different strains. If the proteins from which the antigenic polypeptides are derived are OspA or OspB, the antigenic polypeptides can be derived from either the portion of the OspA or OspB protein present between the amino terminus and the conserved tryptophan of the protein (referred to as a proximal portion), or the portion of the OspA or OspB protein present between the conserved tryptophan of the protein

and the carboxy terminus (referred to as a distal portion). Particular chimeric proteins, and the nucleotide sequences encoding them, are set forth in Figures 23-37 and 43-46.

The chimeric proteins of the current invention provide antigenic polypeptides of a variety of Borrelia strains and/or proteins within a single protein. Such proteins are particularly useful in immunodiagostic assays to detect the presence of antibodies to native Borrelia in potentially infected individuals as well as to measure T-cell reactivity, and can therefore be used as immunodiagnostic reagents. The chimeric proteins of the current invention are additionally useful as vaccine immunogens against Borrelia infection.

For a better understanding of the present invention together with other and further objects, reference is made to the following description, taken together with the accompanying drawings.

Brief Description of the Drawings

Figure 1 summarizes peptides and antigenic domains localized by proteolytic and chemical fragmentation of OspA.

Figure 2 is a comparison of the antigenic domains depicted in Figure 1, for OspA in nine strains of B. burgdorferi.

Figure 3 is a graph depicting a plot of weighted polymorphism versus amino acid position among 14 OspA variants. The marked peaks are: a) amino acids 132-145; b) amino acids 163-177; c) amino acids 208-221. The lower dotted line at polymorphism value 1.395 demarcates statistically significant excesses of polymorphism at p = 0.05. The upper dotted line at 1.520 is the same, except that the first 29 amino acids at the monomorphic N-terminus have been removed from the original analysis.

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Figure 4 depicts the amino acid alignment of residues 200 through 220 for OspAs from strains B31 and K48 as well as for the site-directed mutants 613, 625, 640, 613/625, and 613/640. Arrow indicates Trp216. Amino acid changes are underlined.

Figure 5 is a helical wheel projection of residues 204-217 of B31 OspA. Capital letters indicate hydrophobic residues; lower case letters indicate hydrophilic residues; +/- indicate positively/negatively charged residues. Dashed line indicates division of the alpha-helix into hydrophobic arc (above the line) and polar arc (below the line). Adapted from France et al. (Biochem. Biophys. Acta 1120: 59 (1992)).

Figure 6 depicts a phylogenic tree for strains of

Borrelia described in Table I. The strains are as
follows: 1 = B31; 2 = Pka1; 3 = ZS7; 4 = N40; 5 =

25015; 6 = K48; 7 = DK29; 8 = PHei; 9 = Ip90; 10 =

PTrob; 11 = ACAI; 12 = PGau; 13 = Ip3; 14 = PBo; 15 =

PKo.

Figure 7 depicts the nucleic acid sequence of OspA-B31 (SEW ID NO. 6), and the encoded protein sequence (SEQ ID NO. 7).

Figure 8 depicts the nucleic acid sequence of OspA-K48 (SEQ ID NO. 8), and the encoded protein sequence (SEQ ID NO. 9).

Figure 9 depicts the nucleic acid sequence of OspA-PGau (SEQ ID NO. 10), and the encoded protein sequence (SEQ ID NO. 11).

Figure 10 depicts the nucleic acid sequence of OspA-25015 (SEQ ID NO. 12), and the encoded protein sequence (SEQ ID NO. 13).

Figure 11 depicts the nucleic acid sequence of OspB-B31 (SEQ ID NO. 21), and the encoded protein sequence (SEQ ID NO. 22).

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Figure 12 depicts the nucleic acid sequence of OspC-B31 (SEQ ID NO. 29), and the encoded protein sequence (SEQ ID NO. 30).

Figure 13 depicts the nucleic acid sequence of OspC-K48 (SEQ ID NO. 31), and the encoded protein sequence (SEQ ID NO. 32).

Figure 14 depicts the nucleic acid sequence of OspC-PKo (SEQ ID NO. 33), and the encoded protein sequence (SEQ ID NO. 34).

10 Figure 15 depicts the nucleic acid sequence of OspC-pTrob (SEQ ID NO. 35) and the encoded protein sequence (SEQ ID NO. 36).

Figure 16 depicts the nucleic acid sequence of p93-B31 (SEQ ID NO. 65) and the encoded protein sequence (SEQ ID NO. 66).

Figure 17 depicts the nucleic acid sequence of p93-K48 (SEQ ID NO. 67).

Figure 18 depicts the nucleic acid sequence of p93-PBo (SEQ ID NO. 69).

20 Figure 19 depicts the nucleic acid sequence of p93pTrob (SEQ ID NO. 71).

Figure 20 depicts the nucleic acid sequence of p93-pGau (SEQ ID NO. 73).

Figure 21 depicts the nucleic acid sequence of p93-25 25015 (SEQ ID NO. 75).

Figure 22 depicts the nucleic acid sequence of p93-pKo (SEQ ID NO. 77).

Figure 23 depicts the nucleic acid sequence of the OspA-K48/OspA-PGau chimer (SEQ ID NO. 85) and the encoded chimeric protein sequence (SEQ ID NO. 86).

Figure 24 depicts the nucleic acid sequence of the OspA-B31/OspA-PGau chimer (SEQ ID NO. 88) and the encoded chimeric protein sequence (SEQ ID NO. 89).

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Figure 25 depicts the nucleic acid sequence of the OspA-B31/OspA-K48 chimer (SEQ ID NO. 91) and the encoded chimeric protein sequence (SEQ ID NO. 92).

Figure 26 depicts the nucleic acid sequence of the OspA-B31/OspA-25015 chimer (SEQ ID NO. 94) and the encoded chimeric protein sequence (SEQ ID NO. 95).

Figure 27 depicts the nucleic acid sequence of the OspA-K48/OspA-B31/OspA-K48 chimer (SEQ ID NO. 97) and the encoded chimeric protein sequence (SEQ ID NO. 98).

Figure 28 depicts the nucleic acid sequence of the OspA-B31/OspA-K48/OspA-B31/OspA-K48 chimer (SEQ ID NO. 100) and the encoded chimeric protein sequence (SEQ ID NO. 101).

Figure 29 depicts the nucleic acid sequence of the OspA-B31/OspB-B31 chimer (SEQ ID NO. 103) and the encoded chimeric protein sequence (SEQ ID NO. 104).

Figure 30 depicts the nucleic acid sequence of the OspA-B31/OspB-B31/OspC-B31 chimer (SEQ ID NO. 106) and the encoded chimeric protein sequence (SEQ ID NO. 107).

Figure 31 depicts the nucleic acid sequence of the OspC-B31/OspA-B31/OspB-B31 chimer (SEQ ID NO. 109) and the encoded chimeric protein sequence (SEQ ID NO. 110).

Figure 32 depicts the nucleic acid sequence of the OspA-B31/p93-B31 chimer (SEQ ID NO. 111) and the encoded chimeric protein sequence (SEQ ID NO. 112).

Figure 33 depicts the nucleic acid sequence of the OspB-B31/p41-B31 (122-234) chimer (SEQ ID NO. 113) and the encoded chimeric protein sequence (SEQ ID NO. 114).

Figure 34 depicts the nucleic acid sequence of the OspB-B31/p41-B31 (122-295) chimer (SEQ ID NO. 115) and the encoded chimeric protein sequence (SEQ ID NO. 116).

Figure 35 depicts the nucleic acid sequence of the OspB-B31/p41-B31 (140-234) chimer (SEQ ID NO. 117) and the encoded chimeric protein sequence (SEQ ID NO. 118).

Figure 36 depicts the nucleic acid sequence of the OspB-B31/p41-B31 (140-295) chimer (SEQ ID NO. 119) and the encoded chimeric protein sequence (SEQ ID NO. 120).

Figure 37 depicts the nucleic acid sequence of the 0spB-B31/p41-B31 (122-234)/OspC-B31 chimer (SEQ ID NO. 121) and the encoded chimeric protein sequence (SEQ ID NO. 122).

Figure 38 depicts an alignment of the nucleic acid sequences for OspC-B31 (SEQ ID NO. 29), OspC-PKO (SEQ ID NO. 33), OspC-pTrob (SEQ ID NO. 35), and OspC-K48 (SEQ ID NO. 31). Nucleic acids which are identical to those in the lead nucleic acid sequence (here, OspC-B31) are represented by a period (.); differing nucleic acids are shown in lower case letters.

Figure 39 depicts an alignment of the nucleic acid sequences for OspD-pBO (SEQ ID NO. 123), OspD-PGau (SEQ ID NO. 124), OspD-DK29 (SEQ ID NO. 125), and OspD-K48 (SEQ ID NO. 126). Nucleic acids which are identical to those in the lead nucleic acid sequence (here, OspD-pBo) are represented by a period (.); differing nucleic acids are shown in lower case letters.

Figure 40 depicts the nucleic acid sequence of p41-B31 (SEq ID NO. 127) and then encoded protein sequence (SEQ ID NO. 128).

Figure 41 depicts an alignment of the nucleic acid sequences for p41-B31 (SEQ ID NO. 127), p41-pKa1 (SEQ ID NO. 129), p41-PGau (SEQ ID NO. 51), p41-PBo (SEQ ID NO. 130), p41-DK29 (SEQ ID NO. 53), and p41-PKo (SEQ ID NO. 131). Nucleic acids which are identical to those in the lead nucleic acid sequence (here, p41-B31) are represented by a period (.); differing nucleic acids are shown in lower case letters.

Figure 42 depicts an alignment of the nucleic acid sequences for OspA-B31 (SEQ ID NO. 6), OspA-pKa1 (SEQ ID NO. 132), OspA-N40 (SEQ ID NO. 133), OspA-ZS7 (SEQ ID

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NO. 134), OspA-25015 (SEQ ID NO. 12), OspA-pTrob (SEQ ID NO. 135), OspA-K48 (SEQ ID NO. 8), OspA-Hei (SEQ ID NO. 136), OspA-DK29 (SEQ ID NO. 49), OSpA-Ip90 (SEQ ID NO. 50), OspA-pBo (Seq ID NO. 55), OspA-Ip3 (SEQ ID NO. 56), OspA-PKo (SEQ ID NO. 57), OspA-ACAI (SEQ ID NO. 58), and OspA-PGau (SEQ ID NO. 10). Nucleic acids which are identical to those in the lead nucleic acid sequence (here, OspA-B31) are represented by a period (.); differing nucleic acids are shown in lower case letters.

Figure 43 depicts the nucleic acid sequence of the OspA-Tro/OspA-Bo chimer (SEQ ID NO. 137) and the encoded chimeric protein sequence (SEQ ID NO. 138).

Figure 44 depicts the nucleic acid sequence of the OspA-PGau/OspA-Bo chimer (SEQ ID NO. 139) and the encoded chimeric protein sequence (SEQ ID NO. 140).

Figure 45 depicts the nucleic acid sequence of the OspA-B31/OspA-PGau/OspA-B31/OspA-K48 chimer (SEQ ID NO. 141) and the encoded chimeric protein sequence (SEQ ID NO. 142).

Figure 46 depicts the nucleic acid sequence of the OspA-PGau/OspA-B31/OspA-K48 chimer (SEQ ID NO. 143) and the encoded chimeric protein sequence (SEQ ID NO. 144).

Detailed Description of the Invention

The current invention pertains to chimeric proteins comprising antigenic Borrelia polypeptides which do not occur in nature in the same Borrelia protein. The chimeric proteins are a combination of two or more antigenic polypeptides derived from Borrelia proteins. The antigenic polypeptides can be derived from different proteins from the same species of Borrelia, or different proteins from different Borrelia species, as well as from corresponding proteins from different species. As used herein, the term "chimeric protein" describes a protein comprising two or more polypeptides which are

derived from corresponding and/or non-corresponding native Borrelia protein. A polypeptide "derived from" a native Borrelia protein is a polypeptide which has an amino acid sequence the same as an amino acid sequence 5 present in a Borrelia protein, an amino acid sequence equivalent to the amino acid sequence of a naturally occurring Borrelia protein, or an amino acid sequence substantially similar to the amino acid sequence of a naturally occurring Borrelia protein (e.g., differing by 10 few amino acids) such as when a nucleic acid encoding a protein is subjected to site-directed mutagenesis. "Corresponding" proteins are equivalent proteins from different species or strains of Borrelia, such as outer surface protein A (OspA) from strain B31 and OspA from 15 strain K48. The invention additionally pertains to nucleic acids encoding these chimeric proteins.

As described below, Applicants have identified two separate antigenic domains of OspA and OspB which flank the sole conserved tryptophan present in OspA and in 20 OspB. These domains share cross-reactivity with different genospecies of Borrelia. The precise amino acids responsible for antigenic variability were determined through site-directed mutagenesis, so that proteins with specific amino acid substitutions are available for the development of chimeric proteins. 25 Furthermore, Applicants have identified immunologically important hypervariable domains in OspA proteins, as described below in Example 2. The first hypervariable domain of interest for chimeric proteins, Domain A, includes amino acid residues 120-140 of OspA, the second hypervariable domain, Domain B, includes residues 150-180 and the third hypervariable domain, Domain C, includes residues 200-216 or 217 (depending on the position of the sole conserved tryptophan residue in the OspA of that particular species of Borrelia) (see Figure

3). In addition, Applicants have sequenced the genes for several Borrelia proteins.

These discoveries have aided in the development of novel recombinant Borrelia proteins which include two or more amino acid regions or sequences which do not occur in the same Borrelia protein in nature. The recombinant proteins comprise polypeptides from a variety of Borrelia proteins, including, but not limited to, OspA, OspB, OspC, OspD, p12, p39, p41, p66, and p93.

10 Antigenically relevant polypeptides from each of a number of proteins are combined into a single chimeric protein.

In one embodiment of the current invention, chimers are now available which include antigenic polypeptides flanking a tryptophan residue. The antigenic 15 polypeptides are derived from either the proximal portion from the tryptophan (the portion of the OspA or OspB protein present between the amino terminus and the conserved tryptophan of the protein), or the distal 20 portion from the tryptophan (the portion of the OspA or OspB protein present between the conserved tryptophan of the protein and the carboxy terminus) in OspA and/or OspB. The resultant chimers can be OspA-OspA chimers (i.e., chimers incorporating polypeptides derived from OspA from different strains of Borrelia), OspA-OspB chimers, or OspB-OspB chimers, and are constructed such that amino acid residues amino-proximal to an invariant tryptophan are from one protein and residues carboxyproximal to the invariant tryptophan are from the other 30 protein. For example, one available chimer consists of a polypeptide derived from the amino-proximal region of OspA from strain B31, followed by the tryptophan residue, followed by a polypeptide derived from the carboxy-proximal region of OspA from strain K48 (SEQ ID NO. 92). Another available chimer includes a 35

polypeptide derived from the amino-proximal region of OspA from strain B31, and a polypeptide derived from the carboxy-proximal region of OspB from strain B31 (SEQ ID NO. 104). If the polypeptide proximal to the tryptophan of these chimeric proteins is derived from OspA, the proximal polypeptide can be further subdivided into the three hypervariable domains (Domains A, B, and C), each of which can be derived from OspA from a different strain of Borrelia. These chimeric proteins can further comprise antigenic polypeptides from another protein, in addition to the antigenic polypeptides flanking the tryptophan residue.

In another embodiment of the current invention, chimeric proteins are available which incorporate antigenic domains of two or more *Borrelia* proteins, such as Osp proteins (Osp A, B, C and/or D) as well as p12, p39, p41, p66, and/or p93.

The chimers described herein can be produced so that they are highly soluble, hyper-produced in E. coli, and non-lipidated. In addition, the chimeric proteins 20 can be designed to end in an affinity tag (His-tag) to facilitate purification. The recombinant proteins described herein have been constructed to maintain high levels of antigenicity. In addition, recombinant proteins specific for the various genospecies of Borrelia that cause Lyme disease are now available, because the genes from each of the major genospecies have been sequenced; the sequences are set forth below. These recombinant proteins with their novel biophysical and antigenic properties will be important diagnostic 30 reagent and vaccine candidates.

The chimeric proteins of the current invention are advantageous in that they retain specific reactivity to monoclonal and polyclonal antibodies against wild-type Borrelia proteins, are immunogenic, and inhibit the

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growth or induce lysis of Borrelia in vitro.

Furthermore, in some embodiments, the proteins provide antigenic domains of two or more Borrelia strains and/or proteins within a single protein. Such proteins are particularly useful in immuno-diagostic assays. For example, proteins of the present invention can be used as reagents in assays to detect the presence of antibodies to native Borrelia in potentially infected individuals. These proteins can also be used as immunodiagnostic reagents, such as in dot blots, Western blots, enzyme linked immunosorbed assays, or agglutination assays. The chimeric proteins of the present invention can be produced by known techniques, such as by recombinant methodology, polymerase chain reaction, or mutagenesis.

Furthermore, the proteins of the current invention are useful as vaccine immunogens against Borrelia infection. Because Borrelia has been shown to be clonal, a protein comprising antigenic polypeptides from a variety of Borrelia proteins and/or species, will provide immunoprotection for a considerable time when used in a vaccine. The lack of significant intragenic recombination, a process which might rapidly generate novel epitopes with changed antigenic properties, 25 ensures that Borrelia can only change antigenic type by accumulating mutational change, which is slow when compared with recombination in generating different antigenic types. The chimeric protein can be combined with a physiologically acceptable carrier and 30 administered to a vertebrate animal through standard methods (e.g., intravenously or intramuscularly, for example).

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The current invention is illustrated by the following Examples, which are not to be construed to be limiting in any way.

Example 1. Purification of Borrelia burgorferi Outer Surface Protein A and Analysis of Antibody Binding Domains

This example details a method for the purification of large amounts of native outer surface protein A (OspA) to homogeneity, and describes mapping of the antigenic specificities of several anti-OspA MAbs. OspA was purified to homogeneity by exploiting its resistance to trypsin digestion. Intrinsic labeling with ¹⁴C-palmitic acid confirmed that OspA was lipidated, and partial digestion established lipidation at the aminoterminal cysteine of the molecule.

The reactivity of seven anti-OspA murine monoclonal antibodies to nine different Borrelia isolates was ascertained by Western blot analysis. Purified OspA was fragmented by enzymatic or chemical cleavage, and the monoclonal antibodies were able to define four distinct immunogenic domains (see Figure 1). Domain 3, which included residues 190-220 of OspA, was reactive with protective antibodies known to agglutinate the organism in vitro, and included distinct specificities, some of which were not restricted to a genotype of B. burgdorferi.

A. Purification of Native OspA

Detergent solubilization of B. burgdorferi strips the outer surface proteins and yields partially-purified preparations containing both OspA and outer surface 5 protein B (Osp B) (Barbour, A.G. et al., Infect. Immun. <u>52 (5)</u>: 549-554 (1986); Coleman, J.L. and J.L. Benach, \underline{J} Infect. Dis. 155 (4): 756-765 (1987); Cunningham, T.M. et al., Ann. NY Acad. Sci. 539: 376-378 (1988); Brandt, M.E. et al., <u>Infect. Immun. 58</u>: 983-991 (1990); Sambri, 10 V. and R. Cevenini, Microbiol. 14:307-314 (1991)). Although both OspA and OspB are sensitive to proteinase K digestion, in contrast to OspB, OspA is resistant to cleavage by trypsin (Dunn, J. et al., Prot. Exp. Purif. 1: 159-168 (1990); Barbour, A.G. et al., Infect. Immun. 15 45:94-100 (1984)). The relative insensitivity to trypsin is surprising in view of the fact that Osp A has a high (16% for B31) lysine content, and may relate to the relative configuration of Osp A and B in the outer membrane.

Intrinsic Radiolabeling of Borrelia Labeling for lipoproteins was performed as described by Brandt et al. (<u>Infect. Immun. 58</u>:983-991 (1990)). ¹⁴C-palmitic acid (ICN, Irvine, California) was added to the BSK II media to a final concentration of 0.5 μCi per milliliter (ml). Organisms were cultured at 34°C in this medium until a density of 10⁸ cells per ml was achieved.

Purification of OspA Protein from Borrelia Strain B31

Borrelia burgdorferi, either ¹⁴C-palmitic acidlabeled or unlabeled, were harvested and washed as
described (Brandt, M.E. et al., <u>Infect. Immun. 58</u>:983991 (1990)). Whole organisms were trypsinized according

to the protocol of Barbour et al. (Infect. Immun. 45:94-100 (1984)) with some modifications. The pellet was suspended in phosphate buffered saline (PBS, 10mM, pH 7.2), containing 0.8% tosyl-L-phenylalanine chloromethyl 5 ketone (TPCK)-treated trypsin (Sigma, St. Louis, Missouri), the latter at a ratio of 1 μ g per 10 8 cells. Reaction was carried out at 25°C for 1 hour, following which the cells were centrifuged. The pellet was washed in PBS with 100 $\mu \text{g/ml}$ phenylmethylsulfonyl fluoride (PMSF). Triton X-114 partitioning of the pellet was 10 carried out as described by Brandt et al. (Infect. Immun. 58:983-991 (1990)). Following trypsin treatment, cells were resuspended in ice-cold 2% (v/v) Triton X-114 in PBS at 10° cells per ml. The suspension was rotated 15 overnight at 4°C, and the insoluble fraction removed as a pellet after centrifugation at 10,000 X g for 15 minutes at 4°C. The supernatant (soluble fraction) was incubated at 37°C for 15 minutes and centrifuged at room temperature at 1000 X g for 15 minutes to separate the aqueous and detergent phases. The aqueous phase was 20 decanted, and ice cold PBS added to the lower Triton phase, mixed, warmed to 37°C, and again centrifuged at 1000 X g for 15 minutes. Washing was repeated twice more. Finally, detergent was removed from the preparation using a spin column of Bio-beads SM2 25 (BioRad, Melville, New York) as described (Holloway, P.W., Anal. Biochem. 53:304-308 (1973)).

Ion exchange chromatography was carried out as described by Dunn et al. (Prot. Exp. Purif. 1: 159-168 (1990)) with minor modifications. Crude OspA was dissolved in buffer A (1% Triton X-100, 10mM phosphate buffer (pH 5.0)) and loaded onto a SP Sepharose resin (Pharmacia, Piscataway, New Jersey), pre-equilibrated with buffer A at 25°C. After washing the column with 10

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bed-volumes of buffer A, the bound OspA was eluted with buffer B (1% Triton X-100, 10mM phosphate buffer (pH 8.0)). OspA fractions were detected by protein assay using the BCA method (Pierce, Rockford, Illinois), or as radioactivity when intrinsically labeled material was fractionated. Triton X-100 was removed using a spin column of Bio-beads SM2.

This method purifies OspA from an outer surface membrane preparation. In the absence of trypsin
treatment, OspA and B were the major components of the soluble fraction obtained after Triton partitioning of strain B31. In contrast, when Triton extraction was carried out after trypsin-treatment, the OspB band is not seen. Further purification of OspA-B31 on a SP

Sepharose column resulted in a single band by SDS-PAGE. The yield following removal of detergent was approximately 2 mg per liter of culture. This method of purification of OspA, as described herein for strain B31, can be used for other isolates of Borrelia as well.

For strains such as strain K48, which lack OspB, trypsin treatment can be omitted.

Lipidation site of OspA-B31

14C-palmitic acid labeled OspA from strain B31 was purified as described above and partially digested with endoproteinase Asp-N (data not shown). Following digestion, a new band of lower molecular weight was apparent by SDS-PAGE, found by direct amino-terminal sequencing to begin at Asp₂₅. This band had no trace of radioactivity by autoradiography (data not shown). OspA and B contain a signal sequence (L-X-Y-C) similar to the consensus described for lipoproteins of E. coli, and it has been predicted that the lipidation site of OspA and B should be the amino-terminal cysteine (Brandt, M.E. et

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al., <u>Infect. Immun 58</u>: 983-991 (1990)). The results presented herein support this prediction.

Comparison of OspA Antibody Binding Regions in Nine В. Strains of Borrelia burgdorferi

The availability of the amino acid sequenced for OspA from a number of different isolates, combined with peptide mapping and Western blot analysis, permitted the identification of the antigenic domains recognized by monoclonal antibodies (MAbs) and allowed inference of 10 the key amino acid residues responsible for specific antibody reactivity.

Strains of Borrelia burgdorferi

Nine strains of Borrelia, including seven European strains and two North American strains, were used in this study of antibody binding domains of several 15 proteins. Information concerning the strains is summarized in Table I, below.

Table I. Representative Borrelia Strains

	Table 1. Representation	
Strain	Location and Source	Reference for Strain
K48	Czechoslovakia, Ixodes ricinus	none
PGau	Germany, human ACA	Wilske, B. et al., <u>J. Clin.</u> <u>Microbiol. 32</u> :340-350 (1993)
DK29	Denmark, human EM	Wilske, B. et al.
PKo	Germany, human EM	Wilske, B. et al.
PTrob	Germany, human skin	Wilske, B. et al.
Ip3	Khabarovsk, Russia, I. persulcatus	Asbrink, E. et al., <u>Acta</u> <u>Derm. Venereol. 64</u> : 506-512 (1984)
Ip90	Khabarovsk, Russia, I. persulcatus	Asbrink, E. et al.
25015	Millbrook, NY, I. persulcatus	Barbour, A.G. et al., <u>Curr.</u> <u>Microbiol. 8</u> :123-126 (1983)
B31	Shelter Island, NY, I. scapularis	Luft, B.J. et al., <u>Infect.</u> <u>Immun. 60</u> : 4309-4321 (1992); ATCC 35210
PKa1	Germany, human CSF	Wilske, B. et al.
ZS7	Freiburg, Germany, I. ricinus	Wallich, R. et al., <u>Nucl.</u> <u>Acids Res. 17</u> : 8864 (1989)
N40	Westchester Co., NY	Fikrig, E. et al., <u>Science</u> 250:553-556 (1990)
PHei	Germany, human CSF	Wilske, B. et al.
ACAI	Sweden, human ACA	Luft, B. J. et al., <u>FEMS</u> <u>Microbiol. Lett. 93</u> :73-68 (1992)
PBo	Germany, human CSF	Wilske, B. et al.

ACA = patient with acrodermatitis chronica atrophicans; EM = patient with erythema migrans; CSF = cerebrospinal fluid of patient with Lyme disease

Strains K48, PGau and DK29 were supplied by R. Johnson, University of Minnesota; PKo and pTrob were provided by B. Wilske and V. Preac-Mursic of the

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Pettenkhofer Institute, Munich, Germany; and Ip3 and Ip90 were supplied by L. Mayer of the Center for Disease Control, Atlanta, Georgia. The North American strains included strain 25015, provided by J. Anderson of the Connecticut Department of Agriculture; and strain B31 (ATCC 35210).

Monoclonal Antibodies

Seven monoclonal antibodies (MAbs) were utilized in this study. Five of the MAbs (12, 13, 15, 83 and 336) were produced from hybridomas cloned and subcloned as previously described (Schubach, W.H., et al., Infect. Immun. 59(6):1911-1915 (1991)). MAb H5332 (Barbour, A.G. et al., Infect. Immun. 41:795-804 (1983)) was a gift from Drs. Alan Barbour, University of Texas, and MAb CIII.78 (Sears, J.E. et al., J. Immunol. 147(6):1995-2000 (1991)) was a gift from Richard A. Flavell, Yale University. MAbs 12 and 15 were raised against whole sonicated B3; MAb 336 was produced against whole PGau; and MAbs 13 and 83 were raised to a truncated form of OspA cloned from the K48 strain and expressed in E. coli using the T7 RNA polymerase system 20 (McGrath, B.C. et al., Vaccines, Cold Spring Harbor Laboratory Press, Plainview, New York, pp. 365-370 (1993)). All MAbs were typed as being Immunoglobulin G (IgG).

Methods of Protein Cleavage, Western Blotting, and Amino-Terminal Sequencing

Prediction of the various cleavage sites was achieved by knowledge of the primary amino acid sequence derived from the full nucleotide sequences of OspA, many of which are currently available (see Table II, below). Cleavage sites can also be predicted based on the peptide sequence of OspA, which can be determined by standard techniques after isolation and purification of OspA by the method described above. Cleavage of several OspA isolates was

conducted to determine the localization of monoclonal antibody binding of the proteins.

Hydroxylamine-HCl (HA), N-chlorosuccinimide (NCS), and cyanogen bromide cleavage of OspA followed the methods

5 described by Bornstein (Biochem. 9 (12):2408-2421 (1970)), Shechter et al., (Biochem. 15 (23):5071-5075 (1976)), and Gross (in Hirs, C.H.W. (ed): Methods in Enzymology, (N.Y. Acad. Press), 11:238-255 (1967)) respectively. Protease cleavage by endoproteinase, Asp-N (Boehringer Mannheim,

10 Indianapolis, Indiana), was performed as described by Cleveland D.W. et al., (J. Biol. Chem. 252:1102-1106 (1977)). Ten micrograms of OspA were used for each reaction. The ratio of enzyme to OspA was approximately 1 to 10 (w/w).

Proteins and peptides generated by cleavage were separated by SDS-polyacrylamide gel electrophoresis (SDS-PAGE) (Laemmli, U.K., Nature (London) 227:680-685 (1970)), and electroblotted onto immobilon Polyvinylidine Difluoride (PVDF) membranes (Ploskal, M.G. et al., Biotechniques 20 4:272-283 (1986)). They were detected by amido black staining or by immunostaining with murine MAbs, followed by alkaline phosphatase-conjugated goat antimouse IgG. Specific binding was detected using a 5-bromo-4-chloro-3-indolylphosphate (BCIP)/nitroblue tetrazolium (NBT) developer system (KPL Inc., Gathersburg, Maryland).

In addition, amino-terminal amino acid sequence analysis was carried out on several cleavage products, as described by Luft et al. (<u>Infect. Immun. 57</u>:3637-3645 (1989)). Amido black stained bands were excised from PVDF blots and sequenced by Edman degradation using a Biosystems model 475A sequenator with model 120A PTH analyzer and model 900A control/data analyzer.

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Cleavage Products of Outer Surface Protein A Isolates Purified OspA-B31, labeled with 14C-palmitic acid, was fragmented with hydroxylamine-HCl (HA) into two peptides, designated HA1 and HA2 (data not shown). The HA1 band 5 migrated at 27 KD and retained its radioactivity, indicating that the peptide included the lipidation site at the N-terminus of the molecule (data not shown). From the predicted cleavage point, HA1 should correspond to residues 1 to 251 of OspA-B31. HA2 had a MW of 21.6 KD by SDS-PAGE, 10 with amino-terminal sequence analysis showing it to begin at Gly72, i.e. residues 72 to 273 of OspA-B31. By contrast, HA cleaved OspA-K48 into three peptides, designated HA1, HA2, and HA3 with apparent MWs of 22KD, 16 KD and 12 KD, respectively. Amino-terminal sequencing showed HA1 to start at Gly72, and HA3 at Gly142. HA2 was 15 found to have a blocked amino-terminus, as was observed for the full-length OspA protein. HA1, 2 and 3 of OspA-K48 were predicted to be residues 72-274, 1 to 141 and 142 to 274, respectively.

N-Chlorosuccinimide (NCS) cleaves tryptophan (W), which is at residue 216 of OspA-B31 or residue 217 of OspA-K48 (data not shown). NCS cleaved OspA-B31 into 2 fragments, NCS1, with MW of 23 KD, residues 1-216 of the protein, and NCS2 with a MW of 6.2 KD, residues 217 to 273 (data not shown). Similarly, K48 OspA was divided into 2 pieces, NCS1 residues 1-217, and NCS2 residues 218 to 274 (data not shown).

Cleavage of OspA by cyanogen bromide (CNBr) occurs at the carboxy side of methionine, residue 39. The major fragment, CNBr1, has a MW of 25.7 KD, residues 39-274 by amino-terminal amino acid sequence analysis (data not shown). CNBr2 (about 4 KD) could not be visualized by amido black staining; instead, lightly stained bands of about 20 KD MW were seen. These bands reacted with anti-

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OspA MAbs, and most likely were degradation products due to cleavage by formic acid.

Determination of Antibody Binding Domains for Anti-OspA Monoclonal Antibodies

The cleavage products of OspA-B31 and OspA-K48 were analyzed by Western blot to assess their ability to bind to the six different MAbs. Preliminary Western blot analysis of the cleavage products demonstrated that strains K48 and DK29 have similar patterns of reactivity, as do IP3, PGau and PKo. The OspA of strain PTrob was immunologically distinct from the others, being recognized only by MAb 336. MAb 12 recognized only the two North American strains, B31 and 25015. When the isolates were separated into genogroups, it was remarkable that all the MAbs, except MAb 12, crossed over to react with multiple genogroups.

MAb12, specific for OspA-B31, bound to both HA1 and HA2 of OspA-B31. However, cleavage of OspA-B31 by NCS at residue Trp216 created fragments which did not react with MAb12, suggesting that the relevant domain is near or is structurally dependent upon the integrity of this residue (data not shown). MAb 13 bound only to OspA-K48, and to peptides containing the amino-terminus of that molecule (e.g. HA2; NCS1). It did not bind to CNBrl residues 39 to 274. Thus the domain recognized by MAb13 is in the amino-terminal end of OspA-K48, near Met38.

MAb15 reacts with the OspA of both the B31 and K48 strains, and to peptides containing the N-terminus of OspA, such as HA1 of OspA-B31 and NCS1, but not to peptides HA2 of OspA-B31 and HA1 of OspA-K48 (data not shown). Both peptides include residue 72 to the C-terminus of the molecules. MAb15 bound to CNBr1 of OspA-K48, indicating the domain for this antibody to be residues 39 to 72, specifically near Gly72 (data not shown).

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MAD83 binds to OspA-K48, and to peptides containing the C-terminal portion of the molecule, such as HA1. They do not bind to HA2 of OspA-K48, most likely because the C-terminus of HA2 of OspA-K48 ends at 141. Similar to MAD12 and OspA-B31, binding of MADS 83 and CIII.78 is eliminated by cleavage of OspA at the tryptophan residue. Thus binding of MADS 12, 83 and CIII.78 to OspA depends on the structural integrity of the Trp216 residue, which appears to be critical for antigenicity. Also apparent is that, although these MADS bind to a common antigenic domain, the precise epitopes which they recognize are distinct from one another given the varying degrees of cross-reactivity to these MADS among strains.

Although there is similar loss of binding activity of
MAD336 with cleavage at Trp216, this MAD does not bind to
HA1 of OspA-B31, suggesting the domain for this antibody
includes the carboxy-terminal end of the molecule,
inclusive of residues 251 to 273. Low MW peptides, such as
HA3 (10 KD) and NCS2 (6KD), of OspA-K48 do not bind this
MAD on Western blots. In order to confirm this
observation, we tested binding of the 6 MADs with a
recombinant fusion construct p3A/EC that contains a trpE
leader protein fused with residues 217 to 273 of OspA-B31
(Schubach, W.H. et al., Infect. Immun. 59(6): 1911-1915
(1991)). Only MAD336 reacted with this construct (data not
shown). Peptides and antigenic domains localized by
fragmentation of OspA are summarized in Figure 1.

Mapping of Domains to Define the Molecular Basis for the Serotype Analysis

To define the molecular basis for the serotype analysis of OspA, we compared the derived amino acid sequences of OspA for the nine isolates (Figure 2). At the amino terminus of the protein, these predictions can be more precise given the relatively small number of amino

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acid substitutions in this region compared to the carboxy terminus. Domain 1, which is recognized by MAb13, includes residues Leu34 to Leu41. MAb13 only binds to the OspA of species K48, DK29 and IP90. Within this region, residue 37 5 is variable, however Gly37 is conserved amongst the three reactive strains. When Gly37 is changed to Glu37, as it is in OspA of strains B31, pTrob, PGau, and PKo, MAb13 does not recognize the protein (data not shown). By similar analysis, it can be seen that Asp70 is a crucial residue 10 for Domain 2, which includes residues 65 to 75 and is recognized by MAb15. Domain 3 is reactive with MAbs H5332, 12 and 83, and includes residues 190-220. It is clear that significant heterogeneity exists between MAbs reactive with this domain, and that more than one conformational epitope 15 must be contained within the sequence. Domain 4 binds MAb336, and includes residues 250 to 270. In this region, residue 266 is variable and therefore may be an important determinant. It is apparent, however, that other determinants of the reactivity of this monoclonal antibody 20 reside in the region comprising amino acids 217-250. Furthermore, the structural integrity of Trp216 is essential for antibody reactivity in the intact protein. Finally, it is important to stress that Figure 2 indicates only the locations of the domains, and does not necessarily 25 encompass the entire domain. Exact epitopes are being analyzed by site-directed mutagenesis of specific residues.

Overall, evidence suggests that the N-terminal portion is not the immunodominant domain of OspA, possibly by virtue of its lipidation, and the putative function of the lipid moiety in anchoring the protein to the outer envelope. The C-terminal end is immunodominant and includes domains that account in part for structural heterogeneity (Wilske, B. et al., Med. Microbiol. Immunol. 181: 191-207 (1992)), and may provide epitopes for antibody

neutralization (Sears, J.E. et al., J. Immunol. 147(6):
1995-2000 (1991)), and relate to other activities, such as
the induction of T-cell proliferation (Shanafel, M.M., et
al., J. Immunol. 148: 218-224 (1992)). There are common
epitopes in the carboxy-end of the protein that are shared
among genospecies which may have immunoprotective potential
(Wilske, B., et al., Med. Microbiol. Immunol. 181: 191-207
(1992)).

Prediction of secondary structure on the basis of

hydropathy analysis and circular dichroism and fluorescence spectroscopy measurements (McGrath, B.C., et al., Vaccines, Cold Spring Harbor Laboratory Press, Plainview, New York; pp. 365-370 (1993)) suggest domains 3 and 4 to be in a region of the molecule with a propensity to form alphahelix, whereas domains 1 and 2 occur in regions predicted to be beta-sheets (see Figure 1). These differences may distinguish domains in accessibility to antibody or to reactive T-cells (Shanafel, M.M. et al., J. Immunol. 148: 218-224 (1992)). Site-directed mutagenesis of specific epitopes, as described below in Example 2, aids in identifying exact epitopes.

Example 2. Identification of an Immunologically Important Hypervariable Domain of the Major Outer Surface Protein A of Borrelia

This Example describes epitope mapping studies using chemically cleaved OspA and TrpE-OspA fusion proteins. The studies indicate a hypervariable region surrounding the single conserved tryptophan residue of OspA (at residue 216, or in some cases 217), as determined by a moving window population analysis of OspA from fifteen European and North American isolates of Borrelia. The hypervariable region is important for immune recognition.

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Site-directed mutagenesis was also conducted to examine the hypervariable regions more closely. Fluorescence and circular dichroism spectroscopy have indicated that the conserved tryptophan is part of an 5 alpha-helical region in which the tryptophan is buried in a hydrophobic environment (McGrath, B.C., et al., Vaccines, Cold Spring Harbor Laboratory Press, Plainview, New York; pp. 365-370 (1993)). More polar amino acid side-chains flanking the tryptophan are likely to be exposed to the 10 hydrophilic solvent. The hypervariability of these solvent-exposed residues among the various strains of Borrelia suggested that these amino acid residues may contribute to the antigenic variation in OspA. Therefore, site-directed mutagenesis was performed to replace some of the potentially exposed amino acid side chains in the 15 protein from one strain with the analogous residues of a second strain. The altered proteins were then analyzed by Western Blot using monoclonal antibodies which bind OspA on the surface of the intact, non-mutated spirochete. 20 results indicated that certain specific amino acid changes near the tryptophan can abolish reactivity of OspA to these monoclonal antibodies.

A. <u>Verification of Clustered Polymorphisms in Outer</u> <u>Surface Protein A Sequences</u>

Cloning and sequencing of the OspA protein from fifteen European and North American isolates (described above in Table I) demonstrated that amino acid polymorphism is not randomly distributed throughout the protein; rather, polymorphism tended to be clustered in three regions of OspA. The analysis was carried out by plotting the moving, weighted average polymorphism of a window (a fixed length subsection of the total sequence) as it is slid along the sequence. The window size in this analysis was thirteen amino acids, based upon the determination of the largest

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number of significantly deviating points as established by the method of Tajima (J. Mol. Evol. 33: 470-473 (1991)). The average weighted polymorphism was calculated by summing the number of variant alleles for each site. Polymorphism calculations were weighted by the severity of amino acid replacement (Dayhoff, M.O. et al., in: Dayhoff, M.O. (ed.) Atlas of Protein Sequence and Structure NBRF, Washington, Vol. 5, Suppl. 3: 345 (1978)). The sum was normalized by the window size and plotted. The amino acid sequence position corresponds to a window that encompasses amino acids 1 through 13. Bootstrap resampling was used to generate 95% confidence intervals on the sliding window analysis. Since Borrelia has been shown to be clonal, the bootstrap analysis should give a reliable estimate of the 15 expected variance out of polymorphism calculations. bootstrap was iterated five hundred times at each position, and the mean was calculated from the sum of all positions. The clonal nature of Borrelia ensures that the stochastic variance that results from differing genealogical histories of the sequence positions (as would be expected if 20 recombination were prevalent) will be minimized.

This test verified that the three regions around the observed peaks all have significant excesses of polymorphism. Excesses of polymorphism were observed in the regions including amino acid residues 132-145, residues 163-177, and residues 208-221 (Figure 3). An amino acid alignment between residues 200 and 220 for B31, K48 and the four site-directed mutants is shown in Figure 4. The amino acid 208-221 region includes the region of OspA which has been modeled as an oriented alpha-helix in which the single tryptophan residue at amino acid 216 is buried in a hydrophobic pocket, thereby exposing more polar amino acids to the solvent (Figure 5) (France, L.L., et al., Biochem. Biophys. Acta 1120: 59 (1992)). These potentially solvent-exposed residues showed considerable variability among the

OspAs from various strains and may be an important component of OspA antigenic variation. For the purposes of generating chimeric proteins, the hypervariable domains of interest are <u>Domain A</u>, which includes amino acid residues 120-140 of OspA; <u>Domain B</u>, which includes residues 150-180; and <u>Domain C</u>, which includes residues 200-216 or 217.

B. Site-Directed Mutagenesis of the Hypervariable Region
Site-directed mutagenesis was performed to convert
residues within the 204-219 domain of the recombinant B31

OspA to the analogous residues of a European OspA variant,
K48. In the region of OspA between residues 204 and 219,
which includes the helical domain (amino acids 204-217),
there are seven amino acid differences between OspA-B31 and
OspA-K48. Three oligonucleotides were generated, each
containing nucleotide changes which would incorporate K48
amino acids at their analogous positions in the B31 OspA
protein. The oligos used to create the site-directed
mutants were:

5'-CTTAATGACTCTGACACTAGTGC-3' (#613, which converts

threonine at position 204 to serine, and serine at 206 to
threonine (Thr204-Ser, Thr206-Ser)) (SEQ ID NO. 1);

5'-GCTACTAAAAAAACCGGGAAATGGAATTCA-3' (#625, which converts
alanine at 214 to glycine, and alanine at 215 to lysine
(Ala214-Gly, Ala215-Lys)) (SEQ ID NO. 2); and

5'-GCAGCTTGGGATTCAAAAACATCCACTTTAACA-3' (#640, which
converts asparagine at 217 to aspartate, and glycine at

219 to lysine (Asn217-Asp, Gly219-Lys)) (SEQ ID NO. 3).

Site-directed mutagenesis was carried out by

performing mutagenesis with pairs of the above oligos.

Three site-directed mutants were created, each with two

changes: OspA 613 (Thr204-Ser, Thr206-Ser), OspA 625

(Ala214-Gly, Ala215-Lys), and 640 (Asn217-Asp, Gly219-Lys).

There were also two proteins with four changes: OspA

613/625 (Thr204-Ser, Thr206-Ser, Ala214-Gly, Ala215-Lys)

and OspA 613/640 (Thr204-Ser, Thr206-Ser, Asn217-Asp, Gly219-Lys).

Specificity of Antibody Binding to Epitopes of the Non-mutated Hypervariable Region

Monoclonal antibodies that agglutinate spirochetes, including several which are neutralizing in vitro, recognize epitopes that map to the hypervariable region around Trp216 (Barbour, A.G. et al., Infect. and Immun. 41: 759 (1983); Schubach, W.H. et al., Infect. and Immun. 59:

1911 (1991)). Western Blot analysis demonstrated that chemical cleavage of OspA from the B31 strain at Trp 216 abolishes reactivity of the protein with the agglutinating Mab 105, a monoclonal raised against B31 spirochetes (data not shown). The reagent, n-chlorosuccinimide (NCS),

cleaves OspA at the Trp 216, forming a 23.2kd fragment and a 6.2kd peptide which is not retained on the Imobilon-P membrane after transfer. The uncleaved material binds Mab 105; however, the 23.2kd fragment is unreactive. Similar Western blots with a TrpE-OspA fusion protein containing

the carboxy-terminal portion of the OspA protein demonstrated that the small 6.2kd piece also fails to bind Mab 105 (Schubach, W.H. et al., <u>Infect. and Immun. 59</u>: 1911 (1991)).

Monoclonal antibodies H5332 and H3TS (Barbour, A.G. et al., Infect. and Immun. 41: 759 (1983)) have been shown by immunofluorescence to decorate the surface of fixed spirochetes (Wilske, B. et al., World J. Microbiol. 7: 130 (1991)). These monoclonals also inhibit the growth of the organism in culture. Epitope mapping with fusion proteins has confirmed that the epitopes which bind these Mabs are conformationally determined and reside in the carboxy half of the protein. Mab H5332 is cross-reactive among all of the known phylogenetic groups, whereas Mab H3TS and Mab 105 seem to be specific to the B31 strain to which they were

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raised. Like Mab 105, the reactivities of H5332 and H3TS to OspA are abrogated by fragmentation of the protein at Trp216 (data not shown). Mab 336 was raised to whole spirochetes of the strain P/Gau. It cross-reacts to OspA from group 1 (the group to which B31 belongs) but not to group 2 (of which K48 is a member). Previous studies using fusion proteins and chemical cleavage have indicated that this antibody recognizes a domain of OspA in the region between residues 217 and 273 (data not shown). All of these Mabs will agglutinate the B31 spirochete.

Western Blot Analysis of Antibody Binding to Mutated Hypervariable Regions

Mabs were used for Western Blot analysis of the sitedirected OspA mutants induced in E.coli using the T7 expression system (Dunn, J.J. et al., Protein Expression 15 and Purification 1: 159 (1990)). E. coli cells carrying Pet9c plasmids having a site-directed OspA mutant insert were induced at mid-log phase growth with IPTG for four. hours at 37°C. Cell lysates were made by boiling an aliquot of the induced cultures in SDS gell loading dye, 20 and this material was then loaded onto a 12% SDS gell (BioRad mini-Protean II), and electrophoresed. proteins were then transferred to Imobilon-P membranes (Millipore) 70V, 2 hour at 4°C using the BioRad mini transfer system. Western analysis was carried out as 25 described by Schubach et al. (Infect. Immun. 59: 1911 (1991)).

Western Blot analysis indicated that only the 625 mutant (Ala214-Gly and Ala215-Lys) retained binding to the agglutinating monoclonal H3TS (data not shown). However, the 613/625 mutant which has additional alterations to the amino terminus of Trp216 (Ser204-Thr and Thr206-Ser) did not bind this monoclonal. Both 640 and 613/640 OspAs which have the Asn217-Asp and Gly219-Lys changes on the carboxy-

terminal side of Trp216 also failed to bind Mab H3TS. This indicated that the epitope of the B31 OspA which binds H3TS is comprised of amino acid side-chains on both sides of Trp216.

The 613/625 mutant failed to bind Mabs 105 and H5332, while the other mutants retained their ability to bind these Mabs. This is important in light of the data using fusion proteins that indicate that Mab 105 behaves more like Mab H3TS in terms of its serotype specificity and binding to OspA (Wilske, B. et al., Med. Microbiol.

Immunol. 181: 191 (1992)). The 613/625 protein has, in addition to the differences at residues Thr204 and Ser206, changes immediately amino-terminal to Trp216 (Ala214-Gly and Ala215-Lys). The abrogation of reactivity of Mabs 105 and H5332 to this protein indicated that the epitopes of OspA which bind these monoclonals are comprised of residues on the amino-terminal side of Trp216.

The two proteins carrying the Asn217-Asp and Gly219-Lys replacements on the carboxy-terminal side of Trp216

20 (OspAs 640 and 613/640) retained binding to Mabs 105 and H5332; however, they failed to react with Mab 336, a monoclonal which has been mapped with TrpE-OspA fusion proteins and by chemical cleavage to a more carboxy-terminal domain. This result may explain why Mab 336 failed to recognize the K48-type of OspA (Group 2).

It is clear that amino acids Ser204 and Thr206 play an important part in the agglutinating epitopes in the region of the B31 OspA flanking Trp216. Replacement of these two residues altered the epitopes of OspA that bind Mabs 105, H3TS and H5332. The ability of the 640 changes alone to abolish reactivity of Mab 336 indicated that Thr204 and Ser206 are not involved in direct interaction with Mab 336.

The results indicated that the epitopes of OspA which are available to Mabs that agglutinate spirochetes are comprised at least in part by amino acids in the immediate

vicinity of Trp216. Since recent circular dichroism analysis indicated that the structures of B31 and K48 OspA differ very little within this domain, it is unlikely that the changes made by mutation have radically altered the overall structure of the OspA protein (France, L.L. et al., Biochem. Biophys. Acta 1120: 59 (1992); and France et al., Biochem. Biophys Acta, submitted (1993)). This hypothesis is supported by the finding that the recombinant, mutant OspAs exhibit the same high solubility and purification properties as the parent B31 protein (data not shown).

In summary, amino acid side-chains at Ser204 and Thr206 are important for many of the agglutinating epitopes. However, a limited set of conservative changes at these sites were not sufficient to abolish binding of all of the agglutinating Mabs. These results suggested that the agglutinating epitopes of OspA are distinct, yet may have some overlap. The results also supported the hypothesis that the surface-exposed epitope around Trp216 which is thought to be important for immune recognition and neutralization is a conformationally-determined and complex domain of OspA.

EXAMPLE 3. Borrelia Strains and Proteins

Proteins and genes from any strain of Borrelia can be utilized in the current invention. Representative strains are summarized in Table I, above.

A. Genes Encoding Borrelia Proteins

The chimeric peptides of the current invention can comprise peptides derived from any Borrelia proteins.

Representative proteins include OspA, OspB, OspC, OspD, p12, p39, p41 (fla), p66, and p93. Nucleic acid sequences encoding several Borrelia proteins are presently available (see Table II, below); alternatively, nucleic acid

sequences encoding Borrelia proteins can be isolated and characterized using methods such as those described below.

Table II. References for Nucleic Acid Sequences for Several Proteins of Various Borrelia Strains

Strai n	p93	OspA	p41 (fla)
K48	X69602 (SID 67)	X62624 (SID 8)	X69610 (SID 49)
PGau	SID 73	X62387 (SID 10)	X69612 (SID 51)
DK29	-	X63412 (SID 137)	X69608 (SID 53)
PKo	X69803 (SID 77)	X65599 (SID 141)	X69613 (SID 131)
PTrob	X69604 (SID 71)	X65598 (SID 135)	X69614 (SID 55)
Ip3	-	X70365 (SID 140)	-
Ip90	ND	Kryuchechnikov, V.N. et al., J.Microbiol. Epid. Immunobiol. 12:41-44 (1988) (SID 138)	-
25015	X70365 (SID 75)	Fikrig, E.S. et al., <u>J. Immunol. 7</u> :2256- 2260 1992) SID 12)	-
B31	Perng, G.C. et al., <u>Infect.</u> <u>Immun. 59:</u> 2070- 74 (1992); Luft, B.J. et al., <u>Infect.</u> <u>Immun. 60:</u> 4309- 4321 (1992) (SID 65)	Bergstrom, S. et al., Mol. Microbiol. 3:479-486 (1989) (SID 6)	Gassmann, G.S. et al., <u>Nucl.</u> <u>Acids Res. 17</u> : 3590 (1989) (SID 127)
PKal	-	X69606 (SID 132)	X69611 (SID 129)
ZS7	-	Jonsson, M. et al., <u>Infect. Immun.</u> <u>60</u> :1845-1853 (1992) (SID 134)	-
N40	-	Kryuchechnikov, V.N. et al. (SID 133)	-
PHei	-	X65600 (SID 136)	-
ACAI	-	Kryuchechnikov, V.N. et al. (SID 142)	-
PBo	X69601 (SID 69)	X65605 (SID 139)	X69610 (SID

Numbers with an "X" prefix are GenBank data base accession numbers. SID = SEQ ID NO.

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B. <u>Isolation of Borrelia Genes</u>

Nucleic acid sequences encoding full length, lipidated proteins from known Borrelia strains were isolated using the polymerase chain reaction (PCR) as described below. addition, nucleic acid sequences were generated which encoded truncated proteins (proteins in which the lipidation signal has been removed, such as by eliminating the nucleic acid sequence encoding the first 18 amino acids, resulting in non-lipidated proteins). proteins were generated which encoded polypeptides of a particular gene (i.e., encoding a segment of the protein which has a different number of amino acids than the protein does in nature). Using similar methods as those described below, primers can be generated from known 15 nucleic acid sequences encoding Borrelia proteins and used to isolate other genes encoding Borrelia proteins. Primers can be designed to amplify all of a gene, as well as to amplify a nucleic acid sequence encoding truncated protein sequences, such as described below for OspC, or nucleic ... acid sequences encoding a polypeptide derived from a 20 Borrelia protein. Primers can also be designed to incorporate unique restriction enzyme cleavage sites into the amplified nucleic acid sequences. Sequence analysis of the amplified nucleic acid sequences can then be performed using standard techniques. 25

Cloning and Sequencing of OspA Genes and Relevant Nucleic Acid Sequences

Borrelia OspA sequences were isolated in the following manner: 100 μ l reaction mixtures containing 50 mM KCl, 10 mM TRIS-HCl (pH 8,3), 1.5 mM MgCl₂, 200 μ M each NTP, 2.5 units of TaqI DNA polymerase (Amplitaq, Perkin-Elmer/Cetus) and 100 pmol each of the 5' and 3' primers (described below) were used. Amplification was performed in a Perkin-Elmer/Cetus thermal cycler as described (Schubach, W.H. et

al., <u>Infect. Immun. 59</u>:1811-1915 (1991)). The amplicon was visualized on an agarose gel by ethidium bromide staining. Twenty nanograms of the chloroform-extracted PCR product were cloned directly into the PC-TA vector (Invitrogen) by following the manufacturer's instructions. Recombinant colonies containing the amplified fragment were selected, the plasmids were prepared, and the nucleic acid sequence of each OspA was determined by the dideoxy chaintermination technique using the Sequenase kit (United States Biochemical). Directed sequencing was performed with M13 primers followed by OspA-specific primers derived from sequences, previously obtained with M13 primers.

Because the 5' and 3' ends of the OspA gene are highly conserved (Fikrig, E.S. et al., J. Immunol. 7:2256-2260

15 (1992); Bergstrom, S. et al., Mol. Microbiol. 3: 479-486 (1989); Zumstein, G. et al., Med. Microbiol. Immunol. 181: 57-70 (1992)), the 5' and 3' primers for cloning can be based upon any known OspA sequences. For example, the following primers based upon the OspA nucleic acid sequence 20 from strain B31 were used:

5'-GGAGAATATTATGAAA-3' (-12 to +6) (SEQ ID NO. 4); and 5'-CTCCTTATTTTAAAGCG-3' (+826 to +809) (SEQ ID NO. 5). (Schubach, W.H. et al., Infect. Immun 59:1811-1915 (1991)).

OspA genes isolated in this manner include those for strains B31, K48, PGau, and 25015; the nucleic acid sequences are depicted in the sequence listing as SEQ ID NO. 6 (OspA-B31), SEQ ID NO. 8 (OspA-K48), SEQ ID NO. 10 (OspA-PGau), and SEQ ID NO. 12 (OspA-25015). An alignment of these and other OspA nucleic acid sequences is shown in Figure 42. The amino acid sequences of the proteins encoded by these nucleic acid sequences are represented as SEQ ID NO. 7 (OspA-B31), SEQ ID NO. 9 (OspA-K48), SEQ ID NO. 11 (OspA-PGau), and SEQ ID NO. 13 (OspA-25015).

The following primers were used to generate specific nucleic acid sequences of the OspA gene, to be used to

generate chimeric nucleic acid sequences (as described in Example 4):

5'-GTCTGCAAAAACCATGACAAG-3' (plus strand primer #369) (SEQ ID NO. 14);

- 5'-GTCATCAACAGAAGAAAATTC-3' (plus strand primer #357) (SEQ ID NO 15);
 - 5'-CCGGATCCATATGAAAAAATATTTATTGGG-3' (plus strand primer #607) (SEQ ID NO. 16);
- 5'-CCGGGATCCATATGGCTAAGCAAAATGTTAGC-3' (plus strand primer
- 10 #584) (SEQ ID NO. 17); 5'-GCGTTCAAGTACTCCAGA-3' (minus strand primer #200) (SEQ ID NO. 18);
 - 5'-GATATCTAGATCTTATTTTAAAGCGTT-3' (minus strand primer #586) (SEQ ID NO. 19); and
- 5'-GGATCCGGTGACCTTTTAAAGCGTTTTTAAT-3' (minus strand primer #1169) (SEQ ID NO. 20).

Cloning and Sequencing of OspB

Similar methods were also used to isolate OspB genes.

One OspB genes isolated is represented as SEQ ID NO. 21

20 (OspB-B31); its encoded amino acid sequence is SEQ ID NO.

22.

The following primers were used to generate specific nucleic acid sequences of the OspB gene, to be used in generation of chimeric nucleic acid sequences (see Example

- 5'-GGTACAATTACAGTACAA-3' (plus strand primer #721) (SEQ ID NO. 23);
 - 5'-CCGAGAATCTCATATGGCACAAAAAGGTGCTGAGTCAATTGG-3' (plus strand primer #1105) (SEQ ID NO. 24);
- 5'-CCGATATCGGATCCTATTTTAAAGCGTTTTTAAGC-3' (minus strand primer # 1106) (SEQ ID NO. 25); and 5'-GGATCCGGTGACCTTTTAAAGCGTTTTTAAG-3' (minus strand primer #1170) (SEQ ID NO. 26).

Cloning and Sequencing of OspC

Similar methods were also used to isolate OspC genes. The following primers were used to isolate entire OspC genes from *Borrelia* strains B31, K48, PKO, and pTrob:

- 5'-GTGCGCGACCATATGAAAAAGAATACATTAAGTGCG-3' (plus strand primer having Ndel site combined with start codon) (SEQ ID NO. 27), and
- 5'-GTCGGCGGATCCTTAAGGTTTTTTTGGACTTTCTGC-3' (minus strand primer having BamHl site followed by stop codon) (SEQ ID NO. 28).

The nucleic acid sequences of the OspC genes were then determined by the dideoxy chain-termination technique using the Sequenase kit (United States Biochemical). OspC genes isolated and sequenced in this manner include those for strains B31, K48, PKo, and Tro; the nucleic acid sequences are depicted in the sequence listing as SEQ ID NO. 29 (OspC-B31), SEQ ID NO. 31 (OspC-K48), SEQ ID NO. 33 (OspC-PKO), and SEQ ID NO. 35 (OspC-Tro). An alignment of these sequences is shown in Figure 38. The amino acid sequences of the proteins encoded by these nucleic acid sequences are represented as SEQ ID NO. 30 (OspC-B31), SEQ ID NO. 32 (OspC-K48), SEQ ID NO. 34 (OspC-PKO), and SEQ ID NO. 36 (OspC-Tro).

Truncated OspC genes were generated using other

25 primers. These primers were designed to amplify nucleic acid sequences, derived from the OspC gene, that lacked the nucleic acids encoding the signal peptidase sequence of the full-length protein. The primers corresponded to bp 58-75 of the natural protein, with a codon for Met-Ala attached ahead. For strain B31, the following primer was used:

5'-GTGCGCGACCATATGGCTAATAATTCAGGGAAAGAT-3' (SEQ ID NO. 37).

For strain PKo,

5'-GTGCGCGACCATATGGCTAGTAATTCAGGGAAAGGT-3' (SEQ ID NO. 38)
35 was used.

For strains pTrob and K48, 5'-GTGCGCGACCATATGGCTAATAATTCAGGTGGGGAT-3' (SEQ ID NO. 39) was used.

Additional primers were also designed to amplify
nucleic acids encoding particular polypeptides, for use in
creation of chimeric nucleic acid sequences (see Example

- 4). These primers included:
- 5'-CTTGGAAAATTATTTGAA-3' (plus strand primer #520) (SEQ ID NO. 40);
- 5'-CACGGTCACCCCATGGGAAATAATTCAGGGAAAGG-3' (plus strand primer #58) (SEQ ID NO. 41);
 - 5'-TATAGATGACAGCAACGC-3' (minus strand primer #207) (SEQ ID NO. 42); and
- 5'-CCGGTGACCCCATGGTACCAGGTTTTTTTGGACTTTCTGC-3' (minus strand primer #636) (SEQ ID NO. 43).

Cloning and Sequencing of OspD

Similar methods can be used to isolate OspD genes. An alignment of four OspD nucleic acid sequences (from strains pBo, PGau, DK29, and K48) is shown in Figure 39.

20 Cloning and Sequencing of p12

The p12 gene was similarly identified. Primers used to clone the entire p12 gene included: 5'CCGGATCCATATGGTTAAAAAAATAATATTTATTTC-3' (forward primer #
757) (SEQ ID NO. 44); and 5'-

25 GATATCTAGATCTTTAATTGCTCTGCTCACTCTTC-3' (reverse primer #758) (SEQ ID NO. 45).

To amplify a truncated p12 gene (one in which the transcribed protein is non-lipidated, and begins at amino acid 18 of the native sequence), the following primers were used: 5'-CCGGGATCCATATGGCTAGTGCAATTGGTCGTGG-3' (forward primer # 759) (SEQ ID NO. 46); and primer #758 (SEQ ID NO. 45).

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Cloning and Sequencing of p41 (fla)

A similar approach was used to clone and sequence genes encoding the p41 (fla) protein. The p41 sequences listed in Table II with GenBank accession numbers were isolated using the following primers from strain B31: 5'-ATGATTATCAATCATAAT-3' (+1 to +18) (SEQ ID NO. 47); and 5'-TCTGAACAATGACAAAAC-3' (+1008 to +991) (SEQ ID NO. 48). The nucleic acid sequences of p41 isolated in this manner are depicted in the sequence listing as SEQ ID NO. 51 (p41-PGau), and SEQ ID NO. 53 (p41-DK29). An alignment of 10 several p41 nucleic acid sequences, including those for strains B31, pKal, PGau, pBo, DK29, and pKo, is shown in Figure 41. The amino acid sequences of the proteins encoded by these nucleic acid sequences are represented as 15 SEQ ID NO. 50 (p41-K48), SEQ ID NO. 52 (p41-PGau), SEQ ID NO. 54 (p41-DK29), SEQ ID NO. 56 (p41-PTrob), and SEQ ID NO. 58 (p41-PHei).

Other primers were designed to amplify nucleic acid sequences encoding polypeptides of p41, to be used in chimeric nucleic acid sequences. These primers included: 5'-TTGGATCCGGTCACCCCATGGCTCAATATAACCAATG-3' (minus strand primer #122) (SEQ ID NO. 59); 5'-TTGGATCCGGTCACCCCATGGCTTCTCAAAATGTAAG-3' (plus strand primer # 140) (SEQ ID NO. 60);

5'-TTGGATCCGGTGACCAACTCCGCCTTGAGAAGG-3' (minus strand primer # 234) (SEQ ID NO. 61); and 5'-TTGGATCCGGTGACCTATTTGAGCATAAGATGC-3' (minus strand primer #141) (SEQ ID NO. 62).

Cloning and Sequencing of p93

The same approach was also used to clone and sequence p93 protein. Genes encoding p93, as listed in Table II with GenBank accession numbers, were isolated by this method with the following primers from strain B31:

- 5'-GGTGAATTTAGTTGGTAAGG-3' (-54 to -35) (SEQ ID NO. 63); and
- 5'-CACCAGTTTCTTTAAGCTGCTCCTGC-3' (+1117 to +1092) (SEQ ID NO. 64).
- The nucleic acid sequences of p93 isolated in this manner are depicted in the sequence listing as SEQ ID NO. 65 (p93-B31), SEQ ID NO. 67 (p93-K48) SEQ ID NO. 69 (p93-PBo), SEQ ID NO. 71 (p93-PTrob), SEQ ID NO. 73 (p93-PGau), SEQ ID NO. 75 (p93-25015), and SEQ ID NO. 77 (p93-PKo).
- The amino acid sequences of the proteins encoded by these nucleic acid sequences are represented as SEQ ID NO. 66 (p93-B31), SEQ ID NO. 68 (p93-K48) SEQ ID NO. 70 (p93-PBo), SEQ ID NO. 72 (p93-PTrob), SEQ ID NO. 74 (p93-PGau), SEQ ID NO. 76 (p93-25015), and SEQ ID NO. 78 (p93-PKo).
- Other primers were used to amplify nucleic acid sequences encoding polypeptides of p93 to be used in generating chimeric nucleic acid sequences. These primers included:
 - 5'-CCGGTCACCCCATGGCTGCTTTAAAGTCTTTA-3' (plus strand primer #475) (SEQ ID NO. 79);
 - 5'-CCGGTCACCCCATGAATCTTGATAAAGCTCAG-3' (plus strand primer #900) (SEQ ID NO. 80);
 - 5'-CCGGTCACCCCATGGATGAAAAGCTTTTAAAAAGT-3' (plus strand primer #1168) (SEQ ID NO. 81);
- 5'-CCGGTCACCCCCATGGTTGAGAAATTAGATAAG-3' (plus strand primer #1423) (SEQ ID NO. 82); and
 - 5'-TTGGATCCGGTGACCCTTAACTTTTTTAAAG-3' (minus strand primer # 2100) (SEQ ID NO. 83).

C. Expression of Proteins from Borrelia Genes

The nucleic acid sequences described above can be incorporated into expression plasmids, using standard techniques, and transfected into compatible host cells in order to express the proteins encoded by the nucleic acid

sequences. As an example, the expression the pl2 gene and the isolation of pl2 protein is set forth.

Amplification of the pl2 nucleic acid sequence was conducted with primers that included a NdeI restriction 5 site into the nucleic acid sequence. The PCR product was extracted with phenol/chloroform and precipitated with The precipitated product was digested and ligated into an expression plasmid as follows: (approximately 1 μ g) of PCR DNA was combined with 2 μ l 10X restriction buffer for NdeI (Gibco/BRL), 1 μ l NdeI (Gibco/BRL), and 2 μ l distilled water, and incubated overnight at 37°C. This mixture was subsequently combined with 3 μ l 10X buffer (buffer 3, New England BioLabs), 1 μ l BamHI (NEB), and 6 μ l distilled water, and incubated at 37° 15 for two hours. The resultant material was purified by preparative gel electrophoresis using low melting point agarose, and the band was visualized under long wave ultraviolet light and excised from the gel. The gel slice was treated with Gelase using conditions recommended by the 20 manufacturer (Epicentre Technologies). The resulting DNA pelled was resuspended in 25-50 μl of 10 mM TRIS-CL (pH 8.0) and 1 mM EDTA (TE). An aliquot of this material was ligated into the Pet9c expression vector (Dunn, J. J. et al., Protein Expression and Purification 1: 159 (1990)).

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To ligate the material into the Pet9c expression vector, 20-50 ng of p12 nucleic acid sequences cut and purified as described above was combined with 5 μ l 10 One-Phor-All (OPA) buffer (Pharmacia), 30-60 ng Pet9c cut with NdeI and BamHI, 2.5 μ l 20 mM ATP, 2 μ l T4 DNA ligase (Pharmacia) diluted 1:5 in 1X OPA buffer, and sufficient distilled water to bring the final volume to 50 μ l. mixture was incubated at 12°C overnight.

The resultant ligations were transformed into competent DH5-alpha cells and plated on nutrient agar plates containing 50 μ g/ml kanamycin and incubated

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overnight at 37 °C. DH5-alpha is used as a "storage strain" for T7 expression clones, because it is RecA deficient, so that recombination and concatenation are not problematic, and because it lacks the T7 RNA polymerase gene necessary to express the cloned gene. The use of this strain allows for cloning of potentially toxic gene products while minimizing the chance of deletion and/or rearrangement of the desired genes. Other cell lines having similar properties may also be used.

10 Kanamycin resistant colonies were single-colony purified on nutrient agar plates supplemented with kanamycin at 50 μ g/ml. A colony from each isolate was inoculated into 3-5 ml of liquid medium containing 50 μ g/ml kanamycin, and incubated at 37°C without agitation.

Plasmid DNA was obtained from 1 ml of each isolate using a hot alkaline lysis procedure (Mantiatis, T. et al., Molecular Cloning: A Laboratory Manual, cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1982)).

plasmid DNA was digested with EcoRI and BglII in the following manner: 15 μ l plasmid DNA was combined with 2 μ l 10X buffer 3 (NEB), 1 μ EcoRI (NEB), 1 μ l BglII (NEB) and 1 μ l distilled water, and incubated for two hours at 37°C. The entire reaction mixture was electrophoresed on an analytical agarose gel. Plasmids carrying the p12 insert were identified by the presence of a band corresponding to 925 base-pairs (full length p12) or 875 base-pairs (nonlipidated p12).

One or two plasmid DNAs from the full length and nonlipidated p12 clones in Pet9c were used to transform BL21 DE3 pLysS to kanamycin resistance as described by Studier et al. (Methods in Enzymology, Goeddel, D. (Ed.), Academic Press, 185: 60-89 (1990)). One or two transformants of the full length and nonlipidated clones were single-colony purified on nutrient plates containing 25 µg/ml chloramphenicol (to maintain pLysS) and 50 µg/ml

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kanamycin at 37 °C. One colony of each isolate was inoculated into liquid medium supplemented with chloramphenicol and kanamycin and incubated overnight at 37°C. The overnight culture was subcultured the following 5 morning into 500 ml of liquid broth with chloramphenicol (25 μ g/ml) and kanamycin (50 μ g/ml) and grown with aeration at 37°C in an orbital air-shaker until the absorbance at 600 nm reached 0.4-0.7. Isopropyl-thio-galactoside (IPTG) was added to a final concentration of 0.5 mM, for induction, and the culture was incubated for 3-4 hours at 10 37° as before. The induced cells were pelleted by centrifugation and resuspended in 25 ml of 20 mM NaPO4 (pH 7.7). A small aliquot was removed for analysis by gel electrophoresis. Expressing clones produced proteins which migrated at the 12 kDa position.

A crude cell lysate was prepared from the culture as described for recombinant OspA by Dunn, J.J. et al., (Protein Expression and Purification 1: 159 (1990)). crude lysate was first passed over a Q-sepharose column (Pharmacia) which had been pre-equilibrated in Buffer A: 20 10 mM NaPO4 (pH 7.7), 10 mM NaCl, 0.5 mM PMSF. The column was washed with 10 mM NaPO4, 50 mM NaCl and 0.5 mM PMSF and then pl2 was eluted in 10 mM NaPO4, 0.5 mM PMSF with a NaCl gradient from 50-400 mM. pl2 eluted approximately halfway through the gradient between 100 and 200 mM NaCl. The peak 25 fractions were pooled and dialyzed against 10 mM NaPo4 (pH 7.7), 10 mM NaCl, 0.5 mM PMSF. The protein was then concentrated and applied to a Sephadex G50 gel filtration column of approximately 50 ml bed volume (Pharmacia), in 10 mM NaPO, 200 mM NaCl, 0.5 mM PMSF. p12 would typically elute shortly after the excluded volume marker. Peak fractions were determined by running small aliquots of all fractions on a gel. The p12 peak was pooled and stored in small aliquots at -20°C.

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Example 4. Generation of Chimeric Nucleic Acid Sequences and Chimeric Proteins

A. General Protocol for Creation of Chimeric Nucleic Acid Sequences

The megaprimer method of site directed mutagenesis and its modification were used to generate chimeric nucleic acid sequences (Sarkar and Sommer, <u>Biotechniques 8(4): 404-407 (1990)</u>; Aiyar, A. and J. Leis, <u>Biotechniques 14(3): 366-369 (1993)</u>). A 5' primer for the first genomic

template and a 3' fusion oligo are used to amplify the desired region. the fusion primer consists of a 3' end of the first template (DNA that encodes the amino-proximal polypeptide of the fusion protein), coupled to a 5' end of the second template (DNA that encodes the carboxy-proximal polypeptide of the fusion protein).

The PCR amplifications are performed using Taq DNA polymerase, 10X PCR buffer, and MgCl $_2$ (Promega Corp., Madison, WI), and Ultrapure dNTPs (Pharmacia, Piscataway, NJ). One μg of genomic template 1, 5 μ of 10 μ M 5' oligo and 5 μ l of 10 μ M fusion oligo are combined with the following reagents at indicated final concentrations: 10X Buffer-Mg FREE (1X), MgCl $_2$ (2 mM), dNTP mix (200 μ M each dNTP), Taq DNA polymerase (2.5 units), water to bring final volume to 100 μ l. A Thermal Cycler (Perkin Elmer Cetus,

Norwalk, CT) is used to amplify under the following conditions: 35 cycles at 95°C for one minute, 55°C for two minutes, and 72° for three minutes. This procedure results in a "megaprimer".

The resulting megaprimer is run on a 1% TAE, 4% low-melt agarose gel. The megaprimer band is cut from the gel and purified using the Promega Magic PCR Preps DNA purification system. Purified megaprimer is then used in a second PCR step. One μg of genomic template 2, approximately 0.5 μg of the megaprimer, and 5 μ of 10 μM 3'

oligo are added to a cocktail of 10X buffer, MgCl2, dNTPs and Taq at the same final concentrations as noted above, and brought to 100 μl with water. PCR conditions are the same as above. The fusion product resulting from this amplification is also purified using the Promega Magic PCR Preps DNA purification system.

The fusion product is then ligated into TA vector and transformed into E. coli using the Invitrogen (San Diego, CA) TA Cloning Kit. Approximately 50 ng of PCR fusion 10 product is ligated to 50 ng of pCRII vector with 1X Ligation Buffer, 4 units of T4 ligase, and brought to 10 Nl with water. This ligated product mixture is incubated at 12°C overnight (approximately 14 hours). Two μ l of the ligation product mixture is added to 50 μl competent INC F' 15 cells and 2 μ beta mercaptoethanol. The cells are then incubated for 30 minutes, followed by heat shock treatment at 42°C for 60 seconds, and an ice quenching for two minutes. 450 μ l of warmed SOC media is then added to the cells, resulting in a transformed cell culture which is incubated at 37°C for one hour with slight shaking. 20 of the transformed cell culture is plated on LB + 50 $\mu g/\mu l$ ampicillin plates and incubated overnight at 37°C. white colonies are picked and added to individual overnight cultures containing 3 ml LB with ampicillin (50 $\mu g/\mu l$).

The individual overnight cultures are prepared using Promega's Magic Miniprep DNA purification system. A small amount of the resulting DNA is cut using a restriction digest as a check. DNA sequencing is then performed to check the sequence of the fusion nucleic acid sequence, 30 using the United States Biochemical (Cleveland, OH) Sequenase Version 2.0 DNA sequencing kit. Three to five μg of plasmid DNA is used per reaction. 2 μ l 2M NaOH/2mM EDTA are added to the DNA, and the volume is brought to 20 μl with water. The mixture is then incubated at room temperature for five minutes. 7 μ l water, 3 μ l 3M NaAc, 75

 μ l EtOH are added. The resultant mixture is mixed by vortex and incubated for ten minutes at -70°C, and then subjected to microfugation. After microfuge for ten minutes, the supernatant is aspirated off, and the pellet 5 is dried in the speed vac for 30 second. 6 μl water, 2 μl annealing buffer, and 2 μ l of 10 μ M of the appropriate oligo is then added. This mixture is incubated for 10 minutes at 37°C and then allowed to stand at room temperature for 10 minutes. Subsequently, 5.5 μ l of label 10 cocktail (described above) is added to each sample of the mixture, which are incubated at room temperature for an additional five minutes. 3.5 μl labeled DNA is then added to each sample which is then incubated for five minutes at 37°C. 4 μ l stop solution is added to each well. The DNA 15 is denatured at 95° for two minutes, and then placed on ice.

Clones with the desired fusion nucleic acid sequences are then recloned in frame in the pEt expression system in the lipidated (full length) and non-lipidated (truncated, 20 i.e., without first 17 amino acids) forms. The product is amplified using restriction sites contained in the PCR primers. The vector and product are cut with the same enzymes and ligated together with T4 ligase. The resultant plasmid is transformed into competent E. coli using standard transformation techniques. Colonies are screened as described earlier and positive clones are transformed into expression cells, such as E. coli BL21, for protein expression with IPTG for induction. The expressed protein in its bacterial culture lysate form and/or purified form is then injected in mice for antibody production. The mice are bled, and the sera collected for agglutination, in vitro growth inhibition, and complement- dependent and independent lysis tests.

B. Specific Chimeric Nucleic Acid Sequences

Various chimeric nucleic acid sequences were generated. The nucleic acid sequences are described as encoding polypeptides from Borrelia proteins. The chimeric 5 nucleic acid sequences are produced such that the nucleic acid sequence encoding one polypeptide is in the same reading frame as the nucleic acid sequence encoding the next polypeptide in the chimeric protein sequence encoded by the chimeric nucleic acid sequence. The proteins are 10 listed sequentially (in order of presence of the encoding sequence) in the description of the chimeric nucleic acid sequence. For example, if a chimeric nucleic acid sequence consists of bp 1-650 from OspA-1 and bp 651-820 from OspA-2 were sequenced, the sequence of the chimer would include 15 the first 650 base pairs from OspA-1 followed immediately by base pairs 651-820 of OspA-2.

OSPA-K48/OSPA-PGau A chimer of OSPA from strain
K48 (OSPA-K48) and OSPA from strain PGau (OSPA-PGau) was
generated using the method described above. This chimeric
nucleic acid sequence included bp 1-654 from OSPA-K48,
followed by bp 655-820 from OSPA-PGau. Primers used
included: the amino-terminal sequence of OSPA primer #607
(SEQ ID NO. 16); the fusion primer,
5'-AAAGTAGAAGTTTTTGAATCCCATTTTCCAGTTTTTTT-3' (minus strand
primer #668-654) (SEQ ID NO. 84); the carboxy-terminal

primer #668-654) (SEQ ID NO. 84); the carboxy-terminal sequence of OspA primer #586 (SEQ ID NO. 19); and the sequence primers #369 (SEQ ID NO. 14) and #357 (SEQ ID NO. 15). The chimeric nucleic acid sequence is presented as SEQ ID NO. 85; the chimeric protein encoded by this chimeric nucleic acid sequence is presented as SEQ ID NO. 86.

OspA-B31/OspA-PGau A chimer of OspA from strain B31 (OspA-B31) and OspA from strain PGau (OspA-PGau) was generated

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ID NO. 92.

using the method described above. This chimeric nucleic acid sequence included bp 1-651 from OspA-B31, followed by bp 652-820 from OspA-PGau. Primers used included: the fusion primer,

5 5'-AAAGTAGAAGTTTTTGAATTCCAAGCTGCAGTTTT-3' (minus strand primer #668-651) (SEQ ID NO. 87); and the sequence primer, #369 (SEQ ID NO. 14). The chimeric nucleic acid sequence is presented as SEQ ID NO. 88; the chimeric protein encoded by this chimeric nucleic acid sequence is presented as SEQ ID NO. 89.

OspA-B31/OspA-K48 A chimer of OspA from strain B31 (OspA-B31) and OspA from strain K48 (OspA-K48) was generated using the method described above. This chimeric nucleic acid sequence included bp 1-651 from OspA-B31, followed by bp 652-820 from OspA-K48. Primers used included: the fusion primer, 5'-AAAGTGGAAGTTTTTGAATTCCAAGCTGCAGTTTTTTT-3' (minus strand primer #671-651) (SEQ ID NO. 90); and the sequence primer, #369 (SEQ ID NO. 14). The chimeric nucleic acid sequence is presented as SEQ ID NO. 91; the chimeric protein encoded

by this chimeric nucleic acid sequence is presented as SEQ

OspA-B31/OspA-25015 A chimer of OspA from strain B31 (OspA-B31) and OspA from strain 25015 (OspA-25015) was generated using the method described above. This chimeric nucleic acid sequence included bp 1-651 from OspA-B31, followed by bp 652-820 from OspA-25015. Primers used included: the fusion primer, 5'-TAAAGTTGAAGTGCCTGCATTCCAAGCTGCAGTTT-3' (SEQ ID NO. 93). The chimeric nucleic acid sequence is presented as SEQ ID NO. 94; the chimeric protein encoded by

presented as SEQ ID NO. 94; the chimeric protein encoded by this chimeric nucleic acid sequence is presented as SEQ ID NO. 95.

OSPA-K48/OSPA-B31/OSPA-K48 A chimer of OSPA from strain B31 (OSPA-B31) and OSPA from strain K48 (OSPA-K48) was generated using the method described above. This chimeric nucleic acid sequence included bp 1-570 from OSPA-B31, followed by bp 570-651 from OSPA-B31, followed by bp 650-820 from OSPA-K48. Primers used included: the fusion primer, 5'-CCCCAGATTTTGAAATCTTGCTTAAAACAAC-3' (SEQ ID NO. 96); and the sequence primer, #357 (SEQ ID NO. 15). The chimeric nucleic acid sequence is presented as SEQ ID NO. 97; the chimeric protein encoded by this chimeric nucleic acid sequence is presented as SEQ ID NO. 98.

OspA-B31/OspA-K48/OspA-B31/OspA-K48

A chimer of OspA from strain B31 (OspA-B31) and OspA from strain K48 (OspA-K48) was generated using the method described above. This chimeric nucleic acid sequence included bp 1-420 from OspA-B31, followed by 420-570 from OspA-K48, followed by bp 570-650 from OspA-B31, followed by bp 651-820 from OspA-K48. Primers used included: the fusion primer, 5'-CAAGTCTGGTTCCAATTTGCTCTTGTTATTAT-3' (minus strand primer #436-420) (SEQ ID NO. 99); and the sequence primer, #357 (SEQ ID NO. 15). The chimeric nucleic acid sequence is presented as SEQ ID NO. 100; the chimeric protein encoded by this chimeric nucleic acid sequence is presented as SEQ ID NO. 101.

OspA-B31/OspB-B31 A chimer of OspA and OspB from strain
B31 (OspA-B31, OspB-B31) was generated using the method
described above. The chimeric nucleic acid sequence
included bp 1-651 from OspA-B31, followed by bp 652-820
from OspB-B31. Primers used included: the fusion primer,

5'-GTTAAAGTGCTAGTACTGTCATTCCAAGCTGCAGTTTTTTT-3' (minus
strand primer #740-651) (SEQ ID NO. 102); the carboxyterminal sequence of OspB primer #1106 (SEQ ID NO. 25); and
the sequence primer #357 (SEQ ID NO. 15). The chimeric

nucleic acid sequence is presented as SEQ ID NO. 103; the chimeric protein encoded by this chimeric nucleic acid sequence is presented as SEQ ID NO. 104.

OspA-B31/OspB-B31/OspC-B31 A chimer of OspA, OspB and OspC from strain B31 (OspA-B31, OspB-B31, and OspC-B31) was generated using the method described above. The chimeric nucleic acid sequence included bp 1-650 from OspA-B31, followed by bp 652-820 from OspB-B31, followed by bp 74-630 of OspC-B31. Primers used included: the fusion primer, 5'-TGCAGATGTAATCCCATCCGCCATTTTTAAAGCGTTTTT-3' (SEQ ID NO. 105); and the carboxy-terminal sequence of OspC primer (SEQ ID NO. 28). The chimeric nucleic acid sequence is presented as SEQ ID NO. 106; the chimeric protein encoded by this chimeric nucleic acid sequence is presented as SEQ ID NO. 107.

OSPC-B31/OSPA-B31/OSPB-B31 A chimer of OSPA, OSPB and OSPC from strain B31 (OSPA-B31, OSPB-B31, and OSPC-B31) was generated using the method described above. The chimeric nucleic acid sequence included bp 1-630 from OSPC-B31, followed by bp 52-650 from OSPA-B31, followed by bp 650-820 of OSPB-B31. Primers used included: the amino-terminal sequence of OSPC primer having SEQ ID NO. 27; the fusion primer, 5'-GCTGCTAACATTTTGCTTAGGTTTTTTTGGACTTTC-3' (minus strand primer #69-630) (SEQ ID NO. 108); and the sequence primers #520 (SEQ ID NO. 40) and #200 (SEQ ID NO. 18). The chimeric nucleic acid sequence is presented as SEQ ID NO. 109; the chimeric protein encoded by this chimeric nucleic acid sequence is presented as SEQ ID NO. 110.

30 Additional Chimeric Nucleic Acid Sequences

Using the methods described above, other chimeric nucleic acid sequences were produced. These chimeric

nucleic acid sequences, and the proteins encoded, are summarized in Table 3.

Table III Chimeric Nucleic acid Sequences and the Encoded Proteins

Chimers Generated (base pairs)	SEQ ID NO. (nt)	SEQ ID NO. (protein)
OspA (52-882) / p93 (1168-2100)	111	112
OspB (45-891) / p41 (122-234)	113	114
OspB (45-891) / p41 (122-295)	115	116
OspB (45-891) / p41 (140-234)	117	118
OspB (45-891) / p41 (140-295)	119	120
OspB (45-891) / p41 (122-234) / OspC (58-633)	121	122
OspA-Tro/OspA-Bo	137	138
OspA-PGau/OspA-Bo	139	140
OspA-B31/OspA-PGau/OspA-B31/ OspA-K48	141	142
OspA-PGau/OspA-B31/OspA-K48	143	144

C. <u>Purification of Proteins Generated by Chimeric Nucleic</u> Acid <u>Sequences</u>

The chimeric nucleic acid sequences described above, as well as chimeric nucleic acid sequences produced by the methods described above, are used to produce chimeric proteins encoded by the nucleic acid sequences. Standard methods, such as those described above in Example 3, concerning the expression of proteins from Borrelia genes, can be used to express the proteins in a compatible host organism. The chimeric proteins can then be isolated and purified using standard techniques.

If the chimeric protein is soluble, it can be purified on a Sepharose column. Insoluble proteins can be solubilized in guanidine and purified on a Ni++ column;

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alternatively, they can be solubilized in 10 mM NaPO₄ with 0.1 - 1% TRIXON X 114, and subsequently purified over an S column (Pharmacia). Lipidated proteins were generally purified by the latter method. Solubility was determined by separating both soluble and insoluble fractions of cell lysate on a 12% PAGE gel, and checking for the localization of the protein by Coomasie staining, or by Western blotting with monoclonal antibodies directed to an antigenic polypeptide of the chimeric protein.

10 Equivalents

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. such equivalents are intended to be encompassed in the scope of the following claims.

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CLAIMS

What is claimed is:

- A chimeric protein comprising two or more antigenic
 Borrelia polypeptides, wherein the antigenic Borrelia
 polypeptides which comprise the chimeric protein do
 not occur naturally in the same protein in Borrelia.
 - The chimeric protein of Claim 1, wherein the antigenic Borrelia polypeptides are from two or more different species of Borrelia.
- The chimeric protein of Claim 2, wherein the antigenic Borrelia polypeptides are derived from Borrelia proteins selected from the group consisting of: outer surface protein A, outer surface protein B, outer surface protein C, outer surface protein D, p12, p39, p41, p66, and p93.
 - 4. The chimeric protein of Claim 3, wherein the antigenic Borrelia polypeptides are from corresponding proteins from two or more different species of Borrelia.
- 5. The chimeric protein of Claim 3, wherein the antigenic
 Borrelia polypeptides are from non-corresponding
 proteins from at least two different species of
 Borrelia.
- 6. The chimeric protein of Claim 1, wherein two or more antigenic Borrelia polypeptides are from the same species of Borrelia.

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- 7. The chimeric protein of Claim 6, wherein the antigenic Borrelia polypeptides are derived from Borrelia proteins selected from the group consisting of: outer surface protein A, outer surface protein B, outer surface protein C, outer surface protein D, pl2, p39, p41, p66, and p93.
 - 8. The chimeric protein of Claim 7, wherein the antigenic Borrelia polypeptides are from the same protein.
- 9. The chimeric protein of Claim 6, wherein the antigenic Borrelia polypeptides are from different proteins.
- 10. A chimeric protein comprising two antigenic Borrelia polypeptides flanking a tryptophan residue, wherein the amino-proximal polypeptide consists of a polypeptide that is proximal from the single tryptophan residue of a first outer surface protein of Borrelia, and the carboxy-proximal polypeptide consists of a polypeptide that is distal from the single tryptophan residue of a second outer surface protein of Borrelia.
- 20 11. The chimeric protein of Claim 10, wherein the first and second outer surface proteins are from the same species of Borrelia.
- 12. The chimeric protein of Claim 11, wherein the first outer surface protein is outer surface protein A and the second outer surface protein is outer surface protein B.
 - 13. The chimeric protein of Claim 11, wherein the first outer surface protein is outer surface protein B, and

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the second outer surface protein is outer surface protein A.

- 14. The chimeric protein of Claim 10, wherein the first and second outer surface proteins are from different species of *Borrelia*.
- 15. The chimeric protein of Claim 14, wherein the first outer surface protein is outer surface protein A and the second outer surface protein is outer surface protein B.
- 10 16. The chimeric protein of Claim 14, wherein the first outer surface protein is outer surface protein B, and the second outer surface protein is outer surface protein A.
- 17. The chimeric protein of Claim 14, wherein the first
 and second outer surface proteins are corresponding
 proteins selected from the group consisting of: outer
 surface protein A and outer surface protein B.
- 18. The chimeric protein of Claim 10, wherein the first outer surface protein is outer surface protein A and the second outer surface protein is outer surface protein B.
- 19. The chimeric protein of Claim 18, wherein the aminoproximal polypeptide further comprises a first,
 second, and third hypervariable domain, the first
 hypervariable domain consisting of residues 120
 through 140 of outer surface protein A, the second
 hypervariable domain consisting of residues 150
 through 180 of outer surface protein A, and the third

hypervariable domain consisting of residues 200 through 217 of outer surface protein A.

- 20. The chimeric protein of Claim 19, wherein the first and second hypervariable domains are derived from outer surface protein A from different species of Borrelia.
- 21. The chimeric protein of Claim 10, further comprising an antigenic Borrelia polypeptide derived from a Borrelia protein selected from the group consisting of: outer surface protein A, outer surface protein B, outer surface protein C, outer surface protein D, p12, p39, p41, p66, and p93.
- 22. A nucleic acid sequence encoding a chimeric protein comprising two antigenic Borrelia polypeptides,
 wherein the two antigenic Borrelia polypeptides which comprise the chimeric protein do not occur naturally in the same protein in Borrelia.
- 23. The nucleic acid sequence of Claim 22, wherein the antigenic Borrelia polypeptides are from two or more different species of Borrelia.
 - 24. The nucleic acid sequence of Claim 23, wherein the antigenic Borrelia polypeptides are derived from Borrelia proteins selected from the group consisting of: outer surface protein A, outer surface protein B, outer surface protein C, outer surface protein D, p12, p39, p41, p66, and p93.
 - 25. The nucleic acid sequence of Claim 24, wherein the antigenic Borrelia polypeptides are from corresponding

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proteins from two or more different species of Borrelia.

- 26. The nucleic acid sequence of Claim 24, wherein two or more of the antigenic *Borrelia* polypeptides are from non-corresponding proteins from different species of *Borrelia*.
 - 27. The nucleic acid sequence of Claim 22, wherein two or more antigenic *Borrelia* polypeptides are from the same species of *Borrelia*.
- 10 28. The nucleic acid sequence of Claim 27, wherein the antigenic Borrelia polypeptides are derived from Borrelia proteins selected from the group consisting of: outer surface protein A, outer surface protein B, outer surface protein C, outer surface protein D, p12, p39, p41, p66, and p93.
 - 29. The nucleic acid sequence of Claim 28, wherein the antigenic *Borrelia* polypeptides are from the same protein.
- 30. The nucleic acid sequence of Claim 27, wherein the antigenic *Borrelia* polypeptides are from different proteins.

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- 31. A nucleic acid sequence encoding a chimeric protein comprising two antigenic Borrelia polypeptides flanking a tryptophan residue, wherein the aminoproximal polypeptide consists of a polypeptide that is proximal from the single tryptophan residue of a first outer surface protein of Borrelia, and the carboxy-proximal polypeptide consists of a polypeptide that is distal from the single tryptophan residue of a second outer surface protein of Borrelia.
- 10 32. The nucleic acid sequence of Claim 31, wherein the first and second outer surface proteins are from the same species of *Borrelia*.
- 33. The nucleic acid sequence of Claim 32, wherein the first outer surface protein is outer surface protein A and the second outer surface protein is outer surface protein B.
 - 34. The nucleic acid sequence of Claim 32, wherein the first outer surface protein is outer surface protein B, and the second outer surface protein is outer surface protein A.
 - 35. The nucleic acid sequence of Claim 31, wherein the first and second outer surface proteins are from different species of Borrelia.
- 36. The nucleic acid sequence of Claim 35, wherein the
 first outer surface protein is outer surface protein A
 and the second outer surface protein is outer surface
 protein B.

- 37. The nucleic acid sequence of Claim 35, wherein the first outer surface protein is outer surface protein B, and the second outer surface protein is outer surface protein A.
- 5 38. The nucleic acid sequence of Claim 35, wherein the first and second outer surface proteins are corresponding proteins selected from the group consisting of: outer surface protein A and outer surface protein B.
- 10 39. The nucleic acid sequence of Claim 31, wherein the first outer surface protein is outer surface protein A and the second outer surface protein is outer surface protein B.
- 40. The nucleic acid sequence of Claim 39, wherein the
 amino-proximal polypeptide further comprises a first
 and a second hypervariable domain, the first
 hypervariable domain consisting of amino acid residues
 1 through 140 of outer surface protein A, and the
 second hypervariable domain consisting of amino acid
 residues 150 through 217 of outer surface protein A.
 - 41. The nucleic acid sequence of Claim 40, wherein the first and second hypervariable domains are derived from outer surface protein A from different species of Borrelia.
- 25 42. The nucleic acid sequence of Claim 31, further comprising an antigenic Borrelia polypeptide derived from a Borrelia protein selected from the group consisting of: outer surface protein A, outer surface protein B, outer surface protein C, outer surface protein D, p12, p39, p41, p66, and p93.

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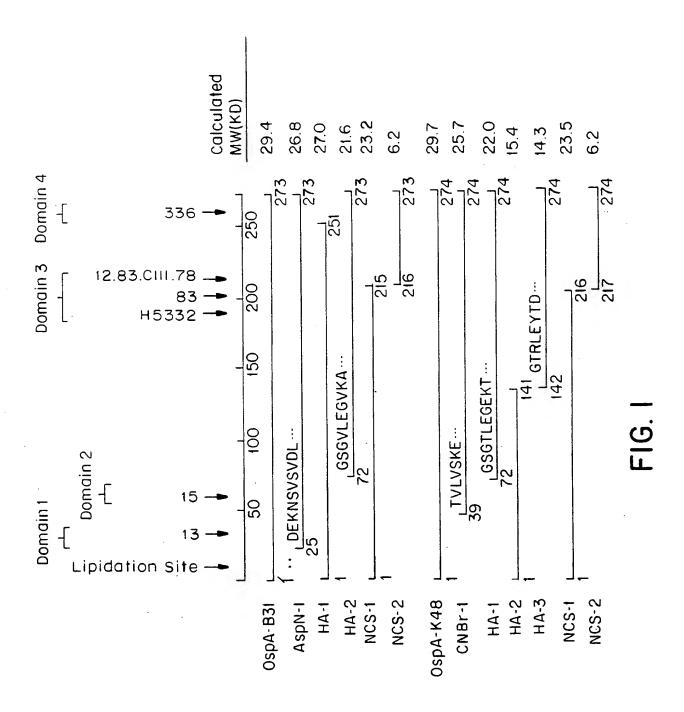
- 43. A nucleic acid sequence having a sequence selected from the group consisting of: SEQ ID NO. 85, SEQ ID NO. 88, SEQ ID NO. 91, SEQ ID NO. 94, SEQ ID NO. 97, SEQ ID NO. 100, SEQ ID NO. 103, SEQ ID NO. 106, SEQ ID NO. 109, SEQ ID NO. 111, SEQ ID NO. 113, SEQ ID NO. 115, SEQ ID NO. 117, SEQ ID NO. 119, SEQ ID NO. 121, SEQ ID NO. 137, SEQ ID NO. 139, SEQ ID NO. 141, and SEQ ID NO. 143.
- 44. A protein having an amino acid sequence selected from the group consisting of: SEQ ID NO. 86, SEQ ID NO. 89, SEQ ID NO. 92, SEQ ID NO. 95, SEQ ID NO. 98, SEQ ID NO. 101, SEQ ID NO. 104, SEQ ID NO. 107, SEQ ID NO. 110, SEQ ID NO. 112, SEQ ID NO. 114, SEQ ID NO. 116, SEQ ID NO. 118, SEQ ID NO. 120, SEQ ID NO. 122, SEQ ID NO. 138, SEQ ID NO. 140, SEQ ID NO. 142, and SEQ ID NO. 144.
- 45. A chimeric protein according to any one of claims 1 to 21 and 44 for use in therapy or diagnosis, for example as a vaccine against <u>Borrelia</u> infection, in immunodiagnostic assays to detect the presence of antibodies to <u>Borrelia</u> or to measure T-cell reactivity.
- 46. A chimeric protein according to claim 45, wherein the immunodiagnostic assay is a dot blot, Western blot, ELISA or agglutination assay.

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- 47. Use of the chimeric protein according to any one of claims 1 to 21 and 44, or the nucleic acid sequence of any one of claims 22 to 43, for the manufacture of a compound for use in therapy or diagnosis, for example as a vaccine against <u>Borrelia</u> infection, in immunodiagnostic assays to detect the presence of antibodies to <u>Borrelia</u> or to measure T-cell reactivity.
- 48. Use according to claim 47, wherein the

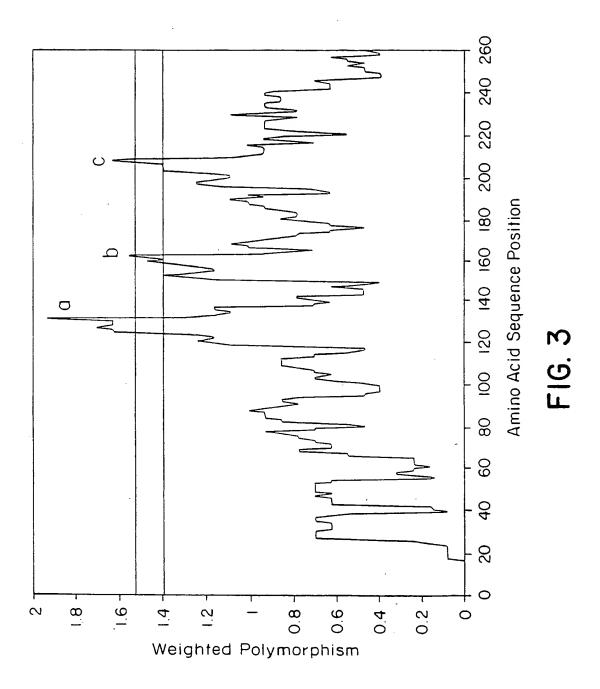
 immunodiagnostic assay is a dot blot, Western blot,

 ELISA or agglutination assay.



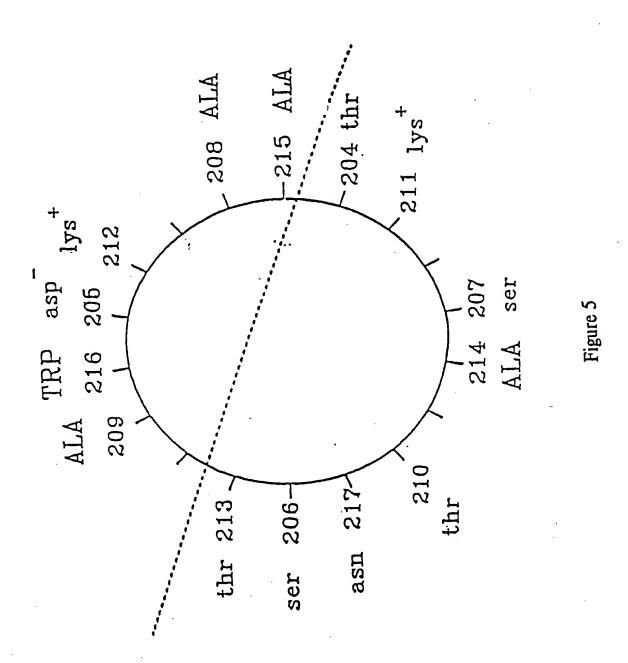
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		A-B31	A-TRO	A-K48	A-DK29	A-P/Gau	A-PKo	A-IP3	A-IP90	A-25015			A-B31	A-TRo	A-K48	A-DK29	A-P/Gau	A-PKo	A-IP3	A-IP90	
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	40	>	>	>	>	>	>	>	>	>			TAAW	VELNDSNSTQATKKTGKWDSNT	NILKSGEITVALDDSDTTQATKKTGKWDSKT	TGKW	TGAW	TGAW	TGAW	TGTW	HISNSGEITVELNDSDTTQATKKTGTWDSKT
7	39	×	×	H	H	×	×	×	ပ	×		210	ATKK			ATKK	ATKK	ATKK	ATKK	ATKK	
Domain	38	Σ	Σ	Σ	Σ	Σ	Σ	T	Σ	Σ	м		DSSA			DTTR	NTTQ.	NTTQ	NTTO	DTTO	
i r	37	ធ	ш	ပ	១	ы	ш	ш	უ	ធា	Domain	200	NISKSGEVSVELNDTDSSAATKKTAAWNSGT	ELNDS		NILKSGEITAALDDSDTTRATKKTGKWDSKI	VALNDTNTTQATKKTGAWDSKT	4LND1	VALNDTNTTQATKKTGAWDSKT	SUNTE	
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	35	Д	Q,	Ā	ď	Д	Δ,	đ	ď	Q.			SKSG					EIAKSGEVTVALNDTNTTQATKKTGAWDSKT	EIAKSGEVT	SNSG	
	34	u	ı	ឯ	IJ	ı	ı	J	1	긔		190	IN	HI	IN	IN	EI	EI	Ξ	HI	
		B31	-TRo	1-K48	-DK29	-P/Gau	PKo	-IP3	-IP90	-25015			1-B31	-TRo	-K48	DK29	P/Gau	-PKo	-1P3	-IP90	

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B31: ELNDTDSSAATKKTAAWNSGT
K48: ALDDSDTTQATKKTGKWDSKT
613: ELNDTDSSAATKKTAAWNSGT
640: ELNDTDSSAATKKTAAWDSKT
613/625: ELNDSDISAATKKTAAWDSKT
613/640: ELNDSDISAATKKTAAWDSKT

igure 4



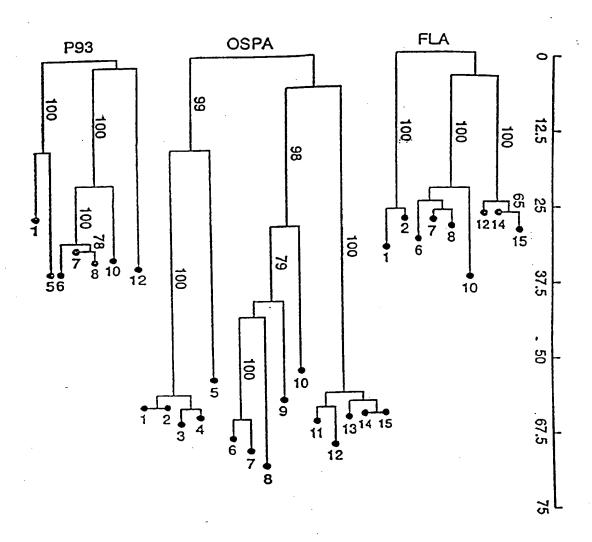


Figure 6

ATG Met 1	AAA Lys	AAA Lys	TAT Tyr	TTA Leu 5	TTG Leu	GGA Gly	ATA Ile	GGT Gly	CTA Leu 10	ATA Ile	TTA Leu	GCC Ala	TTA Leu	ATA Ile 15	GCA Ala	48
						AGC Ser										96
						AAA Lys										144
						ATT Ile 55										192
						AAT Asn										240
						AAA Lys										288
						AAA Lys										336
AAA Lys	GTA Val	ACT Thr 115	TCC Ser	AAA Lys	GAC Asp	AAG Lys	TCA Ser 120	TCA Ser	ACA Thr	GAA Glu	GAA Glu	AAA Lys 125	TTC Phe	AAT Asn	GAA Glu	384
AAA Lys	GGT Gly 130	GAA Glu	GTA Val	TCT Ser	GAA Glu	AAA Lys 135	ATA Ile	ATA Ile	ACA Thr	AGA Arg	GCA Ala 140	GAC Asp	GGA Gly	ACC Thr	AGA Arg	432
						AAA Lys										480
GTT Val	TTA Leu	AAA Lys	GGC Gly	TAT Tyr 165	GTT Val	CTT Leu	GAA Glu	GGA Gly	ACT Thr 170	CTA Leu	ACT Thr	GCT Ala	GAA Glu	AAA Lys 175	ACA Thr	528
						GGA Gly						Lys				576
AAA Lys	TCT Ser	GGG Gly 195	GAA Glu	GTT Val	TCA Ser	GTT Val	GAA Glu 200	CTT Leu	AAT Asn	GAC Asp	ACT Thr	GAC Asp 205	AGT Ser	AGT Ser	GCT Ala	624

Figure 7 (1 of 2)

				GCT Ala 215					•	672
				AAA Lys						720
				CAA Gln						768
				ACA Thr						816
AAA	TA									822

Figure 7 (2 of 2)

OSPA K48

ATG AAA AAA TAT TTA TTG GGA ATA GGT CTA ATA TTA GCC TTA ATA GCA TAC TIT TIT ATA AAT AAC CCT TAT CCA GAT TAT AAT CGG AAT TAT CGT Met Lys Lys Tyr Leu Leu Gly Ile Gly Leu Ile Leu Ala Leu Ile Ala> 70 80 60 TGT AAG CAA AAT GTT AGC AGC CTT GAT GAA AAA AAT AGC GTT TCA GTA ACA TTC GTT TTA CAA TCG TCG GAA CTA CTT TTT TTA TCG CAA AGT CAT Cys Lys Gln Asn Val Ser Ser Leu Asp Glu Lys Asn Ser Val Ser Val> 110 120 GAT TTA CCT GGT GGA ATG ACA GTT CTT GTA AGT AAA GAA AAA GAC AAA CTA AAT GGA CCA CCT TAC TGT CAA GAA CAT TCA TTT CTT TTT CTG TTT Asp Leu Pro Gly Gly Met Thr Val Leu Val Ser Lys Glu Lys Asp Lys> 170 180 160 150 GAC GGT AAA TAC AGT CTA GAG GCA ACA GTA GAC AAG CTT GAG CTT AAA CTG CCA TTT ATG TCA GAT CTC CGT TGT CAT CTG TTC GAA CTC GAA TTT Asp Gly Lys Tyr Ser Leu Glu Ala Thr Val Asp Lys Leu Glu Leu Lys> 230 240 210 220 GGA ACT TCT GAT AAA AAC AAC GGT TCT GGA ACA CTT GAA GGT GAA AAA CCT TGA AGA CTA TTT TTG TTG CCA AGA CCT TGT GAA CTT CCA CTT TTT Gly Thr Ser Asp Lys Asn Asn Gly Ser Gly Thr Leu Glu Gly Glu Lys> 260 270 280 250 ACT GAC AAA AGT AAA GTA AAA TTA ACA ATT GCT GAT GAC CTA AGT CAA TGA CTG TTT TCA TTT CAT TTT AAT TGT TAA CGA CTA CTG GAT TCA GTT Thr Asp Lys Ser Lys Val Lys Leu Thr Ile Ala Asp Asp Leu Ser Gln> 300 310 320 330 290 ACT AAA TTT GAA ATT TTC AAA GAA GAT GCC AAA ACA TTA GTA TCA AAA TGA TTT AAA CTT TAA AAG TTT CTT CTA CGG TTT TGT AAT CAT AGT TTT Thr Lys Phe Glu Ile Phe Lys Glu Asp Ala Lys Thr Leu Val Ser Lys> 370 360 340 350 AAA GTA ACC CTT AAA GAC AAG TCA TCA ACA GAA GAA AAA TTC AAC GAA TTT CAT TGG GAA TTT CTG TTC AGT AGT TGT CTT CTT TTT AAG TTG CTT Lys Val Thr Leu Lys Asp Lys Ser Ser Thr Glu Glu Lys Phe Asn Glu>

FIGURE 8 (1 of 3)

OSP A K48

400 420 430 390 AAG GGT GAA ACA TCT GAA AAA ACA ATA GTA AGA GCA AAT GGA ACC AGA TTC CCA CTT TGT AGA CTT TTT TGT TAT CAT TCT CGT TTA CCT TGG TCT Lys Gly Glu Thr Ser Glu Lys Thr Ile Val Arg Ala Asn Gly Thr Arg> 450 460 470 CTT GAA TAC ACA GAC ATA AAA AGC GAT GGA TCC GGA AAA GCT AAA GAA GAA CTT ATG TGT CTG TAT TTT TCG CTA CCT AGG CCT TTT CGA TTT CTT Leu Glu Tyr Thr Asp Ile Lys Ser Asp Gly Ser Gly Lys Ala Lys Glu> 500 510 520 490 GTT TTA AAA GAC TTT ACT CTT GAA GGA ACT CTA GCT GCT GAC GGC AAA CAA AAT TTT CTG AAA TGA GAA CTT CCT TGA GAT CGA CGA CTG CCG TTT Val Leu Lys Asp Phe Thr Leu Glu Gly Thr Leu Ala Ala Asp Gly Lys> 550 560 570 530 540 ACA ACA TTG AAA GTT ACA GAA GGC ACT GTT GTT TTA AGC AAG AAC ATT TGT TGT AAC TTT CAA TGT CTT CCG TGA CAA CAA AAT TCG TTC TTG TAA Thr Thr Leu Lys Val Thr Glu Gly Thr Val Val Leu Ser Lys Asn Ile> 600 590 610 580 TTA AAA TCC GGA GAA ATA ACA GTT GCA CTT GAT GAC TCT GAC ACT ACT AAT TTT AGG CCT CTT TAT TGT CAA CGT GAA CTA CTG AGA CTG TGA TGA Leu Lys Ser Gly Glu Ile Thr Val Ala Leu Asp Asp Ser Asp Thr Thr> 660 640 650 CAG GCT ACT AAA AAA ACT GGA AAA TGG GAT TCA AAA ACT TCC ACT TTA GTC CGA TGA TTT TTT TGA CCT TTT ACC CTA AGT TTT TGA AGG TGA AAT Gln Ala Thr Lys Lys Thr Gly Lys Trp Asp Ser Lys Thr Ser Thr Leu> 700 680 690 710 720 ACA ATT AGT GTG AAT AGC CAA AAA ACC AAA AAC CTT GTA TTC ACA AAA TGT TAA TCA CAC TTA TCG GTT TTT TGG TTT TTG GAA CAT AAG TGT TTT Thr Ile Ser Val Asn Ser Gln Lys Thr Lys Asn Leu Val Phe Thr Lys> 730 750 760 GAA GAC ACA ATA ACA GTA CAA AAA TAC GAC TCA GCA GGC ACC AAT CTA CTT CTG TGT TAT TGT CAT GTT TTT ATG CTG AGT CGT CCG TGG TTA GAT Glu Asp Thr Ile Thr Val Gln Lys Tyr Asp Ser Ala Gly Thr Asn Leu>

FIGURE 8 (2 of 3)

BASCOCIO: -MIC DESCOCA

Osp A K-48

GAA GGC AAA GCA GTC GAA ATT ACA ACA CTT AAA GAA CTT AAA AAC GCT CTT CCG TTT CGT CAG CTT TAA TGT TGT GAA TTT CTT GAA TTT TTG CGA Glu Gly Lys Ala Val Glu Ile Thr Thr Leu Lys Glu Leu Lys Asn Ala>

OSP A K48

· #

820

TTA AAA TAA AAT TTT ATT Leu Lys ***>

FIGURE 8 (3 of 3)

OSP A PGAU

10					20 . 30				40						
	•		•	•		•		•	•		•		•	•	
ATG .	AAA	AAA	ŢĄŢ	TTA	TTG	GGA	ATA	GGT	CTA	ATA	TTA	GCC	TTA	ATA	GCA
TAC '	TTT	TTT	ATA	AAT	AAC	CCT	TAT	CCA	GAT	TAT	TAA	CGG	AAT	TAT	CGT
Met	Lys	Lys	Tyr	Leu	Leu	Gly	Ile	Gly	Leu	Ile	Leu	Ala	Leu	Ile	Ala>
						-				80			90		
50			60		•	•	0			*		•	90		•
TGC .	A A G	- -		СТТ	AGC	AGC	CTT	GAT	GAA	AAA	AAC	AGC	GCT	TCA	GTA
ACG	ተጥር	GTT	TTA	CAA	TCG	TCG	GAA	CTA	CTT	TTT	TTG	TCG	CGA	AGT	CAT
CVS	Lvs	Gln	Asn	Val	Ser	Ser	Leu	Asp	Glu	Lys	Asn	Ser	Ala	Ser	Val>
٠,٠	_, _							•			-				
10	0		1	10			120			13	0		1	140	
	•.	•		*		•	•		•		•	•		•	
GAT															
CTA	AAC	GGA	CCA	CTC	TAC	TTT	CAA	GAA	CAT	TCA	111	CTT	TTT	CTG	TTT
Asp	Leu	Pro	Gly	Glu	met	Lys	vaı	Leu	vai	Ser	Lys	GIU	Lys	ASP	Lys>
	150			16	50		1	70			180			19	90
•			•	•	•	•		•		•	•		•		•
GAC	GGT	AAG	TAC	AGT	CTA	AAG	GCA	ACA	GTA	GAC	AAG	ATT	GAG	CTA	AAA
				TCA											
Asp	Gly	Lys	Tyr	Ser	Leu	Lys	Ala	Thr	Val	Asp	Lys	Ile	Glu	Leu	Lys>
															5.4.5
_	7	200		_	210			22	20		•	230			240
663	N C-T	TCT	C 2 T	AAA	GAC	דממ	GGT	тст	GGA	GTG	ښک	GAA	ССТ	ACA	AAA
				TTT											
															Lys>
017	••••			_, _			-		-				-		-
		2	50		;	260			270			- 2	В0		
	•		•	•		•		•	•		•		•	•	
				AAA											
			TCA	$ extbf{TTT}$	CGT	TTT		TGT	TAA	CGA					
	_	_	. .			•	•	5			>		*		
Asp	Asp	Lys	Ser	Lys		Lys	Leu	Thr			Asp	Asp	Leu	Ser	-, -
, -	Asp	Lys	÷					Thr	Ile	Ala	Asp	Asp		Ser	-, -
290	Asp	Lys	Ser 300				Leu 10	Thr	Ile		Asp	Asp	330	Ser	•
290 •	ACA	·	300 • GAA	Lys	Ala TTA	3 AAA	10 GAA	• GAT	Ile	Ala 320 . •	ACA	• TTA	330 •	TCA	• AGA
290 ACC TGG	ACA TGT	• TTC AAG	300 GAA CTT	Lys CTT GAA	Ala TTA AAT	3 AAA TTT	10 GAA	GAT	Ile GGC CCG	Ala 320 AAA TTT	ACA TGT	• TTA	330 • GTG CAC	TCA AGT	AGA TCT
290 ACC TGG	ACA TGT	• TTC AAG	300 GAA CTT	Lys CTT GAA	Ala TTA AAT	3 AAA TTT	10 GAA	GAT	Ile GGC CCG	Ala 320 AAA TTT	ACA TGT	• TTA	330 • GTG CAC	TCA AGT	• AGA
290 ACC TGG Thr	ACA TGT Thr	• TTC AAG	300 GAA CTT Glu	Lys CTT GAA Leu	Ala TTA AAT	3 AAA TTT	GAA CTT	GAT CTA Asp	Ile GGC CCG	Ala 320 AAA TTT Lys	ACA TGT Thr	• TTA	330 • GTG CAC	TCA AGT Ser	AGA TCT
290 ACC TGG Thr	ACA TGT	• TTC AAG	300 GAA CTT Glu	Lys CTT GAA	Ala TTA AAT	3 AAA TTT	10 GAA	GAT CTA Asp	Ile GGC CCG	Ala 320 AAA TTT Lys	ACA TGT	• TTA	330 • GTG CAC	TCA AGT	AGA TCT
290 ACC TGG Thr	ACA TGT Thr	TTC AAG Phe	GAA CTT Glu	CTT GAA Leu	TTA AAT Leu	3 AAA TTT Lys	GAA CTT Glu	GAT CTA Asp	GGC CCG Gly	Ala 320 AAA TTT Lys	ACA TGT Thr	TTA	330 GTG CAC Val	TCA AGT Ser	AGA TCT Arg>
290 ACC TGG Thr	ACA TGT Thr 40	TTC AAG Phe	300 GAA CTT Glu	CTT GAA Leu 350	TTA AAT Leu	AAA TTT Lys	GAA CTT Glu 360	GAT CTA Asp	GGC CCG Gly	Ala 320 AAA TTT Lys 3	ACA TGT Thr 70	* TTA AAT Leu	330 GTG CAC Val	TCA AGT Ser 380	AGA TCT
290 ACC TGG Thr	ACA TGT Thr	• TTC AAG	300 GAA CTT Glu	Lys CTT GAA Leu	Ala TTA AAT	3 AAA TTT	GAA CTT	GAT CTA Asp	Ile GGC CCG	Ala 320 AAA TTT Lys	ACA TGT Thr	• TTA	330 • GTG CAC	TCA AGT Ser	AGA TCT

FIGURE 9 (1 of 3)

OSP A PGAU

400 410 420 AAA GGT GAA TTG TCT GCA AAA ACC ATG ACA AGA GAA AAT GGA ACC AAA TTT CCA CTT AAC AGA CGT TTT TGG TAC TGT TCT CTT TTA CCT TGG TTT Lys Gly Glu Leu Ser Ala Lys Thr Met Thr Arg Glu Asn Gly Thr Lys> 470 460 450 CTT GAA TAT ACA GAA ATG AAA AGC GAT GGA ACC GGA AAA GCT AAA GAA GAA CTT ATA TGT CTT TAC TTT TCG CTA CCT TGG CCT TTT CGA TTT CTT Leu Glu Tyr Thr Glu Met Lys Ser Asp Gly Thr Gly Lys Ala Lys Glu> 500 510 520 490 GTT TTA AAA AAG TTT ACT CTT GAA GGA AAA GTA GCT AAT GAT AAA GTA CAA AAT TTT TTC AAA TGA GAA CTT CCT TTT CAT CGA TTA CTA TTT CAT Val Leu Lys Lys Phe Thr Leu Glu Gly Lys Val Ala Asn Asp Lys Val> 550 560 570 540 530 ACA TTG GAA GTA AAA GAA GGA ACC GTT ACT TTA AGT AAG GAA ATT GCA TGT AAC CTT CAT TTT CTT CCT TGG CAA TGA AAT TCA TTC CTT TAA CGT Thr Leu Glu Val Lys Glu Gly Thr Val Thr Leu Ser Lys Glu Ile Ala> 610 590 600 AAA TCT GGA GAA GTA ACA GTT GCT CTT AAT GAC ACT AAC ACT ACT CAG TTT AGA CCT CTT CAT TGT CAA CGA GAA TTA CTG TGA TTG TGA TGA GTC Lys Ser Gly Glu Val Thr Val Ala Leu Asn Asp Thr Asn Thr Thr Gln> 660 650 640 630 GCT ACT AAA AAA ACT GGC GCA TGG GAT TCA AAA ACT TCT ACT TTA ACA CGA TGA TTT TTT TGA CCG CGT ACC CTA AGT TTT TGA AGA TGA AAT TGT Ala Thr Lys Lys Thr Gly Ala Trp Asp Ser Lys Thr Ser Thr Leu Thr> 680 .. 690 700 710 ATT AGT GTT AAC AGC AAA AAA ACT ACA CAA CTT GTG TTT ACT AAA CAA TAA TCA CAA TTG TCG TTT TTT TGA TGT GTT GAA CAC AAA TGA TTT GTT Ile Ser Val Asn Ser Lys Lys Thr Thr Gln Leu Val Phe Thr Lys Gln> 740 750 TAC ACA ATA ACT GTA AAA CAA TAC GAC TCC GCA GGT ACC AAT TTA GAA ATG TGT TAT TGA CAT TTT GTT ATG CTG AGG CGT CCA TGG TTA AAT CTT Tyr Thr Ile Thr Val Lys Gln Tyr Asp Ser Ala Gly Thr Asn Leu Glu>

FIGURE 9 (2 of 3)

OSP A PGAU

770 780 790 800 810

GGC ACA GCA GTC GAA ATT AAA ACA CTT GAT GAA CTT AAA AAC GCT TTA

CCG TGT CGT CAG CTT TAA TTT TGT GAA CTA CTT GAA TTT TTG CGA AAT

Gly Thr Ala Val Glu Ile Lys Thr Leu Asp Glu Leu Lys Asn Ala Leu>

820 • AAA TAA

TTT ATT
Lys ***>

FIGURE 9 (3 of 3)

ATG Met 1	AAA Lys	AAA Lys	TAT Tyr	TTA Leu 5	TTG Leu	GGA Gly	ATA Ile	GGT Gly	CTA Leu 10	ATA Ile	TTA Leu	GCT Ala	TTA Leu	ATA Ile 15	GCA Ala	48
TGT Cys	AAG Lys	CAA Gln	AAT Asn 20	GTT Val	AGC Ser	AGC Ser	CTT Leu	GAC Asp 25	GAG Glu	AAA Lys	AAC Asn	AGC Ser	GTT Val 30	TCA Ser	GTA Val	96
GAT Asp	TTG Leu	CCT Pro 35	GGT Gly	GAA Glu	ATG Met	AAA Lys	GTT Val 40	CTT Leu	GTA Val	AGC Ser	AAA Lys	GAA Glu 45	AAA Lys	GAC Asp	AAA Lys	144
						ATG Met 55										192

Figure 10 (1 of 2)

GGA Gly 65	ACA Thr	TCT Ser	GAT Asp	AAA Lys	AAC Asn 70	AAT Asn	GGA Gly	TCT Ser	GGG Gly	GTG Val 75	CTT Leu	GAA Glu	GGC Gly	GTA Val	AAA Lys 80	240
GCT Ala	GAC Asp	AAA Lys	AGC Ser	AAA Lys 85	GTA Val	AAA Lys	TTA Leu	ACA Thr	GTT Val 90	TCT Ser	GAC Asp	GAT Asp	CTA Leu	AGC Ser 95	ACA Thr	288
ACC Thr	ACA Thr	CTT Leu	GAA Glu 100	GTT Val	TTA Leu	AAA Lys	GAA Glu	GAT Asp 105	GGC Gly	AAA Lys	ACA Thr	TTA Leu	GTG Val 110	TCA Ser	AAA Lys	336
AAA Lys	AGA Arg	ACT Thr 115	TCT Ser	AAA Lys	GAT Asp	AAG Lys	TCA Ser 120	TCA Ser	ACA Thr	GAA Glu	GAA Glu	AAG Lys 125	TTC Phe	TAA Asn	GAA Glu	384
AAA Lys	GGC Gly 130	GAA Glu	TTA Leu	GTT Val	GAA Glu	AAA Lys 135	ATA Ile	ATG Met	GCA Ala	AGA Arg	GCA Ala 140	113	GGA Gly	ACC Thr	ATA Ile	432
CTT Leu 145	GAA Glu	TAC Tyr	ACA Thr	GGA Gly	ATT Ile 150	AAA Lys	AGC Ser	GAT Asp	GGA Gly	TCC Ser 155	GGA Gly	AAA Lys	GCT Ala	AAA Lys	GAA Glu 160	480
ACT Thr	TTA Leu	AAA Lys	GAA Glu	TAT Tyr 165	Val	CTT Leu	GAA Glu	GGA Gly	ACT Thr 170	Leu	ACT Thr	GCT Ala	GAA Glu	AAA Lys 175	GCA Ala	528
ACA Thr	TTG Leu	GTG Val	GTT Val 180	Lys	GAA Glu	GGA Gly	ACT Thr	GTT Val 185	THE	TTA Leu	AGT Ser	AAG	CAC His 190		TCA Ser	576
AAA Lys	TCT Ser	GGA Gly 195	Glu	GTA Val	ACA Thr	GCT	GAA Glu 200	Leu	AAT Asn	GAC Asp	ACT	GAC Asp 205	, 561	ACT Thr	CAA Gln	624
GCT Ala	ACT Thr 210	Lys	AAA Lys	ACI Thr	G GGG	AAA Lys 215	Trp	GAT Asp	GCA Ala	GGC	Thr 220	. ser	ACT Thr	TTA Leu	ACA Thr	672
ATT 11e 225	Thr	GTA Val	AAC Asr	AAC A Ast	AAA Lye 230	Lys	ACI The	AAA Lys	A GCC s Ala	CTI Lev 235	ı va.	A TTT	ACA Thr	LYS	CAA Gln 240	720
GAC Asp	ACA Thr	ATT	ACI Thi	A TCA Sea 249	: Glr	AAA Lys	TAC Tyl	C GAG	TCA Ser 250	AL	GG Gl	A ACC	C AAC Ran	25	GAA 1 Glu 5	768
GG(Gl _y	ACI Thi	A GCA	A GTG a Vai 26	l Gl	A AT	C AAI E Lys	A AC	A CT r Lev 26	n wai	GAI Glu	A CT	T AA u Ly	A AA(6 Asi 27(• ••	r TTA a Leu	816
AG! Arg																819

Figure 10 (2 of 2)

17//33

OSP B B-31 Sequence Range: 1 to 891

30 20 10 ATG AGA TTA TTA ATA GGA TTT GCT TTA GCG TTA GCT TTA ATA GGA TGT TAC TOT AAT AAT TAT COT AAA CGA AAT CGC AAT CGA AAT TAT COT ACA Met Arg Leu Leu Ile Gly Phe Ala Leu Ala Leu Ile Gly Cys> 80 90 70 60 50 GCA CAA AAA GGT GCT GAG TCA ATT GGT TCT CAA AAA GAA AAT GAT CTA CGT GTT TTT CCA CGA CTC AGT TAA CCA AGA GTT TTT CTT TTA CTA GAT Ala Gln Lys Gly Ala Glu Ser Ile Gly Ser Gln Lys Glu Asn Asp Leu> 120 130 110 • AAC CTT GAA GAC TCT AGT AAA AAA TCA CAT CAA AAC GCT AAA CAA GAC TTG GAA CTT CTG AGA TCA TTT TTT AGT GTA GTT TTG CGA TTT GTT CTG Asn Leu Glu Asp Ser Ser Lys Lys Ser His Gln Asn Ala Lys Gln Asp> 190 150 170 160 150 CTT CCT GCG GTG ACA GAA GAC TCA GTG TCT TTG TTT AAT GGT AAA AAA GAA GGA CGC CAC TGT CTT CTG AGT CAC AGA AAC AAA TTA CCA TTA TTT Leu Pro Ala Val Thr Glu Asp Ser Val Ser Leu Phe Asm Gly Asm Lys> 230 210 220 ATT TTT GTA AGC AAA GAA AAA AAT AGC TCC GGC AAA TAT GAT TTA AGA TAA AAA CAT TCG TTT CTT TTT TTA TCG AGG CCG TTT ATA CTA AAT TCT Ile Phe Val Ser Lys Glu Lys Asn Ser Ser Gly Lys Tyr Asp Leu Arg> 280 270 260 250 GCA ACA ATT GAT CAG GTT GAA CTT AAA GGA ACT TCC GAT AAA AAC AAT CGT TGT TAA CTA GTC CAA CTT GAA TTT CCT TGA AGG CTA TTT TTG TTA Ala Thr Ile Asp Gln Val Glu Leu Lys Gly Thr Ser Asp Lys Asn Asn> 330 310 320 300 290 • GGT TCT GGA ACC CTT GAA GGT TCA AAG CCT GAC AAG AGT AAA GTA AAA CCA AGA CCT TGG GAA CTT CCA AGT TTC GGA CTG TTC TCA TTT CAT TTT Gly Ser Gly Thr Leu Glu Gly Ser Lys Pro Asp Lys Ser Lys Val Lys> 370 360 350 340 • TTA AÇA GTT TCT GCT GAT TTA AAC ACA GTA ACC TTA GAA GCA TTT GAT AAT TGT CAA AGA CGA CTA AAT TTG TGT CAT TGG AAT CTT CGT AAA CTA Leu Thr Val Ser Ala Asp Leu Asn Thr Val Thr Leu Glu Ala Phe Asp> 420 400 410

FIGURE 11 (1 of 3)

GCC AGC AAC CAA AAA ATT TCA AGT AAA GTT ACT AAA AAA CAG GGG TCA CGG TCG TTG GTT TTT TAA AGT TCA TTT CAA TGA TTT TTT GTC CCC AGT Ala Ser Asn Gln Lys Ile Ser Ser Lys Val Thr Lys Lys Gln Gly Ser> 450 470 460 450 ATA ACA GAG GAA ACT CTC AAA GCT AAT AAA TTA GAC TCA AAG AAA TTA TAT TGT CTC CTT TGA GAG TTT CGA TTA TTT AAT CTG AGT TTC TTT AAT Ile Thr Glu Glu Thr Leu Lys Ala Asn Lys Leu Asp Ser Lys Lys Leu> 510 500 490 • ACA AGA TCA AAC GGA ACT ACA CTT GAA TAC TCA CAA ATA ACA GAT GCT TGT TCT AGT TTG CCT TGA TGT GAA CTT ATG AGT GTT TAT TGT CTA CGA Thr Arg Ser Asn Gly Thr Thr Leu Glu Tyr Ser Gln Ile Thr Asp Ala> 570 550 560 540 530 • . GAC AAT GCT ACA AAA GCA GTA GAA ACT CTA AAA AAT AGC ATT AAG CTT CTG TTA CGA TGT TTT CGT CAT CTT TGA GAT TTT TTA TCG TAA TTC GAA Asp Asn Ala Thr Lys Ala Val Glu Thr Leu Lys Asn Ser Ile Lys Leu> 600 610 590 580 GAA GGA AGT CTT GTA GTC GGA AAA ACA ACA GTG GAA ATT AAA GAA GGT CTT CCT TCA GAA CAT CAG CCT TTT TGT TGT CAC CTT TAA TTT CTT CCA Glu Gly Ser Leu Val Val Gly Lys Thr Thr Val Glu Ile Lys Glu Gly> 650 660 640 ACT GTT ACT CTA AAA AGA GAA ATT GAA AAA GAT GGA AAA GTA AAA GTC TGA CAA TGA GAT TTT TCT CTT TAA CTT TTT CTA CCT TTT CAT TTT CAG Thr Val Thr Leu Lys Arg Glu Ile Glu Lys Asp Gly Lys Val Lys Val> 700 710 690 680 TTT TTG AAT GAC ACT GCA GGT TCT AAC AAA AAA ACA GGT AAA TGG GAA AAA AAC TTA CTG TGA CGT CCA AGA TTG TTT TTT TGT CCA TTT ACC CTT Phe Leu Asn Asp Thr Ala Gly Ser Asn Lys Lys Thr Gly Lys Trp Glu> 750 740 • GAC AGT ACT AGC ACT TTA ACA ATT AGT GCT GAC AGC AAA AAA ACT AAA CTG TCA TGA TCG TGA AAT TGT TAA TCA CGA CTG TCG TTT TTT TGA TTT Asp Ser Thr Ser Thr Leu Thr Ile Ser Ala Asp Ser Lys Lys Thr Lys> 790 800 780 770 • GAT TTG GTG TTC TTA ACA GAT GGT ACA ATT ACA GTA CAA CAA TAC AAC CTA AAC CAC AAG AAT TGT CTA CCA TGT TAA TGT CAT GTT ATG TTG Asp Leu Val Phe Leu Thr Asp Gly Thr Ile Thr Val Gln Gln Tyr Asn>

820 830 840 850 860

ACA GCT GGA ACC AGC CTA GAA GGA TCA GCA AGT GAA ATT AAA AAT CTT
TGT CGA CCT TGG TCG GAT CTT CCT AGT CGT TCA CTT TAA TTT TTA GAA
Thr Ala Gly Thr Ser Leu Glu Gly Ser Ala Ser Glu Ile Lys Asn Leu>

870 880 890

TCA GAG CTT AAA AAC GCT TTA AAA TAA
AGT CTC GAA TTT TTG CGA AAT TTT ATT
Ser Glu Leu Lys Asn Ala Leu Lys ***>

FIGURE 11 (3 of 3)

OspC-B31 Sequence Range: 1 to 633

40 30 20 ATG AAA AAG AAT ACA TTA AGT GCG ATA TTA ATG ACT TTA TTT TTA TTT TAC TIT TIC TIA TGT AAT TCA CGC TAT AAT TAC TGA AAT AAA AAT AAA Met Lys Lys Asn Thr Leu Ser Ala Ile Leu Met Thr Leu Phe Leu Phe> 80 70 60 50 ATA TCT TGT AAT AAT TCA GGG AAA GAT GGG AAT ACA TCT GCA AAT TCT TAT AGA ACA TTA TTA AGT CCC TTT CTA CCC TTA TGT AGA CGT TTA AGA Ile Ser Cys Asn Asn Ser Gly Lys Asp Gly Asn Thr Ser Ala Asn Ser> 130 120 110 100 GCT GAT GAG TCT GTT AAA GGG CCT AAT CTT ACA GAA ATA AGT AAA AAA CGA CTA CTC AGA CAA TTT CCC GGA TTA GAA TGT CTT TAT TCA TTT TTT Ala Asp Glu Ser Val Lys Gly Pro Asn Leu Thr Glu Ile Ser Lys Lys> 180 170 160 150 ATT ACG GAT TCT AAT GCG GTT TTA CTT GCT GTG AAA GAG GTT GAA GCG TAA TGC CTA AGA TTA CGC CAA AAT GAA CGA CAC TTT CTC CAA CTT CGC Ile Thr Asp Ser Asn Ala Val Leu Leu Ala Val Lys Glu Val Glu Ala> 230 220 210 200 * TTG CTG TCA TCT ATA GAT GAA ATT GCT GCT AAA GCT ATT GGT AAA AAA AAC GAC AGT AGA TAT CTA CTT TAA CGA CGA TTT CGA TAA CCA TTT TTT Leu Leu Ser Ser Ile Asp Glu Ile Ala Ala Lys Ala Ile Gly Lys Lys> 280 270 260 250 ATA CAC CAA AAT AAT GGT TTG GAT ACC GAA TAT AAT CAC AAT GGA TCA TAT GTG GTT TTA TTA CCA AAC CTA TGG CTT ATA TTA GTG TTA CCT AGT Ile His Gln Asn Asn Gly Leu Asp Thr Glu Tyr Asn His Asn Gly Ser> 330 320 310 300 290 TTG TTA GCG GGA CGT TAT GCA ATA TCA ACC CTA ATA AAA CAA AAA TTA AAC AAT CGC CCT GCA ATA CGT TAT AGT TGG GAT TAT TTT GTT TTT AAT Leu Leu Ala Gly Arg Tyr Ala Ile Ser Thr Leu Ile Lys Gln Lys Leu> 380 370 360 350 340 CTA CCT AAC TIT TTA CTT CCT AAT TTC CTT TTT TAA CTA CGC CGA TTC

FIGURE 12 (1 of 2)

Asp Gly Leu Lys Asn Glu Gly Leu Lys Glu Lys Ile Asp Ala Ala Lys>

OspC-B31

430 400 410 390 AAA TGT TCT GAA ACA TTT ACT AAT AAA TTA AAA GAA AAA CAC ACA GAT TTT ACA AGA CTT TGT AAA TGA TTA TTT AAT TTT CTT TTT GTG TGT CTA Lys Cys Ser Glu Thr Phe Thr Asn Lys Leu Lys Glu Lys His Thr Asp> 480 470 460 450 440 CTT GGT AAA GAA GGT GTT ACT GAT GCT GAT GCA AAA GAA GCC ATT TTA GAA CCA TTT CTT CCA CAA TGA CTA CGA CTA CGT TTT CTT CGG TAA AAT Leu Gly Lys Glu Gly Val Thr Asp Ala Asp Ala Lys Glu Ala Ile Leu> 510 ----500 490 AAA ACA AAT GGT ACT AAA ACT AAA GGT GCT GAA GAA CTT GGA AAA TTA TTT TGT TTA CCA TGA TTT TGA TTT CCA CGA CTT CTT GAA CCT TTT AAT Lys Thr Asn Gly Thr Lys Thr Lys Gly Ala Glu Glu Leu Gly Lys Leu> 570 560 550 540 530 TTT GAA TCA GTA GAG GTC TTG TCA AAA GCA GCT AAA GAG ATG CTT GCT AAA CTT AGT CAT CTC CAG AAC AGT TTT CGT CGA TTT CTC TAC GAA CGA Phe Glu Ser Val Glu Val Leu Ser Lys Ala Ala Lys Glu Met Leu Ala> 600 610 620 590 AAT TCA GTT AAA GAG CTT ACA AGC CCT GTT GTG GCA GAA AGT CCA AAA TTA AGT CAA TTT CTC GAA TGT TCG GGA CAA CAC CGT CTT TCA GGT TTT Asn Ser Val Lys Glu Leu Thr Ser Pro Val Val Ala Glu Ser Pro Lys> 630 AAA CCT TAA TTT GGA ATT Lys Pro ***>

FIGURE 12 (2 of 2)

OspC-K48
Sequence Range: 1 to 630

22/133

40 10 30 20 ATG AAA AAG AAT ACA TTA AGT GCG ATA TTA ATG ACT TTA TTT TTA TTT TAC TTT TTC TTA TGT AAT TCA CGC TAT AAT TAC TGA AAT AAA AAT AAA Met Lys Lys Asn Thr Leu Ser Ala Ile Leu Met Thr Leu Phe Leu Phe> 70 80 60 ATA TCT TGT AAT AAT TCA GGT GGG GAT ACC GCA TCT ACT AAT CCT GAT TAT AGA ACA TTA TTA AGT CCA CCC CTA TGG CGT AGA TGA TTA GGA CTA Ile Ser Cys Asn Asn Ser Gly Gly Asp Thr Ala Ser Thr Asn Pro Asp> 100 110 120 GAG TCT GCA AAA GGA CCT AAT CTT ACA GTA ATA AGC AAA AAA ATT ACA CTC AGA CGT TTT CCT GGA TTA GAA TGT CAT TAT TCG TTT TTT TAA TGT Glu Ser Ala Lys Gly Pro Asn Leu Thr Val Ile Ser Lys Lys Ile Thr> 170 160 180 150 190 GAT TCT AAT GCA TTT GTA CTG GCT GTG AAA GAA GTT GAG GCT TTG ATC CTA AGA TTA CGT AAA CAT GAC CGA CAC TTT CTT CAA CTC CGA AAC TAG Asp Ser Asn Ala Phe Val Leu Ala Val Lys Glu Val Glu Ala Leu Ile> 230 200 210 220 240 TCA TCT ATA GAT GAA CTT GCT AAT AAA GCT ATT GGT AAA GTA ATA CAT AGT AGA TAT CTA CTT GAA CGA TTA TTT CGA TAA CCA TTT CAT TAT GTA Ser Ser Ile Asp Glu Leu Ala Asn Lys Ala Ile Gly Lys Val Ile His> 250 260 270 CAN ANT ANT GGT TTA ANT GCT ANT GCG GGT CAN ANC GGA TCA TTG TTA GTT TTA TTA CCA AAT TTA CGA TTA CGC CCA GTT TTG CCT AGT AAC AAT Gln Asn Asn Gly Leu Asn Ala Asn Ala Gly Gln Asn Gly Ser Leu Leu> 290 300 310 320 GCA GGA GCC TAT GCA ATA TCA ACC CTA ATA ACA GAA AAA TTA AGT AAA CGT CCT CGG ATA CGT TAT AGT TGG GAT TAT TGT CTT TTT AAT TCA TTT Ala Gly Ala Tyr Ala Ile Ser Thr Leu Ile Thr Glu Lys Leu Ser Lys> 340 350 360 370 380 TTG AAA AAT TCA GAA GAG TTA AAT AAA AAA ATT GAA GAG GCT AAG AAC AAC TIT TTA AGT CTT CTC AAT TTA TTT TTT TAA CTT CTC CGA TTC TTG Leu Lys Asn Ser Glu Glu Leu Asn Lys Lys Ile Glu Glu Ala Lys Asn>

OspC-K48

400 430 390 CAT TCT GAA GCA TTT ACT AAT AGA CTA AAA GGT TCT CAT GCA CAA CTT GTA AGA CTT CGT AAA TGA TTA TCT GAT TTT CCA AGA GTA CGT GTT GAA His Ser Glu Ala Phe Thr Asn Arg Leu Lys Gly Ser His Ala Gln Leu> 460 470 450 440 GGA GTT GCT GCT ACT GAT GAT CAT GCA AAA GAA GCT ATT TTA AAG CCT CAA CGA CGA CGA TGA CTA CTA GTA CGT TTT CTT CGA TAA AAT TTC Gly Val Ala Ala Ala Thr Asp Asp His Ala Lys Glu Ala Ile Leu Lys> 490 500 510 ି ଅଟେ ହ TCA AAT CCT ACT AAA GAT AAG GGT GCT AAA GCA CTT AAA GAC TTA TCT AGT TTA GGA TGA TTT CTA TTC CCA CGA TTT CGT GAA TTT CTG AAT AGA Ser Asn Pro Thr Lys Asp Lys Gly Ala Lys Ala Leu Lys Asp Leu Ser> 550 560 570 530 540 GAA TCA GTA GAA AGC TTG GCA AAA GCA GCG CAA GAA GCA TTA GCT AAT CTT AGT CAT CTT TCG AAC CGT TTT CGT CGC GTT CTT CGT AAT CGA TTA Glu Ser Val Glu Ser Leu Ala Lys Ala Ala Gln Glu Ala Leu Ala Asn> 590 600 610 620 TCA GTT AAA GAA CTT ACA AAT CCT GTT GTG GCA GAA AGT CCA AAA AAA AGT CAA TTT CTT GAA TGT TTA GGA CAA CAC CGT CTT TCA GGT TTT TTT Ser Val Lys Glu Leu Thr Asn Pro Val Val Ala Glu Ser Pro Lys Lys> 630 CCT TAA GGA ATT Pro ***>

FIGURE 13 (2 of 2)

OspC-PKO

Sequence Range: 1 to 639

20 ATG AAA AAG AAT ACA TTA AGT GCG ATA TTA ATG ACT TTA TTT TTA TTT TAC TTT TTC TTA TGT AAT TCA CGC TAT AAT TAC TGA AAT AAA AAT AAA Met Lys Lys Asn Thr Leu Ser Ala Ile Leu Met Thr Leu Phe Leu Phe> 70 80 90 60 50 ATA TOT TGT AGT AAT TCA GGG AAA GGT GGG GAT TCT GCA TCT ACT AAT TAT AGA ACA TCA TTA AGT CCC TTT CCA CCC CTA AGA CGT AGA TGA TTA Ile Ser Cys Ser Asn Ser Gly Lys Gly Gly Asp Ser Ala Ser Thr Asn> 120 140 100 110 CCT GCT GAC GAG TCT GCG AAA GGG CCT AAT CTT ACA GAA ATA AGC AAA GGA CGA CTG CTC AGA CGC TTT CCC GGA TTA GAA TGT CTT TAT TCG TTT Pro Ala Asp Glu Ser Ala Lys Gly Pro Asn Leu Thr Glu Ile Ser Lys> 180 160 170 AAA ATT ACA GAT TCT AAT GCA TTT GTA CTT GCT GTT AAA GAA GTT GAG TTT TAA TGT CTA AGA TTA CGT AAA CAT GAA CGA CAA TTT CTT CAA CTC Lys Ile Thr Asp Ser Asn Ala Phe Val Leu Ala Val Lys Glu Val Glu> 220 230 240 210 200 ACT TTG GTT TTA TCT ATA GAT GAA CTT GCT AAG AAA GCT ATT GGT CAA TGA AAC CAA AAT AGA TAT CTA CTT GAA CGA TTC TTT CGA TAA CCA GTT Thr Leu Val Leu Ser Ile Asp Glu Leu Ala Lys Lys Ala Ile Gly Gln> 270 250 260 280 AAA ATA GAC AAT AAT AAT GGT TTA GCT GCT TTA AAT AAT CAG AAT GGA TTT TAT CTG TTA TTA CCA AAT CGA CGA AAT TTA TTA GTC TTA CCT Lys Ile Asp Asn Asn Asn Gly Leu Ala Ala Leu Asn Asn Gln Asn Gly> 290 300 310 320 330 TCG TTG TTA GCA GGA GCC TAT GCA ATA TCA ACC CTA ATA ACA GAA AAA AGC AAC AAT CGT CCT CGG ATA CGT TAT AGT TGG GAT TAT TGT CTT TTT Ser Leu Leu Ala Gly Ala Tyr Ala Ile Ser Thr Leu Ile Thr Glu Lys> 350 360 370 TTG AGT AAA TTG AAA AAT TTA GAA GAA TTA AAG ACA GAA ATT GCA AAG AAC TCA TTT AAC TTT TTA AAT CTT CTT AAT TTC TGT CTT TAA CGT TTC Leu Ser Lys Leu Lys Asn Leu Glu Glu Leu Lys Thr Glu Ile Ala Lys>

OspC-PKO

CCA AAA AAA CCT TAA GGT TTT TTT GGA ATT Pro Lys Lys Pro ***>

430 420 400 390 GCT AAG AAA TGT TCC GAA GAA TTT ACT AAT AAA CTA AAA AGT GGT CAT CGA TTC TTT ACA AGG CTT CTT AAA TGA TTA TTT GAT TTT TCA CCA GTA Ala Lys Lys Cys Ser Glu Glu Phe Thr Asn Lys Leu Lys Ser Gly His> 470 460 450 440 GCA GAT CTT GGC AAA CAG GAT GCT ACC GAT GAT CAT GCA AAA GCA GCT CGT CTA GAA CCG TTT GTC CTA CGA TGG CTA CTA GTA CGT TTT CGT CGA Ala Asp Leu Gly Lys Gln Asp Ala Thr Asp Asp His Ala Lys Ala Ala> 520 510 500 490 ATT TTA AAA ACA CAT GCA ACT ACC GAT AAA GGT GCT AAA GAA TTT AAA TAA AAT TIT TGT GTA CGT TGA TGG CTA TIT CCA CGA TIT CIT AAA TIT Ile Leu Lys Thr His Ala Thr Thr Asp Lys Gly Ala Lys Glu Phe Lys> 560 570 550 540 530 GAT TTA TTT GAA TCA GTA GAA GGT TTG TTA AAA GCA GCT CAA GTA GCA CTA AAT AAA CTT AGT CAT CTT CCA AAC AAT TTT CGT CGA GTT CAT CGT Asp Leu Phe Glu Ser Val Glu Gly Leu Leu Lys Ala Ala Gln Val Ala> 620 600 610 590 CTA ACT AAT TCA GTT AAA GAA CTT ACA AGT CCT GTT GTA GCA GAA AGT GAT TGA TTA AGT CAA TTT CTT GAA TGT TCA GGA CAA CAT CGT CTT TCA Leu Thr Asn Ser Val Lys Glu Leu Thr Ser Pro Val Val Ala Glu Ser> 630

FIGURE 14 (2 of 2)

OspC-TRO

Sequence Range: 1 to 624

20 ATG AAA AAG AAT ACA TTA AGT GCG ATA TTA ATG ACT TTA TTT TTA TTT TAC TIT TTC TTA TGT AAT TCA CGC TAT AAT TAC TGA AAT AAA AAT AAA Met Lys Lys Asn Thr Leu Ser Ala Ile Leu Met Thr Leu Phe Leu Phe> 70 60 ATA TCT TGT AAT AAT TCA GGT GGG GAT TCT GCA TCT ACT AAT CCT GAT TAT ÁGA ACA TTA TTA AGT CCA CCC CTA AGA CGT AGA TGA TTA GGA CTA Ile Ser Cys Asn Asn Ser Gly Gly Asp Ser Ala Ser Thr Asn Pro Asp> 120 130 110 100 GAG TCT GCA AAA GGA CCT AAT CTT ACC GTA ATA AGC AAA AAA ATT ACA CTC AGA CGT TTT CCT GGA TTA GAA TGG CAT TAT TCG TTT TTT TAA TGT Glu Ser Ala Lys Gly Pro Asn Leu Thr Val Ile Ser Lys Lys Ile Thr> 160 170 180 150 GAT TCT AAT GCA TTT TTA CTG GCT GTG AAA GAA GTT GAG GCT TTG CTT CTA AGA TTA CGT AAA AAT GAC CGA CAC TTT CTT CAA CTC CGA AAC GAA Asp Ser Asn Ala Phe Leu Leu Ala Val Lys Glu Val Glu Ala Leu Leu> 220 230 210 TCA TCT ATA GAT GAA CTT TCT AAA GCT ATT GGT AAA AAA ATA AAA AAT AGT AGA TAT CTA CTT GAA AGA TTT CGA TAA CCA TTT TTT TAT TTT TTA Ser Ser Ile Asp Glu Leu Ser Lys Ala Ile Gly Lys Lys Ile Lys Asn> 270 250 260 280 GAT GGT ACT TTA GAT AAC GAA GCA AAT CGA AAC GAA TCA TTG ATA GCA CTA CCA TGA AAT CTA TTG CTT CGT TTA GCT TTG CTT AGT AAC TAT CGT Asp Gly Thr Leu Asp Asn Glu Ala Asn Arg Asn Glu Ser Leu Ile Ala> 310 320 330 290 300 GGA GCT TAT GAA ATA TCA AAA CTA ATA ACA CAA AAA TTA AGT GTA TTG CCT CGA ATA CTT TAT AGT TTT GAT TAT TGT GTT TTT AAT TCA CAT AAC Gly Ala Tyr Glu Ile Ser Lys Leu Ile Thr Gln Lys Leu Ser Val Leu> 350 360 370 380 AAT TCA GAA GAA TTA AAG AAA AAA ATT AAA GAG GCT AAG GAT TGT TCC TTA AGT CTT CTT AAT TTC TTT TTT TAA TTT CTC CGA TTC CTA ACA AGG Asn Ser Glu Glu Leu Lys Lys Lys Ile Lys Glu Ala Lys Asp Cys Ser>

OSPC-TRO

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FIGURE 15 (2 of 2)

PCT/US94/12352

28/133

P93

まならりへいつ

Sequence Range: 1 to 2102

30 20 10 ATG AAA AAA ATG TTA CTA ATC TTT AGT TTT TTT CTT ATT TTC TTG AAT TAC TIT TIT TAC AAT GAT TAG AAA TCA AAA AAA GAA TAA AAG AAC TTA Met Lys Lys Met Leu Leu Ile Phe Ser Phe Phe Leu Ile Phe Leu Asn> 80 70 60 GGA TTT CCT GTT AGT GCA AGA GAA GTT GAT AGG GAA AAA TTA AAG GAC CCT AAA GGA CAA TCA CGT TCT CTT CAA CTA TGG-CTT TTT AAT TTC CTG Gly Phe Pro Val Ser Ala Arg Glu Val Asp Arg Glu Lys Leu Lys Asp> 130 120 110 100 TTT GTT AAT ATG GAT CTT GAG TTT GTA AAT TAT AAA GGC CCT TAT GAT AAA CAA TTA TAC CTA GAA CTC 2 A CAT TTA ATA TTT CCG GGA ATA CTA Phe Val Asn Met Asp Leu Glu Phe Val Asn Tyr Lys Gly Pro Tyr Asp> 180 170 160 150 TCT ACA AAT ACA TAT GAA CAA ATA GTG GGT ATT GGG GAG TTT TTA GCA AGA TGT TTA TGT ATA CTT GTT TAT CAC CCA TAA CCC CTC AAA AAT CGT Ser Thr Asn Thr Tyr Glu Gln Ile Val Gly Ile Gly Glu Phe Leu Ala> 230 220 210 200 AGA CCG TTG ACC AAT TCC AAT AGC AAC TCA AGT TAT TAT GGT AAA TAT TCT GGC AAC TGG TTA AGG TTA TCG TTG AGT TCA ATA ATA CCA TTT ATA Arg Pro Leu Thr Asn Ser Asn Ser Asn Ser Ser Tyr Tyr Gly Lys Tyr> 280 270 260 TTT ATT AAT AGA TTT ATT GAT GAT CAA GAT AAA AAA GCA AGC GTT GAT AAA TAA TTA TCT AAA TAA CTA CTA GTT CTA TTT TTT CGT TCG CAA CTA Phe Ile Asn Arg Phe Ile Asp Asp Gln Asp Lys Lys Ala Ser Val Asp> 320 330 300 310 290 GTT TTT TCT ATT GGT AGT AAG TCA GAG CTT GAC AGT ATA TTG AAT TTA CAA AAA AGA TAA CCA TCA TTC AGT CTC GAA CTG TCA TAT AAC TTA AAT Val Phe Ser Ile Gly Ser Lys Ser Glu Leu Asp Ser Ile Leu Asn Leu> 370 360 350 340

FIGURE 16 (1 of 5)

AGA AGA ATT CTT ACA GGG TAT TTA ATA AAG TCT TTC GAT TAT GAC AGG TCT TCT TAA GAA TGT CCC ATA AAT TAT TTC AGA AAG CTA ATA CTG TCC Arg Arg Ile Leu Thr Gly Tyr Leu Ile Lys Ser Phe Asp Tyr Asp Arg>

390 400 420 430 TCT AGT GCA GAA TTA ATT GCT AAG GTT ATT ACA ATA TAT AAT GCT GTT AGA TCA CGT CTT AAT TAA CGA TTC CAA TAA TGT TAT ATA TTA CGA CAA Ser Ser Ala Glu Leu Ile Ala Lys Val Ile Thr Ile Tyr Asn Ala Val> 440 450 460 470 TAT AGA GGA GAT TTG GAT TAT TAT AAA GGG TTT TAT ATT GAG GCT GCT ATA TCT CCT CTA AAC CTA ATA ATA TTT CCC AAA ATA TAA CTC CGA CGA Tyr Arg Gly Asp Leu Asp Tyr Tyr Lys Gly Phe Tyr Ile Glu Ala Ala> 490 500 510 · = = • • TTA AAG TCT TTA AGT AAA GAA AAT GCA GGT CTT TCT AGG GTT TAT AGT AAT TTC AGA AAT TCA TTT CTT TTA CGT CCA GAA AGA TCC CAA ATA TCA Leu Lys Ser Leu Ser Lys Glu Asn Ala Gly Leu Ser Arg Val Tyr Ser> 530 540 550 560 570 CAG TGG GCT GGA AAG ACA CAA ATA TTT ATT CCT CTT AAA AAG GAT ATT GTC ACC CGA CCT TTC TGT GTT TAT AAA TAA GGA GAA TTT TTC CTA TAA Gln Trp Ala Gly Lys Thr Gln Ile Phe Ile Pro Leu Lys Lys Asp Ile> 580 590 600 610 620 TTG TCT GGA AAT ATT GAG TCT GAC ATT GAT ATT GAC AGT TTA GTT ATA AAC AGA CCT TTA TAA CTC AGA CTG TAA CTA TAA CTG TCA AAT CAA TGT Leu Ser Gly Asn Ile Glu Ser Asp Ile Asp Ile Asp Ser Leu Val Thr> 630 640 650 €€0 GAT AAG GTG GTG GCA GCT CTT TTA AGT GAA AAT GAA GCA GGT GTT AAC CTA TTC CAC CAC CGT CGA GAA AAT TCA CTT TTA CTT CGT CCA CAA TIG Asp Lys Val Val Ala Ala Leu Leu Ser Glu Asn Glu Ala Gly Val Asn> 680 690 700 710 TTT GCA AGA GAT ATT ACA GAT ATT CAA GGC GAA ACT CAT AAG GCA GAT AAA CGT TCT CTA TAA TGT CTA TAA GTT CCG CTT TGA GTA TTC CGT CTA Phe Ala Arg Asp Ile Thr Asp Ile Gln Gly Glu Thr His Lys Ala Asp> 730 740 750 760 CAA GAT AAA ATT GAT ATT GAA TTA GAC AAT ATT CAT GAA AGT GAT TCC GTT CTA TTT TAA CTA TAA CTT AAT CTG TTA TAA GTA CTT TCA CTA AGG Gln Asp Lys Ile Asp Ile Glu Leu Asp Asn Ile His Glu Ser Asp Ser> 770 780 790 800 810 AAT ATA ACA GAA ACT ATT GAA AAT TTA AGG GAT CAG CTT GAA AAA GCT TTA TAT TGT CTT TGA TAA CTT TTA AAT TCC CTA GTC GAA CTT TTT CGA Asn Ile Thr Glu Thr Ile Glu Asn Leu Arg Asp Gln Leu Glu Lys Ala>

FIGURE 16 (2 of 5)

830 820 850 860 ACA GAT GAA GAG CAT AAA AAA GAG ATT GAA AGT CAG GTT GAT GCT AAA TGT CTA CTT CTC GTA TTT TTT CTC TAA CTT TCA GTC CAA CTA CGA TTT Thr Asp Glu Glu His Lys Lys Glu Ile Glu Ser Gln Val Asp Ala Lys> 870 880 890 900 AAG AAA CAA AAG GAA GAG CTA GAT AAA AAG GCA ATA AAT CTT GAT AAA TTC TTT GTT TTC CTT CTC GAT CTA TTT TTC CGT TAT TTA GAA CTA TTT Lys Lys Gln Lys Glu Glu Leu Asp Lys Lys Ala Ile Asn Leu Asp Lys> 940 920 930 950 and the second GCT CAG CAA AAA TTA GAT TCT GCT GAA GAT AAT TTA GAT GTT CAA AGA CGA GTC GTT TTT AAT CTA AGA CGA CTT CTA TTA AAT CTA CAA GTT TCT Ala Gln Gln Lys Leu Asp Ser Ala Glu Asp Asn Leu Asp Val Gln Arg> 970 980 990 AAT ACT GTT AGA GAG AAA ATT CAA GAG GAT ATT AAC GAA ATT AAC AAG TTA TGA CAA TCT CTC TTT TAA GTT CTC CTA TAA TTG CTT TAA TTG TTC Asn Thr Val Arg Glu Lys Ile Gln Glu Asp Ile Asn Glu Ile Asn Lys> 1010 1020 1030 1040 1050 GAA AAG AAT TTA CCA AAG CCT GGT GAT GTA AGT TCT CCT AAA GTT GAT CTT TTC TTA AAT GGT TTC GGA CCA CTA CAT TCA AGA GGA TTT CAA CTA Glu Lys Asn Leu Pro Lys Pro Gly Asp Val Ser Ser Pro Lys Val Asp> 1060 1070 1080 1090 AAG CAA CTA CAA ATA AAA GAG AGC CTG GAA GAT TTG CAG GAG CAG CTT TTC GTT GAT GTT TAT TTT CTC TCG GAC CTT CTA AAC GTC CTC GTC GAA Lys Gln Leu Gln Ile Lys Glu Ser Leu Glu Asp Leu Gln Glu Gln Leu> 1130 1110 1120 1140 AAA GAA ACT GGT GAT GAA AAT CAG AAA AGA GAA ATT GAA AAG CAA ATT TTT CTT TGA CCA CTA CTT TTA GTC TTT TCT CTT TAA CTT TTC GTT TAA Lys Glu Thr Gly Asp Glu Asn Gln Lys Arg Glu Ile Glu Lys Gln Ile> 1160 1170 1180 1190 GAA ATC AAA AAA AGT GAT GAA AAG CTT TTA AAA AGT AAA GAT GAT AAA CTT TAG TTT TCA CTA CTT TTC GAA AAT TTT TCA TTT CTA CTA TTT Glu Ile Lys Lys Ser Asp Glu Lys Leu Leu Lys Ser Lys Asp Asp Lys> 1220 1230 GCA AGT AAA GAT GGT AAA GCC TTG GAT CTT GAT CGA GAA TTA AAT TCT CGT TCA TTT CTA CCA TTT CGG AAC CTA GAA CTA GCT CTT AAT TTA AGA Ala Ser Lys Asp Gly Lys Ala Leu Asp Leu Asp Arg Glu Leu Asn Ser>

FIGURE 16 (3 of 5)

WO 95/12676 PCT/US94/12352

31/133

1260 1270 1260 1250 AAA GCT TCT AGC AAA GAA AAA AGT AAA GCC AAG GAA GAA ATA ACC TTT CGA AGA TCG TTT CTT TTT TCA TTT CGG TTC CTT CTT TAT TGG Lys Ala Ser Ser Lys Glu Lys Ser Lys Ala Lys Glu Glu Glu Ile Thr> 1330 1320 1310 1300 AAG GGT AAG TCA CAG AAA AGC TTA GGC GAT TTG AAT AAT GAT GAA AAT TTC CCA TTC AGT GTC TTT TCG AAT CCG CTA AAC TTA TTA CTA CTT TTA Lys Gly Lys Ser Gln Lys Ser Leu Gly Asp Leu Asn Asn Asp Glu Asn> 1370 - ---1380 1360 1350 CTT ATG ATG CCA GAA GAT CAA AAA TTA CCT GAG GTT AAA AAA TTA GAT GAA TAC TAC GGT CTT CTA GTT TTT AAT GGA CTC CAA TTT TTT AAT CTA Leu Met Met Pro Glu Asp Gln Lys Leu Pro Glu Val Lys Lys Leu Asp> 1430 1420 1410 1400 AGC AAA AAA GAA TTT AAA CCT GTT TCT GAG GTT GAG AAA TTA GAT AAG TCG TTT TTT CTT AAA TTT GGA CAA AGA CTC CAA CTC TTT AAT CTA TTC Ser Lys Lys Glu Phe Lys Pro Val Ser Glu Val Glu Lys Leu Asp Lys> 1480 1470 1460 1450 ATT TTC AAG TCT AAT AAC AAT GTT GGA GAA TTA TCA CCG TTA GAT AAA TAA AAG TTC AGA TTA TTG TTA CAA CCT CTT AAT AGT GGC AAT CTA TTT Ile Phe Lys Ser Asn Asn Asn Val Gly Glu Leu Ser Pro Leu Asp Lys> 1520 1510 - 1500 1490 TCT TCT TAT AAA GAC ATT GAT TCA AAA GAG GAG ACA GTT AAT AAA GAT AGA AGA ATA TTT CTG TAA CTA AGT TTT CTC CTC TGT CAA TTA TTT CTA Ser Ser Tyr Lys Asp Ile Asp Ser Lys Glu Glu Thr Val Asn Lys Asp> 1560 1570 1550 1540 • • GTT AAT TTG CAA AAG ACT AAG CCT CAG GTT AAA GAC CAA GTT ACT TCT CAA TTA AAC GTT TTC TGA TTC GGA GTC CAA TTT CTG GTT CAA TGA AGA Val Asn Leu Gln Lys Thr Lys Pro Gln Val Lys Asp Gln Val Thr Ser> 1610 1620 1600 1590 TTG AAT GAA GAT TTG ACT ACT ATG TCT ATA GAT TCC AGT AGT CCT GTA AAC TTA CTT CTA AAC TGA TGA TAC AGA TAT CTA AGG TCA TCA GGA CAT Leu Asn Glu Asp Leu Thr Thr Met Ser Ile Asp Ser Ser Ser Pro Val> 1660 1670 1650 - 1640 TTT TTA GAG GTT ATT GAT CCA ATT ACA AAT TTA GGA ACT CTT CAA CTT AAA AAT CTC CAA TAA CTA GGT TAA TGT TTA AAT CCT TGA GAA GTT GAA Phe Leu Glu Val Ile Asp Pro Ile Thr Asn Leu Gly Thr Leu Gln Leu>

FIGURE 16 (4 of 5)

1720 1710 1700 1690 ATT GAT TTA AAT ACT GGT GTT AGG CTT AAA GAA AGC ACT CAG CAA GGC TAA CTA AAT TTA TGA CCA CAA TCC GAA TTT CTT TCG TGA GTC GTT CCG Ile Asp Leu Asn Thr Gly Val Arg Leu Lys Glu Ser Thr Gln Gln Gly> 1750 1760 1740 1730 ATT CAG CGG TAT GGA ATT TAT GAA CGT GAA AAA GAT TTG GTT GTT ATT TAA GTC GCC ATA CCT TAA ATA CTT GCA CTT TTT CTA AAC CAA CAA TAA Ile Gln Arg Tyr Gly Ile Tyr Glu Arg Glu Lys Asp Leu Val Val Ile> 1800 . _1810 1790 1780 AAA ATG GAT TCA GGA AAA GCT AAG CTT CAG ATA CTT GAT AAA CTT GAA TTT TAC CTA AGT CCT TTT CGA TTC GAA GTC TAT GAA CTA TTT GAA CTT Lys Met Asp Ser Gly Lys Ala Lys Leu Gln Ile Leu Asp Lys Leu Glu> 1860 1850 1840 1830 AAT TTA AAA GTG GTA TCA GAG TCT AAT TTT GAG ATT AAT AAA AAT TCA TTA AAT TTT CAC CAT AGT CTC AGA TTA AAA CTC TAA TTA TTT TTA AGT Asn Leu Lys Val Val Ser Glu Ser Asn Phe Glu Ile Asn Lys Asn Ser> 1920 1910 1900 1890 1880 TCT CTT TAT GTT GAT TCT AAA ATG ATT TTA GTA GCT GTT AGG GAT AAA AGA GAA ATA CAA CTA AGA TTT TAC TAA AAT CAT CGA CAA TCC CTA TTT Ser Leu Tyr Val Asp Ser Lys Met Ile Leu Val Ala Val Arg Asp Lys> 1960 1940 1950 1930 GAT AGT AGT AAT GAT TGG AGA TTG GCC AAA TTT TCT CCT AAA AAT TTA CTA TCA TCA TTA CTA ACC TCT AAC CGG TTT AAA AGA GGA TTT TTA AAT Asp Ser Ser Asn Asp Trp Arg Leu Ala Lys Phe Ser Pro Lys Asn Leu> 2000 1990 1970 1980 GAT GAG TTT ATT CTT TCA GAG AAT AAA ATT ATG CCT TTT ACT AGC TTT CTA CTC AAA TAA GAA AGT CTC TTA TTT TAA TAC GGA AAA TGA TCG AAA Asp Glu Phe Ile Leu Ser Glu Asn Lys Ile Met Pro Phe Thr Ser Phe> 2050 2040 2030 2020 TCT GTG AGA AAA AAT TTT ATT TAT TTG CAA GAT GAG TTT AAA AGT CTA AGA CAC TCT TTT TTA AAA TAA ATA AAC GTT CTA CTC AAA TTT TCA GAT Ser Val Arg Lys Asn Phe Ile Tyr Leu Gln Asp Glu Phe Lys Ser Leu> 2090 2080 2070 GTT ATT TTA GAT GTA AAT ACT TTA AAA AAA GTT AAG TA CAA TAA AAT CTA CAT TTA TGA AAT TTT TTT CAA TTC AT Val Ile Leu Asp Val Asn Thr Leu Lys Lys Val Lys Xxx>

FIGURE 16 (5 of 5)

SASOUCID- SINO DE SOLTIGAS

p93 - K48

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1	ATGAAAAAAT	TGTTACTAAT	CTTTAGTTTT	TTTCTTATTT	CTTTGAATGG	ATTICCICIT
61	DODG ATTERAC	AACTTGATAA	GGAAAAATTA	AAGGATTTTG	TTAATATGGA	TCTTGAGTTT
121	CTALACTATA	AAGGTCCTTA	TGATTCTACA	AATACATATG	AACAAATAGT	AGSTATIGGT
121	CACALALALASC	CAAGACCATT	GATTAATTCC	AATAGCAACT	CAATTTATTA	TGGTAAATAT
241	ATAATTATTT	GATTTATTGA	TGATCAAGAT	AAAAAAGCAA	GCGTTGATGT	TTTTTCTATT
301	CCTACTACCT	CACAGCTTGA	CAGTATATTG	AATCTAAGAA	GAATTCTTAC	AGGGTATTIG
361	איזי ארבוע מערע מידע	TTGATTATGA	AAGATCTAGT	GCTGAATTAA	TTGCTAAGGT	TATTACAATA
421	CATAATCCTC	TTTATAGAGG	GGATTTAAAT	TATTATAAAG	AGGTTTATAT	TGAGGCTGCT
481	Tele-ALDIA & VALAL	TAACTAAAGA	AAATGCAGGT	CTTTCTAGAG	TGTACAGTCA	ATGGGCTGGA
541	AAGACACAAA	TATTTATTCC	TCTTAAAAAG	AATATTTTAT	CTGGAAAAGT	TGAGTCTGAC
601	ATTGATATTG	ACAGTTTGGT	TACAGATAAG	GTTGTGGCAG	-CTCTTTTAAG	CGAGAATGAA
661	CCACCTCTTA	ACTITICCAAG	AGATATTACA	GATATTCAAG	GCGAAACTCA	TAAAGCAGAT
721	CARGATAAAA	TTGATATTGA	ATTAGATAAT	GTTCATAAAA	GTGATTCCAA	TATAACAGAG
781	ACTATTCAGA	ATTTAAGAGA	TCAGCTTGAA	AAGGCTACAG	ATGAAGAGCA	TAGAAAAGAG
841	ATTGAAAGTC	AGGTTGATGC	TAAAAAGAAA	CAAAAAGAAG	AACTAGATAA	AAAGGCAATC
901	GATCTTGATA	AAGCCCAACA	AAAATTAGAT	TCTTCTGAAG	ATAATTTAGA	TATTCAAAGG
961	GATACTGTTA	GAGAGAAGAT	TCAAGAGGAT	ATTGACGAGA	TTAATAAAGA	AAAGAATTTG
1021	CCAAAACCTG	GTGATGTAAG	TTCTCCTAAA	GTTGATAAGC	AGCTACAAAT	AAAAGAGAGT
1081	CTAGAAGACT	TGCAGGAACA	GCTTAAAGAA	ACTAGCGATG	AAAATCAAAA	AAGAGAAATT
1141	GAAAAGCAAA	TTGAAATCAA	AAAAAGTGAT	GAAGAACTTT	TAAAAAGTAA	AGATCCTAAA
1201	GCATTAGATC	TTAATGGAGA	TTTAAATTCT	AAAGTTTCTA	GTAAAGAAAA	AATTAAAGGC
1261	AAAGAAGGAG	AAATAGTCAA	AGAGGAATCA	AAGGCAAGTT	TAGCTGATTT	GAATAATGAC
1321	GAAAATCITA	TGAGGCCGGA	AGATCAAAAA	TTATCTGAGG	TTAAAAAATT	AGATAGTAAA
1381	AAAAATTTAA	AACCIGITIC	TGAGATTGAG	AGAGTAAATG	AAATTTCGAA	GTCTAACAAC
1441	AATGAGATTA	GTGAATCATC	ACCATTATAT	AAGCCTTCTT	ATAGCGATAT	GGATTCAAAA
1501	GAGGGTATAG	ATAATAAAGA	TGTTAACTTG	CAAGAAACCA	AGTCTCAAAC	TAAAAGTCAA
1561	CCTACTTCTT	TAAATCAAGA	TTTGACTACT	ATGTCTATAG	ATTCTAGTAA	TCCTGTATTT
1621	TTAGAGGTTA	TTGATCCTAT	' TACAAATTTA	GGAACGCTTC	AACITATIGA	TITGAATACC
1681	GGTGTTAGAC	TTAAAGAAAG	TACTCAGCAA	GCATTCAGC	GGTATGGAAT	TTATGAACGT-
1741	GAAAAAGATT	TAGTTGTTAI	TAAAATGGAT	TCAGGAAAAG	CCAAGCTTCA	AATACITAAT
1801	AAACTTGAGA	. ATTTAAAAGI	GATATCGGAG	TCTAATTTTG	AGATTAATAA	AAATTCATCT
1861	CTTTATGTTG	ACTCTAAAAT	GATTTTAGTA	GTTGTGAGAG	ATAGIGGTAA	TGTTTGGAGA
1921	TTGGCTAAAT	TITCICCIA	TAKATTTAAA 1	GAGTTTATTC	TTTCAGAGAA	TAAAATTTTG
1981	CCTTTTACTA	GCTTTTCTGT	GAGAAAGAAT	TTATTTATT	TGCAGGATGA	GTTTAAAAGT
2041	CTTATTACTI	`TAGATGTAAA	A TACTITAAAA	AAAGTTAAGT	A	

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1	ATGAAAAAA	TGTTACTAAT	CTTTAGTTTT	TTTCTTGTTT	TTTTAAATGG	ATTICCTCTT
61	AATGCAAGGG	AAGTTGATAA	GGAAAAATTA	AAGGACTTTG	TTAATATGGA	TCTTGAATTT
121	GTTAATTACA	AGGGTCCTTA	TGATTCTACA	GATACATATG	AACAAATAGT	AGGTATTGGG
181	GAGTTTTTAG	CAAGGCCGTT	GAACAATTCC	AATAGTAATT	CAAGTTATTA	TGGTAAATAT
241	TTTGTTAATA	GATTTATTGA	CGATCAAGAT	AAAAAAGCAA	GTGTTGATAT	TTTTTCTATT
				AATCTAAGAA		
				GCGGAATTAA		
				TATTACAAAG		
				CTTTCTAGGG		
541	AAGACACAAA	TATTTATTCC	TCTTAAAAAG	TATTTTATAA	CTGGAAATGT	TGAGTCTGAC
601	ATTGATATTG	ATAGTTTGGT	TACAGATAAG	GTGGTGGÇAG	CTCTTTTAAG	TGAGAATGAA
661	TCAGGTGTTA	ACTITICAAG	AGATATTACA	GACATTCAAG	GCGAAACTCA	TAAAGCAGAT
721	CAAGATAAAA	TTGATATTGA	ATTAGATAAT	TTTCATGAAA	GTGATTCCAA	TATAACAGAA
				AAAGCTACAG		
				CAAAAGGAAG		
				TTTGCTGAAG		
				ATTAACGAGA		
				GTTGATAAGC		
				GCTAGTGATG		
1141	GAAAAGCAAA	TTGAAATCAA	AAAAAATGAT	GAAGAACTTT	TTAAAAATAA	AGATCATAAA
				AAAGCTTCTA		
				AAAAATTTAG		
				AATGAGGTTA		
				GAGGGTGTAG		
				CCTACTTCGT		
1501	GTGTCTATAG	ATTCCAGTAA	TCCTGTCTTT	TTAGAGGTTA	TCGATCCGAT	TACAAATTTA
				GGTGTTAGAC		
				GAAAAAGATT		
				AAACTCGAGA		
				CTTTATGTTG		
				TIGGCTAAAT		
				CCTTTTACTA		
1921	TTTATTTATT	TGCAAGATGA	ACTTAAAAGC	TTAGTTACTT	TAGATGTAAA	TACTTTAAAA
1981	AAAGTTAAGT	A				

p93 - pIRO

	ATGAAAAAA					
	AATGCAAGGG					
	GTAAACTATA					
	GAGTTTTTAG					
	TTTATTAATA					
	AGTAGTAAGT					
361	ATAAAGTCTT	TTGATTATGA	AAGATCTAGT	GCTGAATTAA	TTGCCAAGGT	TATTACAATA
421	CATAATGCTG	TTTATAGAGG	TGATTTAAAT	TATTATAAAG	AGTTTTATAT	TGAGTCTGCT
481	TTAAAGTCTT	TAACTAAAGA	AAATGCAGGT	CTTTCTAGAG	TGTACAGTCA	ATGGGCTGGA
541	AAGACACAAA	TATTTATTCC	TCTTAAAAAG	AATATTTTAT	CTGGAAAAAT	TGAGTCTGAC
601	ATTGATATTG	ATACTTTCCT	TACAGATAAG	GTTGTGGCAG	CTCTTTTAAG	CGAAAATGAA
661	GCAGGTGTTA	ACTITICAAG	GGATATTACA	GATATTCAAG	GAGAAACTCA	TAAAGCAGAT
721	CAAGATAAAA	TTGATATTGA	ATTAGATAAT	GTTCATGAAA	GTGATTCCAA	TATAACAGAA
781	ACTATTGAGA	ATTTAAGAGA	TCAGCTTGAA	AAGGCTACAG	ATGAAGAGCA	TAGAAAAGAG
841	ATTGAAAGTC	AAGTTGATGC	TAAAAAGAAA	CAAAAAGAAG	AACTAGATAA	AAAGGCAATC
901	GATCTTGATA	AAGCCCAACA	AAAATTAGAT	TTTTCTGAAG	ATAATTTAGA	TATTCAAAGG
961	GATACTGTTA	GAGAGAAGAT	TCAAGAGGAT	ATTAACGAGA	TTAATAAGGA	AAAGAATTTA
1021	CCAAAACCTG	GTGATGTAAG	TTCTCCTAAA	GTTGATAAGC	AGCTACAAAT	AAAAGAGAGT
1081	CTAGAAGACT	TGCAGGAGCA	GCTTAAAGAA	ACTAGCGATG	AAAATCAAAA	AAGAGAAATT
1141	GAAAAGCAAA	TTGAAATCAA	AAAAAGTGAT	GAAGAACTTT	TAAAAAGCAA	AGATCCTAAA
1201	GCATTAGATC	TTAATCGAGA	TTTAAATTCT	AAAGCTTCTA	GTAAAGAAAA	AATTAAAGGC
1261	AAAGAAAAAG	AAATAGTCAA	AGAGAAATCA	AAGGTAAGTT	TAGGTGATTT	GGATAATGAC
1321	GAAACCCTTA	TGACGCCGGA	AGATCAAAAA	TTATCTGAGG	TTAAAAAATT	AGATAGTAAA
1381	AAAAATTTAA	AACCTGTTTC	TGAGATTGAG	AGAGTAAATG	AAATTTCAAA	GTCTAACAAC
1441	AATGAGGTTA	GCAAATCATC	ACCATTAGAT	AAGCCTTCTT	ATAGTGATAT	CGATTCAAAA
1501	GAGGTTGTAG	ATAATAAAGA	TGTTAATTTG	CAAGAAACCA	AGCCTCAAGC	TAAAAGTCAA
1561	TCTACTTCTT	TAAATCAAGA	TTTGATTACT	ATGTCTATAG	ATTCTAGTAA	TCCTGTATTT
1621	TTAGAGGTTA	TTGATCCTAT	TACAAATTTA	GGAATGCTTC	AACTTATTGA	TTTAAATACT
1681	GGTGTTAGAC	TTAAAGAAAG	CACTCAGCAA	GGCATTCAGC	GTTATGGAAT	TTATGAACGT
1741	GAAAAAGATT	TAGTTGTTAT	TAAAATGGAT	TCAGGAAAAG	CTAAGCTTCA	AATACTTAAT!
1801	AAACTTGAGA	ATTTAAAAGT	GATATCAGAG	TCTAATTTTG	AGATTAATAA	AAATTCATCT."
1861	CTTTATGTTG	ACTCTAAAAT	GATTTTAGTA	GCTGTGAAAG	ATAGTGGTAA	TGTTTGGAGA
1921	TTGGCTAAAT	TTTCTCCTAA	AAATTTAGAT	GAGTTTATTC	TTTCAGAGAA	TAAAATTTTY
1981	CCTTTTACTA	GCTTTTCTGT	GAGAAAGAAT	TITATTTATT	TGCAAGATGA	GTTTAAAAGT
2041	CTTATTACTT	TAGATGTAAA	TACTTTAAAA	AAAGTTAAGT	A	

p93 - pGau

1	ATGAAAAAA	TGTTACTAAT	CTTTAGTTTT	TITCTIGITI	TTTTAAATGG	ATTTCCTCTT
61	AATGCAAGGG	AAGTTGATAA	GGAAAAATTA	AAGGACTTTG	TTAATATGGA	TCTTGAATTT
121	GTTAATTACA	AGGGTCCTTA	TGATTCTACA	AATACATATG	AACAAATAGT	AGGTATTGGG
181	GAGTTTTTAG	CAAGGCCGTT	GATCAATTCC	AATAGTAATT	CAAGTTATTA	TGGTAAATAT
241	TTTGTTAATA	GATTTATTGA	CGATCAAGAT	AAAAAAGCAA	GTGTTGATAT	TTTTTCTATT
301	GGTAGTAAGT	CAGAGCTTGA	TAGTATATTA	AATCTAAGAA	GAATTCTTAC	AGGGTATTTA
361	ATGAAGTCTT	TTGATTATGA	GAGGTCTAGT	GCGGAATTAA	TTGCTAAAGC	TATTACAATA
421	TATAATGCTG	TTTATAGAGG	AGATTTAGAT	TATTACAAAG	AGTTTTATAT	TGAGGCTTCT
481	TTGAAGTCTT	TGACTAAAGA	AAATGCAGGT	CTTTCTAGGG	TGTACAGTCA	ATGGGCTGGG
				TATTTTTAT		
601	ATTGATATTG	ATAGTTTGGT	TACAGATAAG	GTGGTGGÇAG	CICTITIAAG	TGAGAATGAA
661	TCAGGTGTTA	ACTITICAAG	AGATATTACA	GACATTCAAG	GCGAAACTCA	TAAAGCAGAT
721	CAAGATAAAA	TTGATATTGA	ATTAGATAAT	ATTCATGAAA	GTGATTCCAA	TATAACAGAA
				AAAGCTACAG		
				CAAAAGGAAG		
				TTTGCTGAAG		
961	GATACTGTTA	GAGAGAAGCT	TCAAGAGAAT	ATTAACGAGA	CTAATAAGGA	AAAGAATTTA
				GTTGATAAGC		
				ACTGGTGATG		
				GAAAAGCTTT		
				GATCGAGAAT		
				ATAACCAAGG		
				ATGCCAGAAG		
				CCTGTTTCTG		
				TTATCACCGT		
				AAAGATGITA		
				GAAGATTTGA		
				CCAATTACAA		
				GAAAGCACTC		
				GTTATTAAAA		
				AAAGTGGTAT		
				AAAATGATTT		
				TTTTCTCCTA		
				AGCTTTTCTG		
		AGTTTAAAAG	TCTAGTTATT	TTAGATGTAA	ATACTITAAA	AAAAGTTAAG
2101	TAAAGCC		•			

p93 - pKO

1	ATGAAAAAA	TGTTACTAAT	CTTTAGTTTT	TTTCTTGTTT	TTTTAAATGG	ATTTCCTCTT
£1	A ATTOCA ACCC	AACTICATAA	GGAAAAATTA	AAGGACTTIG	TTAATATGGA	TCTTGAATIT
727	CTTTA ATTTACA	VCCCLCCLLLY.	TGATTCTACA	AATACATATG	AACAAATAGT	AGGIATIGGG
181	CyCatalalalable	CAAGGCCGTT	GATCAATTCC	AATAGTAATT	CAAGITATTA	TGGTAAATAT
2/1	The Thirthire	CATTTATTGA	CGATCAAGAT	AAAAAAGCAA	GIGITGATAT	TTTTTCTATT
301	CCTACTAACT	CAGAGCTTGA	TAGTATATTA	AATCTAAGAA	GAATTCTTAC	AGGGTATTTA
361	אוריאס ע כיארע	TTCATTATCA	GAGGTCTAGT	GCGGAATTAA	TTGCTAAAGC	TATTACAATA
427	TATALITATION	TTTATAGAGG	AGATTTAGAT	TATTACAAAG	AGTTTTATAT	TGAGGCTTCT
491	JALOUS VESTAL	TGACTAAAGA	AAATGCAGGT	CTTTCTAGGG	TGTACAGTCA	ATGGGCTGGG
541	AACACACAAA	TATTTATTCC	TCTTAAAAAG	TATTTTTAT	CTGGAAATGT	TGAGTCTGAC
601	STREATESTEE	ATACTTTCCT	TACAGATAAG	GTGGTGGCAG	CTCTTTTAAG	TGAGAATGAA
661	ALL SALES COLLEGE	ACTITICAAG	AGATATTACA	GACATTCAAG	GCGAAACTCA	TAAAGCAGAT
721	CANCATARAR	TTGATATTGA	ATTAGATAAT	TTTCATGAAA	GTGATTCCAA	TATAACAGAA
721	ACTEMPTERCA	ATTTAACCCA	TCAGCTTGAA	AAAGCTACAG	ATGAAGAGCA	TAAAAAAGAG
841	ATTICADACTY	AGGTTGATGC	TAAAAAGAAA	CAAAAGGAAG	AATTAGATAA	AAAGGCAATT
901	CATALACTA A	AAGCTCAACA	AAAATTAGAT	TTTGCTGAAG	ATAATCTAGA	TATTCAAAGG
961	ATTOTTO ATTA	GAGAGAAGCT	TCAAGAAAAT	ATTAACGAGA	CTAATAAGGA	AAAGAATTTA
1021	CCAAAGCCTG	GTGATGTAAG	TTCTCCTAAG	GTTGATAAGC	AGTTGCAGAT	AAAAGAGAGT
1081	CTAGAAGATT	TGCAAGAGCA	GCTTAAAGAA	GCTAGTGATG	AAAATCAAAA	AAGAGAAATA
1141	CAAAAGCAAA	TTGAAATCAA	AAAAAATGAT	GAAGAACTTT	TTAAAAATAA	AGATCATAAA
1201	GCATTAGATC	TTAAGCAAGA	ATTAAATTCT	AAAGCTTCTA	GTAAAGAAAA	AATTGAAGGC
1261	GAAGAAGAGG	ATAAAGAATT	AGATAGTAAA	AAAAATTTAG	AGCCTGTTTC	TGAGGCTGAT
1321	AAAGTAGATA	AAATTTCCAA	GTCTAACAAC	AATGAGGTTA	GTAAATTATC	CCCGTTAGAT
1381	GAGCCTTCTT	ATAGCGACAT	TGATTCGAAA	GAGGGTGTAG	ATAACAAAGA	TGTTGATTTG
1441	CAAAAAACTA	AACCCCAAGT	TGAAAGTCAA	CCTACTTCGT	TAAATGAAGA	CTTGATTGAT
1501	GTGTCTATAG	ATTCCAGTAA	TCCTGTCTTT	TTAGAGGTTA	TCGATCCGAT	TACAAATTTA
1561	GGAACGCTTC	AACTTATTGA	TTTGAATACC	GGTGTTAGAC	TTAAAGAAAG	TGCTCAACAA
1621	GGTATTCAGC	GATATGGAAT	TTATGAACGT	GAAAAAGATT	TGGTTGTTAT	TAAAATAGAT
1681	TCAGGAAAAG	CTAAGCTTCA	GATACTTGAT	AAACTCGAGA	ATTTAAAAGT	GATATCAGAG
1741	TCTAATTTTG	AGATTAATAA	AAATTCATCT	CTTTATGTTG	ACTCTAGAAT	GATTTTAGTA
1801	GTTGTTAAGG	ACGATAGTAA	, TGCTTGGAGA	. TTGGCTAAAT	TITCTCCTAA	AAATTTAGAT
1861	GAATTTATTC	TGTCAGAAAA	TAAAATTTTC	CCTTTTACTA	GCTTTGCTGT	GAGAAAGAAT
1921	TTTATTTATI	TGCAAGATGA	. ACTTAAAAGC	TTAGTTACTT	TAGATGTAAA	TACTITAAAA
1981	AAAGTTAAGI	, Y				

p93 - 25015

1	ATGAAAAAA	TGTTACTAAT	CTTTAGTTTT	TITCTIATIT	TTTTGAATGG	ATTTCCTCTT
C 3	* *******	ע ע הער איר איר איר איר ע	מיוייים ממממכו	AACTATITIO	TIMMINIOUN	1011040111
401	COLE & POST BOOK	א ארביירירידיי	A JAMMAN TA	AATACGTATG	WWWWINGI	COSTATION
101	C > CALALIALIALIS C	CARCACCCC	CACCAATTCC	AATAGCAACI	CANGITATIA	IGGCWWINI
241	ጥተራጥ አጥተኮአ አጥ አ	COLLEGATION	TYCATY A AGAT	AAAAAAGCAA	GIGIIONIGI	TITITICIAIA
201	ACCACCA AAT	CACACCTTCA	CACTATATIG	AATTTAAGAA	GAATICTIAC	AGGGIATATA
261	TOTAL	TYCEATTATEA	CAGGTCTAGT	GCAGAATTAA	TIGCTAAGGT	TATTACAATA
421	THE STREET	TTTATACACC	AGATTTGGAT	TATTATAAAG	GGITTATAT	TUAGCCTGCT
401	JALLY & CALLAL	TAACTAAAGA	AAACGCAGGT	CTTTCTAGGG	TITACAGICA	GIGGGCIGGA
E 4 1	A A C A C T C A A A	J. Malaka Talaka Ta	TCTTAAAAAG	GATATTTIGT	CIGGAAATAT	TGAATCTGAC
601	TYPEATATIVE	ACACTITICGT	TACAGATAAG	GTGATAGCAG	CICITITAAG	CGAAAATGAA
ĊC1	ATTENCOCO A	DAY CAPTURE A	AGATATTACA	GATATTCAAG	GCGAAACICA	TAAGGCAGAT
721	CANCATARCA	TTCATACTCA	ATTAGACAAT	ATCCATGAAA	GCGATTCTAA	TATAACAGAA
791	ACTATTCAAA	ATTTAAGGGA	TCAGCTTGAA	AAAGCTACAG	ATGAAGAGCA	TAAAAAAGAG
0/1	NAME OF THE PARTY	ACCTTCATCC	TAAAAAGAAA	GAAAAGGAAG	AGCTAGATAA	AAAGGCAATC
001	A TATATATA A	AAGCTCAGCA	AAAATTAGAC	TCTGCTGAAG	ATAATTTAGA	TGTTCAAAGA
961	CATACTCTTA	CAGAGAAAAT	TCAAGAGGAT	ATTAATGAGA	TTAATAAGGA	AAAGAATITIG
1021	CCDAGACCTC	CTYCATYCTAAG	TTCTCCTAAA	GTTGATAAGC	AACTGCAAAT	AAAAGAGAGT
1001	CTACAACATT	TGCAGGAGCA	GCTTAAAGAA	GCTGGTGATG	AAAATCAGAA	AAGAGAAATT
11/1	CACAACCAAA	AAYTAAAYTT	AAAAAGGGAC	GAAGAACTTT	TAAAAAGTAA	AGATGGCAAA
1201	CTAACTAAAG	ATTATGAAGC	ATTAGATCTT	GATCGAGAAT	TATCCAAAGC	TICTAGTAAA
1261	CANANACTA	ACCTCAAGGA	AGAAGAAATA	ACTAAAGGTA	AATCACGGGC	AAGCTTAGGC
1321	CATTTGAATA	ATGATAAAAA	CCTTATGTTG	CCAGAAGATC	AAAAATTACC	TGAAGATAAA
1381	AAATTGGATA	GTAAATTAGA	TGGTAAAAAA	GAATTTAAAC	CAGTTTCTGA	GGTTGAAAAA
1441	TTAGATAAGA	TTTCCAAGTC	TAATAACAAT	GAGGTTGGCA	AGTTATCACC	ATTAGATAAG
1501	CCTTCTTATG	ATGATATTGA	TTCAAAAGAG	GAGGTAGATA	ATAAAGCTAT	TAATTIGCAA
1561	AAGATCGACC	CTAAAGTTAA	AGACCAAACT	ACTICITIGA	ATGAAGATTT	GGATAAAGAT
1621	TTGACTACTA	TGTCTATAGA	TTCCAGCAGT	CCTGTATITC	TAGAGGTTAT	TGATCCTATT
1681	ACAAATTTAG	GAACCCTGCA	GCTTATTGAT	TTAAATACTG	GGGTTAGGCT	TAAGGAAAGC
1741	ACTCAGCAAG	GCATTCAGCG	GTATGGAATT	TATGAACGIG	AAAAAGATTT	GGTTGTTATT
1801	AAAATGGATT	CAGGAAAGGC	TAAGCTTCAA	ATACTTAATA	. AGCTTGAAAA	TTTGAAAGTG
1861	GTATCAGAGT	CTAATTITGA	GATCAATAAA	AATTCATCTC	TTTATGTTGA	CTCTAAAATG
1921	ATTTTGGCAG	CTGTTAGAGA	TAAGGATGAT	AGCAATGCTT	' GGAGATTGGC	TAAATTTTCT
1981	CCTAAAAATT	TGGATGAGTT	TATTCTTTCA	. GAGAATAAAA	. TTTTGCCTTT	TACTAGCTTT
2041	TCTGTGAGAA	TATTTAAAA	TTATTTGCAA	GATGAGCTTA	. AAAATCTAGT	TATTTTAGAT
2101	GTAAATACTI	TAAAAAAAGT	TAAGTA		•	

K48 OSP A/PGAU OSP A FUSION

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	10			20			30			4	40				
	•		•	•		•		•	•		•		•	٠	
ATG	AAA	AAA	TAT	TTA	TTG	GGA	ATA	GGT	CTA	ATA	TTA	GCC	TTA	ATA	GCA
TAC	TTT	TTT	ATA	AAT	AAC	CCT	TAT	CCA	GAT	TAT	AAT	CGG	AAT	TAT	CGT
Met	Lys	Lys	Tyr	Leu	Leu	Gly	Ile	Gly	Leu	Ile	Leu	Ala	Leu	Ile	Ala>
50			60			•	70			80			90		
		•	•		•		•	•		•		•	•		•
TGT	AAG	CAA	TAA	GTT	AGC	AGC	CTT	GAT	GAA	AAA	TAA-	AGC	GTT	TCA	GTA .
ACA	Tree	GIT	TTA	CAA	TCG	100	GAA	CIA	CIT	TTT	TTA	TCG	CAA	AGT	CAT
Cys	Lys	GIN	ASN	vaı	Ser	ser	Leu	ASP	GIU	Lys	Asn	Ser	Val	Ser	Val>
1	00		:	110			120			1:	30			140	
	*	•		•		•.	•		•		*	•		•	•
GAT	ATT	CCT	GGT	GGA	ATG	ACA	GTT	CTT	GTA	AGT	AAA	GAA	AAA	GAC	ጹጹኡ
CTA	AAT	GGA	CCA	CCT	TAC	TGT	CAA	GAA	CAT	TCA	TTT	CTT	TIT	CTG	TŢT
Asp	Leu	Pro	Gly	Gly	Met	Thr	Val	Leu	Val	Ser	Lys	Glu	Lys	Asp	Lys>
	150			10	60			170			180			1 (90
•	•				•	•		•		•	•		•		•
GAC	GGT	AAA	TAC	AGT	CTA	GAG	GCA	ACA	GTA	GAC	AAG	CTT	GAG	CTT	AAA
CTG	CCA	TTT	ATG	TCA	GAT	CTC	CGT	TGT	CAT	CTG	TTC	GAA	CTC	GAA	TIT .
Asp	Gly	Lys	Tyr	Ser	Leu	Glu	Ala	Thr	Val	Asp	Lys	Leu	Glu	Leu	Lys>
	2	200			210			22	20		2	230			240
		•		•	•		•		•	•		•		•	•
GGA	ACT	TCT	GAT	AAA	AAC	AAC	GGT	TCT	GGA	ACA	CTT	GAA	GGT	GAA	AAA :
CLI	The	AGA	CTA	TTT	11G	TTG	CCA	AGA	CCT	TGT	GAA	CIT	CCA	CTT	TIT _
GIY	THE	Ser	ASP	rys	ASII	ASN	GIÀ	ser	GIĀ	Thr	Leu	Glu	Gly	Glu	Lys
		25	50	_	2	260		_	270			.28	30		
аст	C)C	2 2 2	A CT	222	CTA		mm s				•		•	•	
TGA	GAC CTG	444	TCA	444 444	CAT	777	TIA	TOT	WII.	GCT.	GAT	GAC	CTA	AGT	CAA
Thr	Asp	Lvs	Ser	Lvs	Val	Lvs	Leu	Thr	Tle	Ala	CIN	710	CAT	TCA So-	Glr>
	-			-, -		_, _				7.14	rsp	nap	Dea	Sei	GIES
290			300			31	0		3	320			330		
		•	•		*		•	•		•		•	•		•
		TTT		ATT	TTC	AAA	GAA	GAT	GCC	AAA	ACA	TTA	GTA	TCA	AAA
The	TTT	Pha	CIT	TAA	AAG	TTT	CIT	CTA	CGG	TTT	TGT	TAA	CAT	AGT	TTT
1111	Lys	File	GIU	11e	Pne	Lys	GIU	ASP	ATG	Lys	Thr	Leu	Val	Ser	Lys>
34	10		350				360		370		380		80		
***	CTA			*	~	•	-		•		•	•		*	
ተተተ ተ	GTA	TCC	CIT	AAA	GAC	AAG	TCA	TCA	ACA	GAA	GAA	AAA	TTC	AAC	GAA
Inc	CAT	The	Leu	111	7.LC	THE	AGT'	AGT	TGT	CIT	CTT	TIT	AAG	TTG	CII
Ly 3	Val	TILL	Leu	Lys	ASP	rys	ser	ser	IUL	GIu	Glu	Lys	Phe	Asn	Glu>

FIGURE 23 (1 of 3)

K48 OSP A/ PGAU OSPA FUSION

430 420 400 390 AAG GGT GAA ACA TCT GAA AAA ACA ATA GTA AGA GCA AAT GGA ACC AGA TTC CCA CTT TGT AGA CTT TTT TGT TAT CAT TCT CGT TTA CCT TGG TCT Lys Gly Glu Thr Ser Glu Lys Thr Ile Val Arg Ala Asn Gly Thr Arg> 460 450 CTT GAA TAC ACA GAC ATA AAA AGC GAT GGA TCC GGA AAA GCT AAA GAA GAA CTT ATG TGT CTG TAT TTT TCG CTA CCT AGG CCT TTT CGA TTT CTT : Leu Glu Tyr Thr Asp Ile Lys Ser Asp Gly Ser Gly Lys Ala Lys Glu> 510 520 500 GTT TTA AAA GAC TTT ACT CTT GAA GGA ACT CTA GCT GCT GAC GGC AAA CAA AAT TIT CTG AAA TGA GAA CTT CCT TGA GAT CGA CGA CTG CCG TIT Val Leu Lys Asp Phe Thr Leu Glu Gly Thr Leu Ala Ala Asp Gly Lys> 560 570 550 540 530 ACA ACA TTG AAA GTT ACA GAA GGC ACT GTT GTT TTA AGC AAG AAC ATT TGT TGT AAC TTT CAA TGT CTT CCG TGA CAA CAA AAT TCG TTC TTG TAA Thr Thr Leu Lys Val Thr Glu Gly Thr Val Val Leu Ser Lys Asn Ile> 620 600 610 590 TTA AAA TCC GGA GAA ATA ACA GTT GCA CTT GAT GAC TCT GAC ACT ACT AAT TTT AGG CCT CTT TAT TGT CAA CGT GAA CTA CTG AGA CTG TGA TGA Leu Lys Ser Gly Glu Ile Thr Val Ala Leu Asp Asp Ser Asp Thr Thr> 660 650 640 630 CAG GCT ACT AAA AAA ACT GGA AAA TGG GAT TCA AAA ACT TCT ACT TTA GTC CGA TGA TTT TTT TGA CCT TTT ACC CTA AGT TTT TGA AGA TGA AAT Gln Ala Thr Lys Lys Thr Gly Lys Trp Asp Ser Lys Thr Ser Thr Leu> 720. 700 690 710 680 ACA ATT AGT GTT AAC AGC AAA AAA ACT ACA CAA CTT GTG TTT ACT AAA TGT TAA TCA CAA TTG TCG TTT TTT TGA TGT GTT GAA CAC AAA TGA TTT Thr Ile Ser Val Asn Ser Lys Lys Thr Thr Gln Leu Val Phe Thr Lys> 730 740 750 760 CAA TAC ACA ATA ACT GTA AAA CAA TAC GAC TCC GCA GGT ACC AAT TTA GTT ATG TGT TAT TGA CAT TTT GTT ATG CTG AGG CGT CCA TGG TTA AAT Gln Tyr Thr Ile Thr Val Lys Gln Tyr Asp Ser Ala Gly Thr Asn Leu>

FIGURE 23 (2 of 3)

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K48 OSPA / PGAU OSP A FUSION

770 780 790 800 810

GAA GGC ACA GCA GTC GAA ATT AAA ACA CTT GAT GAA CTT AAA AAC GT
CTT CCG TGT CGT CAG CTT TAA TTT TGT GAA CTA CTT GAA TTT TTG CGA
Glu Gly Thr Ala Val Glu Ile Lys Thr Leu Asp Glu Leu Lys Asr. Ada>

820 TTA AAA TAA AAT TTT ATT Leu Lys ***>

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FIGURE 23 (3 of 3)

B-31/OP A PGAU OSP A FUSION

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	Met	Lys	Lys	Tyr	Leu	Leu	Gly	Ile	Gly	Leu	Ile	Leu	Ala	Leu	Ile	Ala>
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	Asp	Gly	Lys	Tyr	Ser	Leu	Lys	Ala	Thr	Val	Asp	Lys	lie	Glu	Leu	Lys>
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	CCT	TGA	AGA	CTA	TTT	CTG	TTA	CCA	AGA	CCT	CAC	GAA	CII	CCA	TGT	TTT
	Gly	Thr	Ser	Asp	Lys	Asp	Asn	Gly	Ser	Gly	Val	Leu	Glu	Gly	Thr	Lys>
			_				260			270				80		
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A. CLASSIFICATION OF SUBJECT MATTER IPC 6 C12N15/11 C07K14/20

G01N33/569

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According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

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Further documents are listed in the continuation of box C.	Patent family members are listed in annex.
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which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art. "&" document member of the same patent family
Date of the actual completion of the international search 10 March 1999	Date of mailing of the international search report $26/03/1999$
Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Authorized officer Ceder, 0

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